

Interactive Effects of Nitrogen Form and Oxygen Concentration on Growth and Nutritional Status of Eggplant in Hydroponics

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ABSTRACT

A greenhouse study was carried out to determine the effect of nitrogen forms and different O₂ levels on growth and mineral nutrient concentrations of eggplant. The experimental design was a completely randomized factorial experiment with two factors, namely: (i) Two nitrogen forms (Ca(NO₃)₂ and (NH₄)₂SO₄) and (ii) Three O₂ levels of the nutrient solutions (1±0.3, 2±0.3, 3±0.3, and 4±0.3 mg L⁻¹ O₂). The results showed that ammonium application reduced all measured parameters of vegetative growth, whereas high oxygen levels increased the vegetative growth. Comparing with nitrate-N, ammonium application increased the concentrations of NPK and Zn in leaves and Zn and Cu in roots, while it decreased the concentration of Mg, Ca, Cu, Mn, and Na in leaves and Ca, Mg, Mn, and Na in roots. High levels of O₂ increased N, Mg, Ca, Cu, and Mn content of leaves, as well as Mn and Na content in roots, while it decreased the concentration of K in leaves and P and Zn in roots. According to the results, the increase in O₂ amount of the nutrient solutions partly alleviated ammonium toxicity in eggplant. Therefore, in floating hydroponic cultures, O₂ level and its distribution should be controlled and must not be lower than 4 mg L⁻¹.

Keywords: Ammonium, Nitrate-N, Nutrient solution aeration, Soilless culture, *Solanum melongena*.

INTRODUCTION

Nitrogen is a constituent of many plant cell components, including amino acids and nucleic acids. Therefore, nitrogen deficiency restricts the growth of the vegetative organs (Barker and Breyson, 2006). It is available in various inorganic forms such as nitrate, ammonium, and nitrogen molecules, and organic forms such as urea and amino acids, which can be changed to forms available for absorption by the plant (Marschner, 1995). Ammonium (NH₄⁺) and nitrate (NO₃⁻) are

used as nitrogen source for the plant, although the plant response to a specific form of nitrogen varies from species to species (Britto and Krounzucker, 2002). At high concentrations, NH₄⁺ toxicity can often be occurred for different species (Britto and Krounzucker, 2002; Roosta *et al.*, 2009).

Oxygen deficiency is a common phenomenon and may occur in some hydroponic systems, especially floating culture, and in poorly-drained soils after a short period of heavy rain. Inhibitions of root cell division and decrease in root elongation have been found in plants

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hydroponically grown at lower dissolved O₂ concentrations (Atwell *et al.*, 1985). Furthermore, decrease in dissolved O₂ concentration decrease leaf water potential and stomatal conductance (Else *et al.*, 1995).

Because the lack of O₂ affects the energy status of the plant, root physiological functions such as respiration and water uptake are depressed (Drew, 1988; Chun and Takakura, 1994). Lack of O₂ may negatively affect the plant metabolism including nitrogen uptake and assimilation (Greenway and Gibbs, 2003; Morard *et al.*, 2004). Under oxygen deficiency condition, the rates of NO₃⁻ and NH₄⁺ uptake have been found to be very low (Brix *et al.*, 1994). Therefore, plants growing in O₂ deficient conditions often exhibit N deficiency symptoms along with O₂ stress symptoms, because N uptake is inhibited by oxygen deficiency (Greenway and Gibbs, 2003; Morard *et al.*, 2004).

Little information is available about the interaction between inorganic N form and dissolved O₂ concentrations and how it influences plant growth and nutrient concentration. In this study, we aimed to study hydroponically grown eggplant and measure some growth parameters and nutrient concentrations in response to different concentrations of dissolved O₂ and two forms of inorganic N.

MATERIALS AND METHODS

Plant Material and Culture Conditions

This study was performed with eggplant (*Solanum melongena* L. cv Long purple). The experiment was carried out as a factorial combination and completely randomized basic design with three replications. The first factor was N in two forms (including calcium nitrate and ammonium sulfate both at the N concentration of 5 mM) and the second factor was O₂ level of nutrient solution at 4 levels including: 1±0.3, 2±0.3, 3±0.3 and 4±0.3 mg L⁻¹ O₂. Seeds were germinated in the pots containing perlite medium. The seedlings were transferred to

pots containing 4 L of aerated nutrient solution. Four plants were grown together. The nutrient solution contained 0.2 mM KH₂PO₄, 0.2 mM K₂SO₄, 0.3 mM MgSO₄·7H₂O, and 0.1 mM NaCl. Micro-nutrients were 20 μM Fe-NaEDTA, 7 μM MnSO₄·H₂O, 0.7 μM ZnCl₂, 0.8 μM CuSO₄·5H₂O, 2 μM H₃BO₃, and 0.8 μM Na₂MoO₄·2H₂O with either 5 mM N as Ca (NO₃)₂·4H₂O or as (NH₄)₂SO₄ (Roosta and Schjoerring, 2007). Except N form, nutrient solution was the same in all treatments. For Ca adjustment in NH₄⁺ containing solution, CaCl₂ was used, so that Ca concentration was equal in both treatments. The pH was controlled at 6.5 to 7 throughout the growth period using Calcium Carbonate (CaCO₃) as a buffer. Solutions were changed completely every week in the first 5 weeks and subsequently every 4th day. The plants were grown in a greenhouse with 13 hours light (26±3°C) and 11 hours darkness (23±3°C). Three of the plants were harvested after 4 weeks and one plant remained in each pot. Plants after 9 weeks of initial treatment were harvested.

Oxygen Measurement and Adjustment

The atmospheric O₂ needed for the treatment was provided by an air pump with 2W power. Air distribution between pots was carried out with the use of medical serum set. Oxygen levels were controlled using portable O₂ meter (OXi 315, WTW Co., Germany) according to Chorianopoulou and Bouranis (2004).

Vegetative Growth Parameters

The height of the plants was measured on the harvest day by a ruler. Leaf number, stem number, fresh mass of leaves, roots, and stems were measured after harvesting the plants. The leaf area was determined using leaf area meter (model C1 202, USA). The organs of plants were put in the oven for 72 hours at 72°C, and then their dry masses were determined.

Nutrient Elements

Oven dried (72°C for 48 hours) samples of roots and shoots were weighed separately and ground to pass a 40-mesh sieve. The ground plant samples were dry-ashed at 500°C for 4 hours; the ashes were dissolved in 10 mL HCl (2N) and made the volume to 100 mL with distilled water. Concentration of Mg, Ca, Mn, Zn, and Cu was determined by atomic absorption spectrometry (Version 1/33 GBC Avanta, Au). K and Na concentrations were determined by flame photometry (JENWAY, PFP7, Germany) and P concentration in the digest was determined by the Olsen *et al.* (1954) method using spectrophotometry. Kjeldahl method was used to measure the total N.

Statistical Analysis

Analysis of variances were performed by using the SAS software (SAS Institute, Cary, NC), if ANOVA determined that the effects of the treatments were significant ($P < 0.05$ for F-test), then the treatment means were separated by LSD test.

RESULTS AND DISCUSSION

Vegetative Growth

Interactive effect of N-source and O₂ supply on fresh mass of shoot and leaves

were significant (Table 1). Both NO₃⁻ and NH₄⁺-fed plants had the greatest shoot and leaf fresh mass at 4 mg L⁻¹ O₂ concentration and the smallest at concentration of 1 mg L⁻¹ O₂. Growth inhibition by low O₂ may be due to the rise of toxic gases such as CO₂ in the root zone (Araki, 2006). Also, ethanol can be formed in plants under O₂ deficiency which is toxic in high concentrations (Marschner, 1995). The lack of oxygen in root tissues blocks mitochondrial respiration and thus reduces the uptake of some nutrients required for plant growth (Linkermer *et al.*, 1998; Sullivan *et al.*, 2001). The NH₄⁺-fed plants were more affected by high O₂ levels, as their shoot and leaf fresh masses were increased more than 2 and 3 times in 4 mg L⁻¹ O₂ in nutrient solution compared to 1 mg L⁻¹ O₂, respectively. This issue was reported by Jampeetong and Brix (2009) in the *Salvinia natans*, which was probably due to the decrease in toxic NH₄⁺ by rising O₂ concentration in the nutrient solution. High O₂ is needed for respiration and, consequently, carbon skeleton provision for NH₄⁺ assimilation (Roosta and Schjoerring, 2008a).

Nitrogen form and O₂ level had significant interactive effects on leaf area and stem number (Table 2). Both NO₃⁻ and NH₄⁺-fed plants had the greatest values of leaf area and stem number at 4 mg L⁻¹ O₂ concentration. It is reported that the Cytokinin (CK) production by root and its transportation from roots to leaves (von

Table 1. Interactive effects of N sources and O₂ supply on the shoot and leaf fresh masses of eggplant.^a

N form (5 mM)	O ₂ levels (mg L ⁻¹)			
	1±0.3	2±0.3	3±0.3	4±0.3
	Shoot fresh mass (g plant ⁻¹)			
NH ₄ ⁺	99.3 e	121.1 de	174.6 cd	275.2 b
NO ₃ ⁻	212.4 bc	274.0 b	397.8 b	414.9 a
	Leaf fresh mass (g plant ⁻¹)			
NH ₄ ⁺	41.9 e	56.6 de	86.9 cd	175.2 b
NO ₃ ⁻	107.4 c	152.1 b	219.6 a	248.6 a

^a Means in each column or row followed by the same letter(s) are not significantly different, LSD test ($P \leq 0.05$).

**Table 2.** Interactive effects of N sources and O₂ supply on leaf area and stem number of eggplant. ^a

N form (5 mM)	O ₂ levels (mg L ⁻¹)			
	1±0.3	2±0.3	3±0.3	4±0.3
	Leaf area (cm ² plant ⁻¹)			
NH ₄ ⁺	1923 f	1496 g	2895 e	6208 b
NO ₃ ⁻	3265 d	4373 c	6132 b	7210 a
	Stem number (plant ⁻¹)			
NH ₄ ⁺	7.33 d	13.00 c	17.33 b	22.33 a
NO ₃ ⁻	16.00 bc	18.00 b	22.67 a	26.00 a

^a Means in each column or row followed by the same letter are not significantly different, LSD test (P≤ 0.05).

Wiren *et al.*, 2000) and depressed root water uptake at low dissolved O₂ concentrations results in leaf turgor loss which cause decrease in leaf expansion (Yoshida *et al.*, 1997). A decrease in the number of lateral stems during the O₂ deficiency condition and NH₄⁺ presence has already been reported by Linkermer *et al.* (1998). It is reported that O₂ reduction decreased the CK production in the plant (Basra and Basra, 1997). Cytokinin induced lateral bud growth by decrease of apical dominance. Reduction in the number of stems in the current study can be due to the decrease of CK in plants.

Shoot and leaf fresh masses, leaf area, and stem number of NH₄⁺-fed plants were lower than NO₃⁻-fed plants at the same O₂ concentration in nutrient solutions (Table 1). This may be related to the decrease in some nutrient uptake, lowering pH of medium, hormonal imbalance, ethylene evolution, futile trans-membrane NH₄⁺ cycling, carbon skeleton depletion in root (Britto and Keronzucker, 2002; Thomas and Sodek, 2005; Roosta and Schjoerring, 2007; 2008a, b), and the photosynthesis reduction in NH₄⁺-fed plants (Takacs and Tecsı, 1992). Reduction of fresh and dry mass in the NH₄⁺-fed plants has already been reported in many species including cucumber (Roosta *et al.*, 2009), lettuce (Hay and Porter, 2006), onion (Gameily *et al.*, 1991), and soybean (Thomas and Sodek, 2005).

Both variables (N form and O₂ level) were effective on plant height and leaf number, although analysis of variances showed that

their interactive effect was not significant (Data not shown).

Nutrient Elements

The concentrations of Mg, Ca, Mn, and Na in root and shoot of the eggplants were found to be significantly affected by the interaction between N form and oxygen levels (P≤ 0.01).

Mg concentration was higher in leaves of NO₃⁻-fed plants as compared to those of NH₄⁺-fed plants. Both NO₃⁻- and NH₄⁺- fed plants had the highest leaf Mg concentration at 4 mg L⁻¹ O₂ level and the lowest at level of 1 mg L⁻¹ O₂ (Table 3). Decreased Mg concentration in the NH₄⁺-fed plants could be the result of a competition with NH₄⁺, as reported in cucumber plants by Roosta and Schjoerring (2007). Decrease in the content of Mg during O₂ deficiency has already been reported by Board (2008). Uptake of Mg into plant roots occurred by both passive process with high Mg concentration and active process with low Mg concentration in nutrient solution (Morard *et al.*, 2004). Active uptake of Mg is dependent on oxygen levels, thus, the active process may be stopped at low level of O₂, and uptake was merely a passive process.

Calcium concentration in leaves, but not in roots, increased with elevating O₂ concentrations for both NH₄⁺- and NO₃⁻- fed plants, however, NO₃⁻- fed plants had higher concentrations of Ca in the leaves and roots as compared to those of NH₄⁺-

Table 3. Interactive effects of N sources and O₂ supply on leaf and root Mg, Ca, Mn and Na in eggplant.^a

N form (5 mM)	O ₂ levels (mg L ⁻¹)			
	1±0.3	2±0.3	3±0.3	4±0.3
		Leaf Mg (%DM)		
NH ₄ ⁺	0.34 e	0.34 e	0.39 de	0.49 cd
NO ₃ ⁻	0.41 de	0.54 c	0.70 b	0.86 a
		Root Mg (%DM)		
NH ₄ ⁺	0.158 c	0.172 c	0.222 bc	0.336 ab
NO ₃ ⁻	0.331 ab	0.385 a	0.422 a	0.384 a
		Leaf Ca (% DM)		
NH ₄ ⁺	0.49 f	0.47 f	0.82 f	1.31 e
NO ₃ ⁻	1.85 d	2.6 c	3.31 b	3.74 a
		Root Ca (% DM)		
NH ₄ ⁺	0.30 b	0.33 b	0.39 b	0.32 b
NO ₃ ⁻	2.85 a	3.66 a	2.35 a	2.44 a
		Leaf Mn (mg kg ⁻¹ DM)		
NH ₄ ⁺	83.0 d	92.5 d	110.0 cd	168.5 a
NO ₃ ⁻	100.5 cd	129.5 bc	128.5 bc	142.0ab
		Root Mn (mg kg ⁻¹ DM)		
NH ₄ ⁺	18.77 b	17.60 b	21.22 b	27.06 a
NO ₃ ⁻	20.08 b	21.60 b	20.13 b	17.10 b
		Leaf Na (mg kg ⁻¹ DM)		
NH ₄ ⁺	0.09 b	0.11 b	0.14 b	0.15 b
NO ₃ ⁻	0.50 a	0.53 a	0.56 a	0.52 a
		Root Na (mg kg ⁻¹ DM)		
NH ₄ ⁺	1.87 c	0.82 d	0.85 d	0.91 d
NO ₃ ⁻	1.23bc	1.43 ab	1.63 a	1.48 a

^a Means in each column or row followed by the same letter(s) are not significantly different, LSD test (P≤0.01).

fed plants at the same O₂ concentrations (Table 3). Our results about the decrease of Ca concentration in NH₄⁺-fed plants agree with the findings of Tabatabaei *et al.* (2006) and Kotsiras *et al.* (2002). Reduced Ca concentration might have been due to either the reduced uptake of Ca (such as an antagonist effect) or the reduced translocation in the xylem (Marschner, 1995). A reduction in Ca content at the O₂ deficiency agrees with the findings of Board (2008). Although Ca uptake is a passive process and needs no energy, lack of O₂ conditions decreases the water uptake. The reduction in water uptake influences the transport of Ca in plant and thus results in decreasing Ca concentration in the plant organs.

Both NO₃⁻ and NH₄⁺-fed plants had the highest leaf Mn concentration at 4 mg L⁻¹ O₂

level and the lowest at level of 1 mg L⁻¹ O₂. NH₄⁺-fed plants had greater root Mn concentration than NO₃⁻-fed plants at 4 mg L⁻¹ O₂ level but not at the other O₂ concentrations (Table 3). Board (2008) reported that Mn concentrations in soybean leaf were lost due to the lack of O₂, which is consistent with our results.

In contrary to NO₃⁻-fed plants, NH₄⁺-supplied plants generally showed a trend toward decreasing root-Na concentrations with increasing O₂ level (Table 3). Decreased Na concentration in the root of NH₄⁺-fed plants may be due to the competition between these two elements for uptake by roots.

The interaction between N forms and O₂ levels was not found to be significant for leaf and root concentrations of N, P, K, Cu, and Zn (data not shown). The main effects



of N forms and O₂ levels on these nutrients are shown in Tables 4 and 5, respectively. The concentration of N was higher in leaves of plants fed with NH₄⁺ than those fed with NO₃⁻ (Table 4). The highest O₂ level caused a significant increase in N concentration (Table 5). Increased N uptake in NH₄⁺-fed plants has been reported in cucumber (Roosta and Schjoerring, 2007), and *Salvinia natans* (Jampeetong and Brix, 2009). On the other hand, decreased N concentration during the lack of O₂ has been reported in maize (Lizaso et al., 2001), and soybean (Sullivan et al., 2001; Board, 2008). Application of NH₄⁺ increased P concentration in leaf, but did not affect the P concentration in root (Table 4). Increased P concentration in NH₄⁺-fed plants has been reported by Roosta and Schjoerring (2007).

Plants fed with NH₄⁺ contain less low-molecular mass anions (NO₃⁻, carboxylates) and, consequently, have less negative charges to balance. Thus, they may uptake more phosphorus for provision of anion equivalents and charge balance (Roosta and Schjoerring, 2007; Zhang et al., 2005). The highest O₂ level caused a significant decrease in P concentration in leaf and root (Table 5).

Potassium concentration was higher in leaves of plants fed with NH₄⁺ than those fed with NO₃⁻ (Table 4). On the other hand, high O₂ level in nutrient solution decreased K concentration in the leaf, but not in the root (Table 5). The lower K concentration in leaves of NO₃⁻ grown plants in the current experiment could be due to the dilution effects as a result of higher growth.

Table 4. Effect of N form in the nutrient solution on nutrients concentrations in leaf and root of eggplant.^a

Nutrient element	N form (5 mM)	
	NH ₄ ⁺	NO ₃ ⁻
Leaf N (% DM)	4.64 a	3.30 b
Leaf P (% DM)	0.043 a	0.021 b
Root P (% DM)	0.36 a	0.367 a
Leaf K (% DM)	4.54 a	2.02 b
Root K (% DM)	0.973a	0.849a
Leaf Zn (mg kg ⁻¹ DM)	35.00 a	10.00 b
Root Zn (mg kg ⁻¹ DM)	139.5 a	103.1 b
Leaf Cu (mg kg ⁻¹ DM)	58.25 b	63.38 a
Root Cu (mg kg ⁻¹ DM)	167.5 a	125.8 b

^a Means with the different letter in each row are significantly different, LSD test (P≤ 0.01).

Table 5. Effect of O₂ levels in the nutrient solution on nutrients concentrations in leaf and root of eggplant.^a

Element	O ₂ Levels (mg L ⁻¹)			
	1±0.3	2±0.3	3±0.3	4±0.3
Leaf N (% DM)	3.90 b [†]	3.80 b	3.80 b	4.20 a
Leaf P (% DM)	0.42 a	0.44 a	0.33 ab	0.26 b
Root P (% DM)	1.39 ab	1.56 a	1.47 a	0.35 b
Leaf K (% DM)	4.03 a	3.80 a	2.72 b	2.57 b
Root K (% DM)	1.02 a	1.12 a	0.874 a	0.628 a
Leaf Cu (mg kg ⁻¹)	57.25 b	57.50 b	62.25 ab	66.25 a
Root Cu (mg kg ⁻¹)	136.0 b	163.5 a	156.5 a	130.0 b
Leaf Zn (mg kg ⁻¹)	21.5 a	19.8 a	22.8 a	26 a
Root Zn (mg kg ⁻¹)	147.72 a	147.75 a	121.00 a	68.75 b

^a Means with the same letters in each row are not significantly different, LSD test (P≤ 0.01).

Increasing the concentration of K in NH_4^+ grown plants might be also due to the plant demands for increase in NH_4^+ assimilation via elevating of glutamine synthetase activity. Increase of glutamine synthetase activity by K has already been reported (Roosta and Schjoerring, 2008b). Low-level O_2 increased K concentration in the leaves. These results were the same as the results achieved by Sullivan *et al.* (2001) on soybean.

The results of this research showed that NH_4^+ -fed plants had higher concentrations of Zn in the leaves and roots as compared to those of NO_3^- -fed plants (Table 4). Increasing the concentration of Zn in plants fed with NH_4^+ has been reported in lettuce (Roosta and Schjoerring, 2007), spinach (Assimakopoulou, 2006), and azalea (Clark and *et al.*, 2003). At the highest level of O_2 i.e. 4 mg L^{-1} , Zn concentration in the root decreased compared to the other O_2 concentrations (Table 5).

Compared to NO_3^- -fed plants, the NH_4^+ -fed plants had lower Cu concentration in the leaf and higher Cu in the root (Table 4). This is in agreement with the findings of Roosta and Schjoerring (2007) and Clark *et al.* (2003) who reported NH_4^+ application decreased Cu concentration in the leaves of cucumber and azalea, respectively. Leaf Cu concentration generally showed a trend toward increasing with increasing O_2 concentrations, while root Cu concentration varied widely across O_2 treatments (Table 5). Chorianopoulou and Bouranis (2004) observed that short-term lack of O_2 increases the Cu concentration in leaves and roots of water cress.

CONCLUSIONS

This study showed that overall growth of eggplant was significantly decreased by NH_4^+ , whereas high levels of oxygen increased the vegetative growth. Both NO_3^- - and NH_4^+ -fed plants had the greatest shoot and leaf fresh mass at 4 mg L^{-1} O_2 concentration and the smallest at

concentration of 1 mg L^{-1} O_2 . NH_4^+ application decreased the concentration of Mg, Ca, Cu, and Mn in plants, compared to NO_3^- . On the other hand, the increase in O_2 amount of nutrient solutions partly alleviated ammonium toxicity by increasing Mg, Ca, Cu, and Mn concentration in plants. Therefore, in floating hydroponic cultures, O_2 level and its distribution should be controlled and must not be lower than 4 mg L^{-1} .

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REFERENCES

1. Araki, H. 2006. Water Uptake of Soybean (*Glycine max* L. Merr) during Exposure to O_2 Deficiency and Field Level CO_2 Concentration in the Root Zone. *Field Crop Res.*, **96**: 98-105.
2. Assimakopoulou, A. 2006. Effect of Iron Supply and Nitrogen Form on Growth, Nutritional Status and Ferric Reducing Activity of Spinach in Nutrient Solution Culture. *Sci. Horti.*, **110**: 21-29.
3. Atwell, B. J., Thomson, C. J., Greenway, H., Ward, G. and Waters, I. 1985. A Study of the Impaired Growth of Roots of *Zea mays* Seedlings at Low Oxygen Concentrations. *Plant Cell Environ.*, **8**: 179-188.
4. Barker, A. V. and Breyson, G. M. 2006. Nitrogen. In: "*Handbook of Plant Nutrition*", (Eds.): Barker, A. V. and Pilbeam, D. J.. CRC Press, Boca Raton, Florida, PP. 21-50.
5. Basra, A. S. and Basra, P. K. 1997. Mechanisms of Environmental Stress Resistance in Plants. Hardwood Academic Publishers. 83-111.
6. Board, J.E. 2008. Water Logging Effects on Plant Nutrient Concentrations in Soybean. *J. Plant Nutr.*, **31**: 828-838.
7. Britto, D. T. and Krounzucker, H. J. 2002. NH_4^+ Toxicity in Higher Plants. *Plant Physiol.*, **159**: 567-584.



8. Brix, H., Lorenzen, B., Morris, J. T., Schierup, H. H. and Sorrell, B. K. 1994. Effects of Oxygen and Nitrate on Ammonium Uptake Kinetics and Adenylate Pools in *Phalaris arundinacea* L. and *Glyceria maxima*. (Hartm) Holmb. *Proc. R. Soc. Edinb.* **102(B)**: 333–342.
9. Chorianoopoulou, S. N. and Bouranis, D. L. 2004. Alterations in Short-term Effect of Oxygen Deficiency on Iron, Manganese, Zinc, and Copper Homeostasis within Fool's Watercress Organs during Development. *J. Plant Nutr.*, **27**: 157-171.
10. Chun, C. and Takakura, T. 1994. Rat of Root Respiration of Lettuce under Various Dissolved Oxygen Concentrations in Hydroponics. *Environ. Cont. Biol.*, **32(2)**: 125-135.
11. Clark, M. B., Mills, H. A., Robacker, C. D. and Latimer, J. G. 2003. Influence of Nitrate Ammonium Ratios on Growth and Elemental Concentration in Two Azalea Cultivars. *J. Plant Nutr.*, **26(12)**: 2503–2520.
12. Drew, M. C. 1988. Effects of Flooding and Oxygen Deficiency on Plant Mineral Nutrition. In: “*Advances in Plant Nutrition*“, (Eds.): Lauchli, A. and Tinker, P. B.. Praeger, New York, PP. 115-159.
13. Else, M. A., Davies, W. J., Malone, M. and Jackson, M. B. 1995. A Negative Hydraulic Message from Oxygen-deficient Roots of Tomato Plants: Influence of Soil Flooding on Leaf Water potential, Led Expansion, and Synchrony between Stomatal Conductance and Root Hydraulic Conductivity. *Plant Physiol.*, **109(3)**: 1017–1024.
14. Gameily, S., Randle, W. M., Mills, H. A. and Smittle, D. A. 1991. Onion Plant Growth, Bulb Quality, and Water Uptake Following Ammonium and Nitrate Nutrition. *HortSci.*, **26(8)**: 1061-1063.
15. Greenway, H. and Gibbs, J. 2003. Mechanisms of Anoxia Tolerance in Plants: A Review. II. Energy Requirements for Maintenance and Energy Distribution to Essential Processes. *Funct. Plant Biol.*, **30(10)**: 999–1036.
17. Hay, R. K. M. and Porter, J. R. 2006. *The Physiology of Crop Yield*. 2nd Edition, Black Well Publishing, Singapore.
18. Jampeetong, A. and Brix, H. 2009. Oxygen Stress in *Salvinia natans*: Interactive Effects of Oxygen Availability and Nitrogen Source. *Environ. Exp. Bot.*, **66(2)**: 153-159.
19. Kotsiras, A., Olympios, C. M., Drosopoulos, J. and Passam, H. C. 2002. Effect of Nitrogen Form and Concentration on the Distribution of Ions within Cucumber Fruit. *J. Amer. Soc. Horti. Sci.*, **95**: 175–183.
20. Linkermer, G., Board, J. E. and Musgrave, M. E. 1998. Waterlogging Effects on Growth and Yield Components in Late-planted Soybean. *Crop Sci.*, **38**: 1576-1584.
21. Lizaso, J., Melendez. L. M. and Ramirez, R. 2001. Early Flooding of Two Cultivars of Tropical Maize. II. Nutritional Responses. *J. Plant Nutr.*, **24**: 997-1011.
22. Marschner, H. 1995. *Mineral Nutrition of Higher Plants*. 2nd Edition, Academic Press, London.
23. Morard, P., Lacoste, L. and Silvestre, J. 2004. Effect of Oxygen Deficiency on Mineral Nutrition of Excised Tomato Roots. *J. Plant Nutr.*, **27(4)**: 613-626.
24. Olsen, S. R., Cole, C. V., Watanabe, F. S., and Dean, L. A. 1954. *Estimation of Available Phosphorous in Soil by Extraction with Sodium Bicarbonate: USDA Circ. 939*. US Gov. Print. Office, Washington, DC, USA.
25. Roosta, H. R. and Schjoerring, J. K. 2007. Effects of Ammonium Toxicity on Nitrogen Metabolism and Elemental Profile of Cucumber Plants. *J. Plant Nutr.*, **30**: 1933-1951.
26. Roosta, H. R. and Schjoerring J. K. 2008a. Root Carbon Enrichment Alleviates Ammonium Toxicity in Cucumber Plants. *J. Plant Nutr.*, **31**: 941-958.
27. Roosta, H. R. and Schjoerring, J. K. 2008b. Effects of Nitrate and Potassium on Ammonium Toxicity in Cucumber Plants. *J. Plant Nutr.*, **31**: 1270-1283.
28. Roosta, H. R., Sajjadinia, A., Rahimi, A. and Schjoerring, J. K. 2009. Responses of Cucumber Plant to NH_4^+ and NO_3^- Nutrition: The Relative Addition Rate Technique vs. Cultivation at Constant Nitrogen Concentration. *Sci. Horti.*, **121(4)**: 397-403.
29. Sullivan, M., Van Toai, T., Fausey, N., Beuerlein, J., Parkinson, R. and Soboyejo, A. 2001. Evaluating On-farm Flooding Impacts on Soybean. *Crop Sci.*, **41(1)**: 93–100.
30. Tabatabaei, S. J., Fatemi, L. S. and Fallahi, E. 2006. Effect of Ammonium: Nitrate Ratio on Yield, Calcium Concentration, and

- Photosynthesis Rate in Strawberry. *J. Plant Nutr.*, **29(7)**: 1273-1285.
31. Takacs, E. and Tecs, L. 1992. Effects of $\text{NO}_3^-/\text{NH}_4^+$ Ratio on Photosynthetic Rates, Nitrate Reductase Activity and Chloroplast Ultra Structure in Three Cultivars of Red Pepper (*Capsicum annuum L.*). *Plant Physiol.*, **140(3)**: 298-305.
32. Thomas, A. L. and Sodek, L. 2005. Development of the Nodulated Soybean Plant after Flooding of the Root System with Different Sources of Nitrogen. *Braz. J. Plant Physiol.*, **17(3)**: 291-297.
33. Yoshida, S., Kitano, M. and Eguchi, H. 1997. Growth of Lettuce Plants (*Lactuca sativa L.*) under Control of Dissolved O_2 Concentration in Hydroponics. *Biotronic.*, **26**: 39-45.
34. von Wiren, N., Gazzarrini, S., Gojont, A. and Frommer, W. B. 2000. The Molecular Physiology of Ammonium Uptake and Retrieval. *Curr. Opin. Plant Biol.*, **3(3)**: 254-261.
35. Zhang, Y., Lin, X., Zhang, Y., Zheng, S. J. and Du, S. 2005. Effects of Nitrogen Levels and Nitrate/Ammonium ratios on Oxalate Concentrations of Different Forms in Edible Parts of Spinach. *J. Plant Nutr.*, **28**: 2011-2025.

اثرات متقابل شکل نیتروژن و غلظت اکسیژن بر رشد و وضعیت تغذیه‌ای بادمجان در سیستم هیدروپونیک

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چکیده

تحقیق گلخانه‌ای برای تعیین اثر شکل‌های نیتروژن و سطوح مختلف اکسیژن بر رشد و غلظت عناصر غذایی در بادمجان انجام شد. آزمایش بصورت فاکتوریل و در قالب طرح کاملاً تصادفی با دو فاکتور شکل نیتروژن $\text{Ca}(\text{NO}_3)_2$ و $(\text{NH}_4)_2\text{SO}_4$ و سطوح اکسیژن (1 ± 0.3) ، (2 ± 0.3) ، (3 ± 0.3) و (4 ± 0.3) میلی گرم بر لیتر اکسیژن انجام شد. نتایج نشان داد که کاربرد آمونیوم همه پارامترهای رشد رویشی اندازه‌گیری شده را کاهش داد، در صورتیکه سطوح بالای اکسیژن آنها را افزایش داد. کاربرد آمونیوم غلظت نیتروژن، فسفر، پتاسیم و روی را در برگ‌ها و روی و مس را در ریشه‌ها افزایش داد، در صورتیکه غلظت منیزیم، کلسیم، مس، منگنز و سدیم را در برگ‌ها و کلسیم، منیزیم، منگنز و سدیم را در ریشه‌ها کاهش داد. سطوح بالای اکسیژن میزان نیتروژن، منیزیم، کلسیم، مس و منگنز را در برگ‌ها و میزان منگنز و سدیم را در ریشه‌ها افزایش داد، در صورتیکه غلظت پتاسیم برگ‌ها و فسفر و روی ریشه‌ها را کاهش داد. بر طبق نتایج بدست آمده افزایش میزان اکسیژن در محلول غذایی تا حدودی سمیت آمونیوم را در گیاه بادمجان کاهش داد. بنابراین، در کشت هیدروپونیک شناور، سطح اکسیژن و توزیع آن باید کنترل شده و میزان آن نباید پایین‌تر از ۴ میلی گرم بر لیتر باشد.