1 Chemical and Mechanical Properties of Palm Hearts from three Iranian

2 Date Palm Cultivars Affect Physiological Responses of Red Palm Weevil

3 (*Rhynchophorus ferrugineus* Olivier, 1790) (Coleoptera: Dryophthoridae)

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5 ABSTRACT

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The red palm weevil, Rhynchophorus ferrugineus (Olivier, 1790) (Coleoptera: Dryophthoridae) 6 is a serious insect pest of date palm trees worldwide. In this study, we aimed to assess the energy 7 8 reserves and antioxidative defense response of R. ferrugineus larvae on palm hearts of three date 9 palm cultivars (Piarom, Mazafati, and Kalute). Furthermore, the specific biochemical and mechanical properties of the palm hearts were evaluated. The insect was reared in a growth 10 chamber at $29 \pm 2^{\circ}$ C, $80 \pm 5\%$ RH, and a photoperiod of 12:12 (L:D) and the experiments were 11 conducted using the seventh instar larvae. The results revealed that the highest contents of sugar 12 and lipid in R. ferrugineus larvae were on Piarom and Mazafati palm hearts, respectively. The 13 antioxidant enzymes activities in R. ferrugineus larvae were significantly different on the tested 14 palm hearts. According to the mechanical analysis, the Mazafati palm heart had the lowest tissue 15 hardness. Biochemical analysis revealed that the total sugar content was high in Mazafati and 16 Kalute palm hearts, and low in Piarom palm heart. The concentrations of phenol and flavonoid 17 were also different in the tested palm hearts. Moreover, gas chromatography-mass spectrometry 18 19 (GC-MS) profiling showed that alkaloids, terpenes, acids, and amides were the major volatile 20 components of the tested palm hearts. Based on the results, Piarom showed traits associated with 21 reduced larval performance, suggesting its potential use in integrated pest management (IPM) 22 programs.

Keywords: Antioxidant enzymes; mechanical traits; phytochemicals; red palm weevil; energy
 reserves.

25 INTRODUCTION

Palm heart, often called heart-of-palm or palmito, is the palm tree stem cells, located near the apical meristem of tree (Mostafa, 2024). It is conventionally harvested from the inner core of date

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palm trees, Phoenix dactylifera L. (Arecales: Arecaceae), or gathered from a cluster of shoots 28 29 developing from the main branch. This edible white cylindrical substance can be eaten raw or 30 cooked in salads and various recipes (Quast and Bernhardt, 1978; Bovi et al., 2001). Palm heart with its tender texture and delicate flavor is a good source of carbohydrates, lipid, protein, 31 32 vitamins, minerals, and dietary fiber not only for humans, but also for pests (Quast and Bernhardt, 33 1978). Various species of palm trees, particularly the date palm, are globally under severe threats by the attack of Rhynchophorus ferrugineus (Olivier, 1790) (Coleoptera: Dryophthoridae) or 34 commonly known red palm weevil (RPW), which is considered as the most destructive pest of 35 36 palm trees (Kubar et al., 2017; Wang et al., 2021).

The pest usually attacks young palms (generally less than 20 years old) because of their soft and succulent tissues. Although adults may partially feed on stem, leaves, and flowers, but the most damaging life stage of the pest is larvae. The female of RPW uses its rostrum to drill a hole and then, lay eggs in it. While feeding on the internal tissues, ideally the palm heart, the larvae burrow into the trunks of palm trees and create sizable tunnels within it (Kubar *et al.*, 2017; Manzoor *et al.*, 2022), leading to the destruction of the palm trunk's vascular system.

43 A part of sustainable integrated management (IPM) programs to control R. ferrugineus outbreak 44 is using resistant palm tree cultivars, an eco-friendly and long-lasting tool, which are extensively used in numerous countries of the world (Faleiro et al., 2014; Kubar et al., 2017). More than 3000 45 46 cultivars of date palm are known to exist all around the globe (Alrashidi et al., 2023). These 47 cultivars have various morphological, chemical, and nutritional properties (Trabzuni et al., 2014), 48 which their susceptibility or resistance to insect pests, particularly RPW is often linked to these 49 traits (Manzoor et al., 2022). Faleiro et al. (2014) reported that among seven different date palm 50 cultivars, Gaar, Khasab, and Shahal cultivars were poorly prone to attack by R. ferrugineus. 51 According to Manzoor et al. (2022) assessment of R. ferrugineus's feeding preferences across 11 52 different date palm cultivars, the pest heavily infested the Hillawi cultivar. Insect larvae may 53 choose the palm heart as a feeding location due to its soft and delicate tissues and nutritional value 54 (Trabzuni et al., 2014). Nearly, all plants usually use complicated strategies to combat pests (Al-55 Khayri et al., 2023). A phytochemicals variety, i.e. secondary metabolites are involved in their 56 defense against insect pests. These phytochemicals may have repellent or deterrent effects on 57 phytophagous insects, interfering with their physiological and nutritional performances (Bennett 58 and Wallsgrove, 1994). During these interactions, the ingestion of the toxic phytochemicals by

insects may induce toxic reactive oxygen species (ROS), which the associated oxidative stress in insect's cells targets biomolecules, leading to the damage of lipids, proteins, and DNA (Mythri *et al.*, 2013). Investigating the biochemical profile of three Saudi date palm cultivars, Trabzuni *et al.* (2014) reported that their palm hearts were rich in antioxidants. Sahito *et al.* (2017) noted that the vulnerability of Aseel variety to *R. ferrugineus* was due to its soft trunk fiber and high nutritional value. Other researchers showed that plant defense metabolites increased oxidative stress in plant feeding insects (Kaur *et al.*, 2017).

Considering the significance of adopting resistant cultivars for IPM programs, the current 66 research was carried out to examine how the nutritional profile and antioxidant enzyme activities 67 of *R. ferrugineus* larvae may alter while feeding on the palm hearts of various date palm cultivars. 68 69 Furthermore, we wanted to know whether the mechanical and biochemical properties of palm hearts were different in the examined date palm cultivars. The obtained results may aid to identify 70 the physiological response of *R. ferrugineus* larvae to cope with palm heart phytochemicals, which 71 72 could be used in the development of sustainable control approaches against the pest in date palm 73 orchards.

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75 MATERIALS AND METHODS

76 Insect rearing

Rhynchophorus ferrugineus was reared in a growth chamber at $29 \pm 2^{\circ}$ C, $80 \pm 5^{\circ}$ RH, and a 77 78 12:12 (L:D) photoperiod at the Agricultural Research Center of Jiroft, Kerman province, Iran. 79 Adults were collected from date palm orchards in Jiroft County (570637°N, 3152288°E) and 80 placed in plastic boxes containing a cotton ball saturated with 10 % (w/v) bee pollen solution. Eggs were collected and incubated in Petri dishes (8 cm diameter, 2.5 cm height), lined with moist filter 81 papers. Then newly hatched larvae (30 larvae per each cultivar) were maintained in plastic 82 container (10 cm diameter, 20 cm height) containing fresh grated-palm heart (100 g) of each date 83 84 palm cultivar as substrate. The food was provided daily. The palm hearts were grated to facilitate the first instar larvae feeding. After reaching to the second instar, the larvae were individually 85 86 transferred on the newly cut young shoots (one larva per one shoot) (each having a palm heart 87 surrounded with young leaves) from each date palm cultivar. At the seventh instar, as the most damaging larval instar, they were used in experiments for evaluating their antioxidant enzymes 88 89 activities and energy resources.

90 **Preparation of palm heart samples**

91 Palm hearts of three commercially important date palm tree cultivars in Iran, including Piarom, 92 Mazafati, and Kalute were randomly obtained from date palm orchards in Karimabad-e Sofla (28° 93 29' 07" N, 57° 42' 58" E) (Jiroft County, Kerman Province, Iran). The selected cultivars are among the most widely grown date palm cultivars in date-producing areas in Iran (Yektankhodaee et al., 94 95 2010) as well as the most attacked ones by RPW based on our observation in Kerman Province. 96 They were harvested from the same-aged date palm trees (each cultivar) then, carefully peeled and 97 freed from other plant materials. The fresh palm hearts of each studied cultivar were used for 98 further mechanical analysis. For biochemical analysis of fresh palm hearts; they were chopped into 99 small pieces (100-200 g) and dried in shade, in a ventilated room at room temperature. The dried 100 samples were completely powdered using an electric grinder and stored (4°C) in sealed containers 101 for further use.

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103 Determination of energy reserves of *R. ferrugineus* larvae

104 Sugar content

105 The samples were prepared as described by Foray et al. (2012). The larvae (3 larvae) were 106 homogenized using a glass homogenizer. Homogeneity was centrifuged at 5000 g for 15 minutes at 2°C in a refrigerated centrifuge. After centrifugation, the precipitate (containing sugars) was 107 108 collected and oven-dried at 90-110°C. The obtained condensed sample (precipitate) was reacted 109 with anthrone reagent. The reagent was prepared by dissolving 75 mg anthrone in 150 mL distilled 110 water and then, mixed with 380 mL H₂SO₄. After increasing the volume of the mixture to 5 mL. 111 it was added to the condensed samples. The mixture was incubated for 17 min at 90-110°C. The amount of sugar was recorded at 625 nm by spectrophotometer (S2100SUV, UNICO, USA). 112 Estimation of sugar content was carried out in three replicates. 113

Lipid content

The lipid content of *R. ferrugineus* larvae was quantified following the method described by Foray *et al.* (2012). First, the supernatant obtained by centrifugation of samples (3 larvae); was collected and diluted with distilled water (2 mL). After that, the mixture was re-centrifuged at 3000 *g* for 1 min. The precipitate thus obtained was oven-dried at 90-110°C. Then, H₂SO₄ (98 %, 0.2 mL) was added to the lipid sample and incubated at 90-110°C for 10 min. Finally, vanillin reagent

121 (60 mg of vanillin solution dissolved in 10 mL hot water and kept in dark) was added to the 122 mixture. The absorbance was read at 530 nm (S2100SUV, UNICO, USA). Measurements were 123 technically done in three replicates.

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125 **Protein content**

Protein concentration of RPW larvae was assayed in three replicates, according to Greenfield (2018) method. After homogenization of samples (3 larvae) with a glass homogenizer and following their centrifugation, the Bradford reagent (190 μ L) was added to the supernatant (10 μ L). The absorbance was recorded at 630 nm (S2100SUV, UNICO, USA).

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131 Determination of antioxidant activity of *R. ferrugineus* larvae

132 Extraction of enzymes

For preparation of enzymes extract from each cultivar, 10 larvae previously fed on the related date palm cultivars were randomly collected. Then, the larvae were homogenized in phosphate buffer (10 mM, pH 7.5, 200 μ L) using a homogenizer. The homogenates obtained for each treatment were centrifuged at 13000 g at the temperature of 4°C for 15 min. For enzymatic evaluates, the supernatants were collected in micro tubes and stored at -20°C. All enzyme assays were done in three replications.

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140 Catalase activity

141 Catalase activity of *R. ferrugineus* larvae was measured following the methodology of Aebi 142 (1984). For this purpose, 50 μ L of crude enzyme extract, 225 μ L H₂O₂ 225mM, and 70 mM 143 phosphate buffer 225 μ L (pH 7.5) were mixed and the absorbance was read at 240 nm by 144 spectrophotometer.

Esterase activity

147 The activity of this enzyme was calculated according to van Asperen (1983). Briefly, enzyme 148 extract (15 μ L), α -naphthyl acetate (10 mM in acetone, 10 μ L), and phosphate buffer (70 mM, pH 149 7.4, 40 mL) were incubated for 10 min at 30°C. Next, 50 μ L Fast Blue RR salt (50 mg in sodium 150 phosphate buffer (40 mM, pH 7, 50 mL)) was added to the reaction mixture. The absorbance was 151 recorded at 450 nm using a spectrophotometer.

153 **Peroxidase activity**

154 This reaction was performed by Bergmeyer methodology (1974). For this reason, enzyme extract 155 (50 μ L), H₂O₂ (225 mM, 225 μ L), guaiacol buffer (45 mM, pH 7, 225 μ L), and potassium 156 phosphate buffer (50 mM, pH 7, 50 μ L) were used as the assay mixture. Change in absorbance 157 was measured at 470 nm by spectrophotometer.

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159 Determination of palm hearts mechanical properties

160 The mechanical features (force, breaking deformation, stress, modulus of elasticity, and energy) of fresh palm hearts were determined using a Universal testing machine (model MRT-5, Santam, 161 162 Iran) (Fig. 1). First, 500 g fresh palm hearts were placed upon a flat plate and the hardness was 163 measured with a 4.8-mm-diameter stainless steel puncture. Three palm hearts were experimented 164 with 10 mm penetration depth for each cultivar. The maximum force required to penetrate each 165 palm heart was calculated according to Ghanbarzadeh (2019) and Najafi et al. (2022). The highest value recorded by the puncture while passing through each tested palm heart, in Newton (N), was 166 167 considered as hardness of the palm heart. The elasticity was calculated according to the force-time 168 curve, using the same probe. The force-deformation curves were used to determine the mechanical 169 features.

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171 Determination of palm hearts total sugar

To measure the total sugar content, 40 mg the powder of each palm heart cultivar (taken from a 172 173 fresh sample as described previously) was homogenized in ethanol (80 %, 5 mL) to remove sugars. 174 Residue was precipitated by placing the mixture in a water-bath set at 70°C for 10 min. The residue 175 was washed repeatedly with ethanol (80%) until all the soluble sugars were completely extracted. The sample extract was mixed with 3 mL anthrone reagent (150 mg anthrone dissolved in H₂SO₄ 176 177 (13 M, 100 mL). Afterwards, the mixture was put in a water-bath set at 100 °C (20 min) then, 178 cooled to room temperature. Absorbance was measured in triplicate for each treatment at 630 nm 179 (McCready et al., 1950).

Determination of palm hearts phytochemicals

182 **Preparation of extracts**

Each palm heart powder (1 g) was extracted with methanol (20 mL) for a period of 24 h in dark situation. Then, each sample was centrifuged at 1000 g for 5 min.

Total phenol content

The amount of total phenolic content of the examined cultivars of palm hearts was determined using the Folin-Ciocalteu reagent, based on the method defined by Soland and Lima (1999). Briefly, each palm heart extract (0.2 mL) was mixed with Folin-Ciocalteu reagent (50 %, 0.4 mL) and sodium carbonate (Na₂CO₃) solution (5 %, 0.8 mL). The reaction mixture was allowed to develop for 2 h in dark and the absorbance was measured at 765 nm using a spectrophotometer. Gallic acid was used as standard. The assays were carried out in three replicates for each palm cultivar.

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194 **Total flavonoid content**

195 The total flavonoid content in the palm hearts was determined according to the procedure of Jia 196 *et al.* (2015). First, each palm heart extract (500 μ L) was mixed with bicarbonate (5%, 75 μ L). 197 After 5 min, AlCl₃ (10 %, 150 mL) was added to the mixture and let to stand for another 5 min at 198 room temperature. Then, NaOH (1.0 M, 500 ml) and distilled water (275 μ L) were added. The 199 absorbance was read at the wavelength of 510 nm. Each tested extract included three replicates. 200 The total flavonoid of the treatments was expressed as μ g of quercetin equivalent per gram dry 201 weight (DW).

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203 Gas chromatography-mass spectrometry (GC-MS) analysis

204 The palm heart's powder of each date palm cultivar (20 g) was suspended in aqueous methanolic 205 solution (1: 2/v:v) (200 mL) in a container and then, placed on a rotary shaker for 48 h. After that, 206 the mixture filtered and poured into falcon tubes, then centrifuged at 6000 g for 3-5 min. The 207 resulting extracts were subjected to GC-MS (Agilent Technologies 7890 A, Germany) to evaluate 208 the phytochemical profile of the treatments. The analysis was performed on a HP-5-MS capillary 209 column (30 m \times 250 µm i.d., film thickness 0.25 µm, temperature program of 50-280°C). The 210 injector temperature was 50°C raised to 260°C with normal injection mode and the flow rate of 211 carrier gas (Helium) was 1.0 mL/min. All MS was operated at EI mode (70 eV ionization energy, 212 250°C EI ion source temperature). The results were compared by using WILEY, Demo and NIST 213 Spectral libraries search programs.

215 Data analysis

All data were analyzed by one-way analysis of variance (ANOVA) appropriate to completely randomized design using the statistical software of SPSS ver. 26.0 (Stata Corp., College Station, TX, USA). The means separation was performed by a Tukey's test at P < 0.05. The data were examined for normality using Kolmogorov–Smirnov test before analysis. The graphs were created in SigmaPlot.

221

222 **RESULTS**

223 Energy reserves of *R. ferrugineus* larvae

The concentrations of storage macromolecules in *R. ferrugineus* larvae fed on palm hearts from different date palm cultivars are shown in Fig. 2. Larvae fed on Piarom palm heart had the highest sugar content (0.484 mg/g) ($F_{2,9} = 11.96$; P = 0.003). Furthermore, lipid content was highest when larvae fed on Mazafati palm heart (16.2 mg/g), while it was lowest when the larvae reared on Piarom palm heart (8.8 mg/g) ($F_{2,9} = 553.62$; P < 0.0001).

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Antioxidant activity of R. ferrugineus larvae

The antioxidant activities of *R. ferrugineus* larvae were significantly different on the tested cultivar, except that of esterase ($F_{2,9} = 2.51$; P = 0.136) (Fig. 3 B). The catalase activity of larvae was highest on Kalute palm heart (0.290 U/mg) and lowest on Mazafati palm heart (0.202 U/mg) ($F_{2,9} = 5.71$; P = 0.025) (Fig. 3 A). The highest and lowest peroxidase activities were observed on Mazafati (0.154 U/mg) and Piarom (0.128 U/mg) palm hearts, respectively ($F_{2,9} = 6.17$; P = 0.021) (Fig. 3 C).

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Palm hearts mechanical properties

There were significant differences among the cultivars regarding the mechanical properties of their palm hearts. The maximum force required to puncture the palm hearts was recorded in Kalute palm heart, while the minimum force was obtained in Mazafati palm heart ($F_{2,6}$ =62.37; P <0.0001) (Table 1). The highest value of breaking deformation was in Mazafati palm heart ($F_{2,6}$ = 14.41; P < 0.0001) (Table 1). The stress was highest in Kalute palm heart, followed by Piarom and Mazafati palm hearts ($F_{2,6}$ = 11.74; P < 0.0001) (Table 1). The highest and lowest modulus of elasticity were in Kalute and Mazafati palm hearts, respectively ($F_{2,6}$ = 3448.34; P < 0.0001) (Table

1). The highest energy was consumed for puncturing Kalute palm heart. However, the lowest energy was used up in Piarom palm heart ($F_{2,6}$ = 1936.56; P < 0.0001) (Table 1).

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249 Palm hearts total sugar content

According to the results, there was a significant variation in total sugar contents of palm hearts ($F_{2,9} = 50.35$; P < 0.0001). The total sugar content was higher in Mazafati (0.679 mg/g DW) and Kalute (0.588 mg/g DW) palm hearts than in Piarom (0.345 mg/g DW) (Fig. 4).

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254 Palm hearts phytochemicals

The results indicated that the total phenol and flavonoid contents of palm hearts were significantly different ($F_{2,12}$ = 58.86; P < 0.0001, $F_{2,12}$ = 27.35; P < 0.0001, respectively). The higher amounts of phenol were detected in Piarom (34.6 µg/g DW) and Mazafati (34.3 µg/g DW) palm hearts. However, its lowest amount was found in Kalute palm heart (30.1 µg/g DW) (Fig. 5). The flavonoid content was highest in Mazafati palm heart (50.2 µg/g DW), whereas it was lower in Piarom (31.2 µg/g DW) and Kalute (28.2 µg/g DW) palm hearts (Fig. 5).

261 According to GC-MS analysis, different compounds with various antibiotic activities (i.e., 262 antimicrobial, insecticidal, larvicidal, and fungicide activities) were identified in the extracts of 263 palm hearts of Piarom, Mazafati, and Kalute, representing 57.70 %, 89.83 %, and 49.48 % of the 264 total sample, respectively. Two alkaloids including Dimpylate and Bis(2-ethylhexyl) phthalate 265 were the main constituents in Piarom palm heart (Table 2). The predominant constituents in Mazafati palm heart were classified as terpenes (9-octyl-heptadecane and Eicosane) and alkaloids 266 267 (Bis(2-ethylhexyl) phthalate) (Table 2). Furthermore, the major compounds in Kalute palm heart 268 were acids (Acetic acid, chloro-, 2-butoxyethyl ester and Butanoic acid, 2-oxo-) and amids (N-269 methoxymethyl-N-methylacetamide) (Table 2).

DISCUSSION

The consumption of *R. ferrugineus* larvae on palm hearts of various date palm cultivars greatly affected the energy reserves of the larvae. The larvae fed on Piarom palm heart had high level of sugar. This value was about 1.3-fold higher than those fed on Mazafati and Kalute palm hearts. Interestingly, the total sugar content of palm hearts either in Mazafati or in Kalute was higher than that in Piarom, emphasizing that the Mazafati and Kalute palm hearts were high-nutrient foods for the pest. Similarly, Farazmand (2002) reported that Mazafati was nutritionally suitable food for *R*.

278 ferrugineus larvae. For long-term use, the majority of ingested carbohydrates are usually stored as 279 glycogen or triglycerides in fat bodies to meet the insects' energy requirements for reproduction 280 and metamorphosis (Enriquez and Visser, 2023) as well as to assist them withstand harsh 281 environmental conditions (Sinclair, 2015). It seems that feeding of R. ferrugineus larvae on sugar-282 rich diets, i.e. Mazafati and Kalute, can impose low metabolic and fitness costs to the pest, thus it 283 readily converts the received dietary sugars to lipids. However, the high sugar level in larvae body 284 on Piarom palm heart indicates that it was used mainly for instant energy needs, such as energy 285 production, growth and development, immune or stress response, etc., but not or less for future use 286 (in the forms of glycogen or lipids). Based on the results, the lipid level of RPW larvae was highest 287 when fed on Mazafati palm heart, while it was lowest feeding on Piarom palm heart, being almost 1.8-fold less than the related value obtained on Mazafati palm heart. As a result, the significant 288 289 quantity of lipid discovered in larvae feeding on palm heart of Mazafati may support the concept 290 that the insect transforms dietary carbohydrates to lipids to endure adverse circumstances in later 291 stages of development.

292 The Mazafati suitability for R. ferrugineus larvae was supported by the of mechanical properties 293 effects of its palm heart. According to the results, the lowest amounts of force, mechanical stress, 294 and modulus of elasticity were recorded for Mazafati palm heart, emphasizing that it had low tissue 295 hardness, which can be favored by *R. ferrugineus* larvae. However, the palm heart of Kalute had 296 a high puncture resistance, as shown by its increased force, modulus of elasticity, mechanical 297 stress, and puncturing energy. This may decrease the feeding of RPW larvae by supplying 298 comparatively hard tissues. Even though, the sugar content of Kalute was suitable for R. 299 ferrugineus larvae, but the stiffness of its palm heart tissue may impose stress to the pest, probably 300 leading to increased catalase activity of larvae. High sugar intake may increase the insect's metabolism and catalase activity due to production of ROS (Kauldhar and Sooch, 2016). 301 302 Therefore, reduced fitness and host acceptance, as well as low feeding performance in insects may 303 be associated with the mechanical properties of plant tissue (Hochuli, 1996, Stevanato et al., 2020; 304 Nalam *et al.*, 2021).

The increased activities of catalase and even esterase in *R. ferrugineus* larvae feeding on Kalute palm hearts and peroxidase in larvae feeding on Mazafati and Kalute palm hearts indicate the occurrence of oxidative stress in RPW larvae body. Mohamed *et al.* (2022) reported that the upregulation of antioxidant enzymes provided protection to *R. ferrugineus* larvae from the cytotoxic

309 effects of ROS, which is consistent with our findings. Antioxidant enzymes generally reduce the 310 level of ROS (Dmochowska-Ślęzak et al., 2015 Wei et al., 2020). The ingestion of plant secondary 311 substances is reported to generate ROS in plant-feeding insects and gives rise to oxidative radicals (Umar et al., 2022, Krishnan and Kodrik, 2006). Piarom had the highest amount of phenol and 312 Mazafati had the highest amounts of phenol and flavonoid. High phenolic content in a plant 313 314 (Piarom in our study) is usually associated with its resistance, as it may interfere with the digestion or physiology of the pest (Sattari Nasab et al., 2018). However, it is worth to note that just because 315 316 the levels of phenol and flavonoid were high in Mazafati palm heart does not mean that this cultivar 317 was resistant to the pest. Since, the strong antioxidant activity in larvae, especially peroxidase might have contributed in detoxification of these compounds. The phenol concentration levels in 318 319 this investigation are lower than, those reported by Trabzuni et al. (2014) for date palm heart 320 extracts from the Solleg and Naboat Saif cultivars. The amount of defensive metabolites in various 321 date palm cultivars may be influenced by genetic and age variability, farming management 322 variations, and environmental factors (Umar et al., 2022, Al-Farsi et al., 2005). According to the 323 results, the antioxidant enzymes activity of RPW larvae was also high in Kalute palm heart. 324 Surprisingly, the amount of either phenol or flavonoid was lowest in palm heart of this cultivar, 325 emphasizing that phytochemicals other than phenol and flavonoid might have increased oxidative 326 radicals in larvae body. GC-MS analysis indicates that the main component in Kalute palm heart 327 was acidic molecules with insecticidal action, such as Butanoic acid, 2-oxo-, which may disrupt 328 insect pests' normal physiological processes, causing paralysis and death (Robert et al., 2015). 329 Based on the results, alkaloids (Bis (2-ethylhexyl) phthalate) and terpenes (9-octyl-heptadecane, 330 and Eicosane) were the predominant compounds in Mazafati palm heart. Waris et al. (2018) 331 reported that Eicosane in volatiles of rice plants elicited behavioral responses in Nilaparvata 332 lugens Stål by simply attracting the pest to the plants. However, exposure to compounds like Di 333 (2-ethylhexyl) phthalate can induce antioxidative stress in insect pests (Rivas et al., 2023), 334 emphasizing that alkaloids can negatively affect insect pests. The amounts of phenol and alkaloids 335 (Dimpylate and Bis (2-ethylhexyl) phthalate) were high in Piarom palm heart, but the larvae on 336 this cultivar had the lowest activity of peroxidase. It is possible that, some allelochemicals will 337 inhibit the activities of antioxidant enzymes and increase the level of free radicals as well as 338 oxidative stress in some species (Krishnan and Kodrik, 2006, Mythri et al., 2013, Yang et al., 339 2019). Moreover, the larvae may use antioxidant enzymes other than peroxidase to cope with stress

caused by secondary metabolites (Wei *et al.*, 2020). The number of experiments to address the
ambiguities regarding the behavior of RPW is very low, and more experiments are needed to
address the dark spots in the behavior of this pest.

In conclusion, RPW larvae were able to feed on palm hearts of the studied date palm cultivars. 343 However, not all date palm cultivars appear to be equally appropriate as hosts for this pest. The 344 Piarom palm heart was relatively unsuitable host for RPW larvae in terms of low sugar content 345 and high phytochemicals contents (especially phenol). Some mechanical traits and phytochemicals 346 of this palm heart increased stress in larvae. However, the antioxidant enzymes of larvae via 347 348 adaptive responses may moderate the suppressive effect of date palm trees' defenses against R. ferrugineus. Using the date palm-derived resistance traits, i.e., phytochemicals and mechanical 349 350 defense, offers an effective approach for strengthening IPM strategies against RPW, particularly in combination with other control measures. To control R. ferrugineus, collecting further 351 352 applicable data is suggested to assess the demographic parameters and other physiological 353 responses of this pest.

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Table 1. Mechanical properties (mean \pm SE) of palm hearts of three date palm cultivars.						
Cultivar Force (N)		Breaking deformation (mm)	Stress (MPa)	Modulus of elasticity (MPa)	Energy (mJ)	
Piarom	47.16± 0.31ab	$3.24 \pm 0.98b$	4.38±0.97ab	81.99± 1.12b	$90.97 \pm 2.22b$	
Mazafati	$35.13 \pm 1.01b$	$9.87 \pm 0.59a$	$2.88 \pm 0.57 b$	44.62±0.55c	158.51±1.39ab	
Kalute	55.66± 1.99a	$3.50 \pm 1.18 b$	5.5±0.29a	179.86±1.62a	$226.21 \pm 0.57a$	

489 Means followed by different letters in each column are significantly different (Tukey test, P<0.01).

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491	Table 2. Phytochemicals	identified in palm hear	rts of three date palm cultiv	vars using gas
	2	1	1	00

492 chromatography-mass spectrometry.

Cultivars	IUPAC Name (Synonym)	Retention time (min)	Activity	Area (%)	Category
	Dimpylate*	23.82	Antimicrobial activity	30.21	Alkaloid
Piarom	Bis(2-ethylhexyl) phthalate	35.21	Antimicrobial activity Larvicidal activity	27.49	Alkaloid
	9-octyl-heptadecane	30.15	Plant growth promotion activity Insecticidal activity	21.12	Terpene
Mazafati	Bis(2-ethylhexyl) phthalate	32.89	Antimicrobial activity Larvicidal activity	50.89	Alkaloid
	Eicosane	39.89	Plant growth promotion activity Insecticidal activity	17.82	Terpene
	Acetic acid, chloro-, 2-butoxyethyl ester	13.98	Insecticidal activity	19.46	Acid
Kalute	Butanoic acid, 2-oxo-	13.16	Insecticidal activity Antimicrobial activity	22.79	Acid
	N-Methoxymethyl-N- methylacetamide	15.20	Fungicide activity Insecticidal activity Herbicidal activity Antimicrobial activity	7.23	Amid

493 Only major compounds were listed.



495

496 **Figure 1.** The Universal testing machine for determining the mechanical properties of palm hearts.



499 **Figure 2.** Energy reserves (mean \pm SE) in *Rhynchophorus ferrugineus* larvae feeding on palm 500 hearts of three date palm cultivars. Columns with different letters represent significant differences 501 among cultivars (Tukey test, P<0.01).

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508 Figure 3. A, B, C: Antioxidant enzymes activities (mean ± SE) in *Rhynchophorus ferrugineus*

- 509 larvae feeding on palm hearts of three date palm cultivars. Columns with different letters represent
- 510 significant differences among cultivars (Tukey test, P<0.01).



- 511 512 Figure 4. Total sugar content (mean \pm SE) in palm hearts tissues of three date palm cultivars.
- 513 Columns with different letters represent significant differences among cultivars (Tukey test,
- 514 P<0.01). *DW: Dry Weight.



Cultivar Figure 5. Mean (± SE) amounts of phytochemicals (phenol and flavonoid) in palm hearts tissues of three date palm cultivars. Columns with different letters represent significant differences among cultivars (Tukey test, P<0.01). *DW: Dry Weight.

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بررسی ویژگیهای شیمیایی و مکانیکی بنیر سه رقم خرمای ایرانی و تأثیر آن بر یاسخهای فیزیولوژیکی 526 سرخرطومي حنايي خرما، (Coleoptera: سرخرطومي حنايي خرما، (Rhvnchophorus ferrugineus Olivier, 1790) 527 **Dryophthoridae**) 528 فاطمه خدادادی اسفیچار، مریم پهلوان یلی، و پیمان نامور 529 530 چکیدہ سرخرطومي حنايي خرما، (Coleoptera: Dryophthoridae) سرخرطومي حنايي خرما، (Rhvnchophorus ferrugineus (Olivier, 1790) 531 آفت جدی در ختان خرما خرما در سر اسر جهان است. هدف از این مطالعه، ار زیابی ذخایر انر ژی و و اکنش آنتی اکسیدانی 532 لاروهای R. ferrugineus روی پنیر خرمای سه رقم خرما (پیارم، مضافتی و کلوته) بود. علاوه براین ویژگیهای 533 بيوشيميايي و مكانيكي پنير خرما هم مورد بررسي قرار گرفت. پرورش حشر ، در شرايط آز مايشگاهي با دماي 2 ± 29 در جه 534 سانتیگراد و رطوبت نسبی 5 ±80 درصد و دوره نوری 12:12 (روشنایی: تاریکی) روی لاروهای سن هفتم انجام شد. نتایج 535 نشان داد که بیشترین ذخایر انرژی (قند و چربی) لارو بهترتیب روی پنیرهای خرمای پیارم ومضافتی بود. فعالیت آنزیمهای 536 آنتی اکسیدانی در لار و های R. ferrugineus روی بنیر های خر مای آز مایش شده به طور قابل توجهی متفاوت بود. طبق آنالیز 537 مكانيكي، ينير هاي خرماي رقم مضّافتي كمترين سُختي بافت را داشت آناليز هاي بيوشيميايي ينير هاي خرماي مورد مطالعه 538 نشان داد که محتوی قند کل در مضافتی و کلوته بالا و در پیارم پایین بود. همچنین، سطح فنل کل به طور معنیداری در 539 ينير های خرمای آزمايشی متفاوت بود. نتايج کروماتوگرافی گازی-طيف سنج جرمی (GC-MS) نشان داد که آلکالوئيدها، 540 تُرین ها، اسیدها و آمیدها مهمترین ترکیبات فرار پنیر های خرمای مورد بررسی بودند. بر اساس نتایج، پیارم ویژگی هایی 541 مر تبط با کاهش عملکر د لار و ی ر ا نشان داد که نشاندهنده کار بر د بالقوه آن در بر نامههای مدیریت تلفیقی آفات (IPM) است. 542 543 544

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