# High Prevalence of Aflatoxin B<sub>1</sub> in Aspergillus flavus Infecting Stored Rice Grains

M. B. S. Al-Shuhaib<sup>1</sup>, A. H. Albakri<sup>2</sup>, H. O. Hashim<sup>3</sup>, S. L. Alwan<sup>2</sup>, N. B. Almandil<sup>4</sup>, P. Selvaraj<sup>5</sup>, R. Jermy<sup>6</sup>, S. Abdul Azeez<sup>7</sup>, and J. Francis Borgio<sup>7\*</sup>

# **ABSTRACT**

One of the best-known rice-infecting microorganisms is Aspergillus flavus, which produces toxic metabolites known as Aflatoxins (AFs). This study was designed to detect potential simultaneous biosynthesis of the four main AFs (AFB<sub>1</sub>, AFB<sub>2</sub>, AFG<sub>1</sub>, and AFG<sub>2</sub>) in rice-infecting strains of A. flavus. The AF prevalence was studied in 109 strains of A. flavus, which were collected from stored Indian rice grains from 300 locations in the Middle Euphrates region of Iraq from 2015 to 2016. The potential AFs were extracted and quantified simultaneously using High-Performance Liquid Chromatography (HPLC) equipped with a photodiode array detector. The results revealed that 29% (n= 32) of strains were non-aflatoxigenic, while the remaining 71% (n= 77) were confirmed to be aflatoxigenic, with variable ability to produce mono-, bi-, and tri-AFs. AFB<sub>1</sub>, AFG<sub>2</sub>, AFG<sub>1</sub>, and AFB<sub>2</sub> were produced by 49 (45%), 44 (40%), 20 (18%), and 17 (16%) strains, respectively, at various concentrations. The concentration of AFB<sub>1</sub> was the highest among the A. flavus strains, with a mean value of 3,561.9 µg kg<sup>-1</sup>. In conclusion, the most abundant AF synthesized by the rice-infecting A. flavus strains was AFB<sub>1</sub>. Contamination with AFs continues to pose potential health risks to animals as well as humans. These results clearly indicate that the improper storage conditions of rice in Iraq were favourable for the growth of A. flavus and contamination with AFs. National-level studies are mandatory to avoid foodborne intoxications. Strict regulations should be devised and imposed to prevent synthesis of AFs on rice grains.

Keywords: Aflatoxigenic, Contamination, Indian rice, HPLC, Hygiene.

# **INTRODUCTION**

Aflatoxins (AFs) are toxic secondary metabolites that are mainly synthesised by Aspergillus flavus, A. parasiticus, and A. nomius, which infect several economically important crops (Al-Shuhaib et al., 2018;

Hasanvand *et al.*, 2016; Heshmati *et al.*, 2019). The growth of aflatoxigenic *A. flavus* strains on certain foods and feeds may result in production of AFs, which can cause illness or lead to the death of humans and animals; therefore, it is an important public health concern (Aydin *et al.*, 2007; Giray *et* 

<sup>&</sup>lt;sup>1</sup> Department of Animal Production, College of Agriculture, Al-Qasim Green University, Al-Qasim, Babil 51001, Iraq.

<sup>&</sup>lt;sup>2</sup> Department of Plant Protection, College of Agriculture, University of Kufa, Kufa, Najaf 54001, Iraq.

<sup>&</sup>lt;sup>3</sup> Department of Clinical Laboratory Sciences, College of Pharmacy, University of Babylon, Babil 51001, Iraq.

<sup>&</sup>lt;sup>4</sup>Department of Clinical Pharmacy Research, Institute for Research and Medical Consultation (IRMC), Imam Abdulrahman Bin Faisal University (Formerly: University of Dammam), Dammam, Saudi Arabia.

<sup>&</sup>lt;sup>5</sup> Department of Zoology, St. Xavier's College (Autonomous), Palayamkottai-627002, Tamil Nadu, India.

<sup>&</sup>lt;sup>6</sup> Department of Nanomedicine, Institute for Research and Medical Consultation (IRMC), Imam Abdulrahman Bin Faisal University, Dammam 31441, Saudi Arabia.

<sup>&</sup>lt;sup>7</sup> Department of Genetic Research, Institute for Research and Medical Consultation (IRMC), Imam Abdulrahman Bin Faisal University, Dammam, Saudi Arabia.

<sup>\*</sup>Corresponding author; e-mail: fbalexander@iau.edu.sa



al., 2007). At present, about 20 AFs have been identified, but those most frequently found in foods like cereals, rice, and corn are AFB<sub>1</sub>, AFB<sub>2</sub>, AFG<sub>1</sub>, and AFG<sub>2</sub> (Li et al., 2014), of which AFB<sub>1</sub> is the most toxic (Erkmen and Bozoglu, 2008). Due to their toxic and carcinogenic potential, AFs have garnered considerable attention. Moreover, AFs are the most hazardous and mutagenic natural substances (Paterson and Lima, 2009). AFs are associated with aflatoxicosis in a variety of mammals, birds, and fishes (Frisvad et al., 2006); hepatocytes are believed to be the primary target cells (Towner et al., 2000). AFs are generally classified as group I carcinogenic substances by the International Agency of Research on Cancer (IARC), with a particular emphasis on AFB<sub>1</sub> (IARC, 1993). The increasing prevalence of AF contamination in rice has become a major concern for the scientific community because it poses a public health Several rice-based dishes threat. essential components of daily meals in the Middle East and Indian subcontinent, which are well known for having high rice consumption. During storage, mycotoxigenic mould may grow on rice and cause mycotoxicosis when ingested (Villers, 2014). Rice represents an attractive substrate for fungal growth and toxinogenesis, and can be used as an ideal growth medium to investigate the potential toxicity of fungal strains (Bars and Bars, 1992). Previous studies have reported high natural levels of AFs in stored rice samples (Tanaka et al., 2007). Recently, high levels of AF contamination by moulds due to improper storage have been widely reported (Al-Zoreky and Saleh, 2017; Tournas and Niazi, 2018; Majeed et al., 2018). Development of AF contamination during storage is largely due to delayed crop mortality and excessive moisture, which is relatively high in tropical countries with high temperature and humidity (Cho et al., 2008; Rahman et al., 2017). In addition to climatic conditions, storage conditions of rice can exert a considerable influence on the fungal load and AF contents of the rice. Therefore,

examination of stored rice grains intended for human consumption is necessary to determine the risk of AFs for consumers. Several countries have introduced strict guidelines regarding AF contamination in food commodities. Given the widespread occurrence of AF-producing fungi and AFs in some crops, several robust biological and chemical methods have been developed for AF detection. However, these methods can be time-consuming (Zheng et al., 2016) and require specialized expertise (Sirhan et al., 2014) or expensive enzymatic preparations (Xiao-han et al., 2017). High-Performance Liquid Chromatography (HPLC) coupled to a fluorescence detector remains the most versatile technique due to its high sensitivity and relatively low cost (Wen et al., 2013; Mashak et al., 2016). The present study was conducted to identify and quantify the AFs from A. flavus isolated from stored rice grains, and to estimate the pattern and severity of mycotoxin contamination in the rice. To the best of our knowledge, data concerning AF production in contaminating A. flavus strains from Iraq are therefore, the present constitutes the first large-scale qualitative and quantitative investigation of naturally biosynthesized AFs in this country.

# MATERIALS AND METHODS

# Rice-Grain Sources and Isolation of A. flavus

A total of 300 Indian rice samples were purchased at random from local markets in several Middle Euphrates regions in Iraq during 2015–2016. Among the microorganisms observed in the cultured rice grains, 109 fungal strains of *A. flavus* were isolated based on macro- and microscopic morphological characteristics. All characteristics (i.e. spore arrangement, spore morphology, and pattern and colour of the fungal colony) of the targeted *A. flavus* strains were identified using a classic species description reference (Klich, 2002).

The *A. flavus* colonies were identified genetically, as described in our previous study (Al-Shuhaib *et al.*, 2018).

# **Extraction of Aflatoxins**

All 109 isolates of *A. flavus* were grown separately on 100 mL of potato dextrose liquid culture medium at 35°C for 18 days. Then, 50 mL of the extracellular mycelial growth was mixed with an equal volume of chloroform and incubated on a rotary shaker for 4 hours. After the toxins dissolved in the solvent, the aqueous layer was discarded, and the remaining fraction was stored at 50°C until all of the chloroform evaporated. The leftover precipitate was dissolved thoroughly with 1 mL of methanol.

#### **HPLC**

HPLC experiments were performed at the Department of Clinical Laboratory Analysis, College of Pharmacy, University Babylon. Before injection, the samples were filtered through 0.45-mm filters (Millipore Corporation, Bedford, MA, USA). Then, 20 µL of the supernatant was injected into an HPLC system equipped with a photodiode array detector, which provides sensitivity, baseline stability, and analytical reliability. The system control and data acquisition were performed using LabSolutions LC WorkStation software (Ver.5.51; Shimadzu, Kyoto, Japan). The analyses conducted were SUPELCOSIL<sup>TM</sup> LC-18 HPLC column (L×ID: 25 cm×4.6 mm, Particle size: 5 μm; Cat. No. 58298; Supelco, Bellefonte, PA, USA) at 40°C using acetonitrile, methanol, and water (20, 20, and 60%, respectively) as a mobile phase with a flow rate of 1.5 mL min<sup>-1</sup> for 15 minutes, following the manufacturer's instructions. To identify the AFs, the observed retention time and UV absorbance spectra of sample peaks were compared and matched to those obtained from an AF mix reference standard (Cat. No. 46304-U; Supelco). Quantitation of AFs was conducted by comparing the peak area of the target peaks (at 365 nm) to the standard curve generated using several dilutions of the AF mix reference standard.

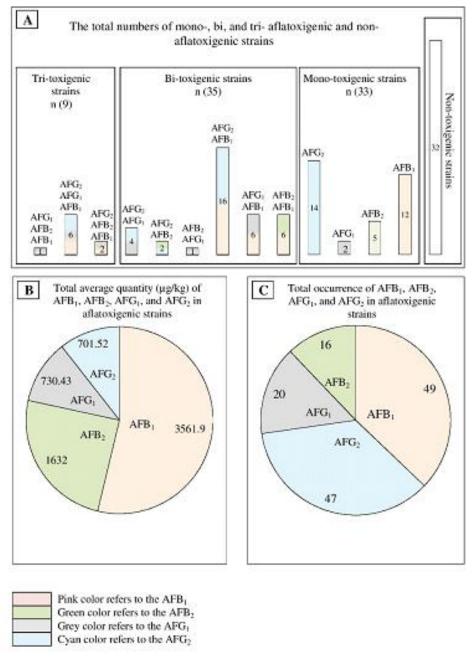
# RESULTS AND DISCUSSION

This detailed study on AF production by 109 A. flavus isolates revealed that 71% (n= 77) of the total isolates were aflatoxigenic (Figure 1 and Table 1). The low number (n= 32; 29%) of non-aflatoxigenic A. flavus clearly demonstrates the importance of screening for rice-infecting A. flavus strains and the impact of AFs in general (and AFB<sub>1</sub> in particular) on human and animal health. The analyzed strains presented a variety of synthesized AF combinations concentrations. Vaamonde et al. (2003) observed similar differences in the ability of A. flavus strains to produce AFs in various crops. Notably, the isolates included mono-, bi-, and tri-aflatoxigenic strains, but no observed isolate appeared capable of producing all four AFs together. Although a previous report indicated the concomitant absence of AFB<sub>2</sub>, AFG<sub>1</sub>, and AFG<sub>2</sub> (Dhanasekaran et al., 2011), the ricecontaminating A. flavus isolates in the present study produced four types of AFs (Table 1). These results contrast with those of previous studies indicating the ability of A. flavus strains to produce only AFB<sub>1</sub> and AFB<sub>2</sub> (Ehrlich et al., 2004; Frisvad et al., 2005). However, the present results were consistent with the observed production of AFG<sub>1</sub>, AFG<sub>2</sub>, AFB<sub>1</sub>, and AFB<sub>2</sub> by A. parasiticus strains (González-Salgado et al., 2008). It has been reported that a high percentage of A. flavus strains synthesize AFG<sub>1</sub>, and a minor group also accumulates AFG<sub>2</sub> (Karaaslan and Arslangray, 2015). Certain A. flavus strains have been shown to produce G-type AFs (Pildain et al., 2008). Another study reported that AFB- and AFGproducing A. flavus strains belong to group II of the same species, which differ from the main group I strains that produce only AFBs



(Geiser *et al.*, 2000). In the present study, although most of the strains were found to produce the four main AFs, AFB<sub>1</sub> concentration and prevalence were consistently higher than those of the other three AFs (Table 1). In fact, AFB<sub>1</sub> accounted for more than half of the total AF content

(Figure 1-b). The HPLC results showed that AFB<sub>1</sub>, AFG<sub>2</sub>, AFG<sub>1</sub>, and AFB<sub>2</sub> were produced by 49, 45, 16, and 14 aflatoxigenic isolates, respectively, out of the 109 *A. flavus* strains analyzed (Figure 1-c). The results clearly demonstrated that AFB<sub>1</sub> had the highest concentration in many *A. flavus* 



**Figure 1.** The pattern of aflatoxigenicity in 109 *Aspergillus flavus* isolates infecting rice grains: (**A**) The numbers of mono-, bi-, tri-, and non-aflatoxigenic isolates; (**B**) Mean concentration of Aflatoxins (AFs) observed in *Aspergillus flavus* isolates infecting rice grains, (**C**) The prevalence of AF types observed in the *Aspergillus flavus* isolates.

Table 1. The concentrations of aflatoxins B<sub>1</sub>, B<sub>2</sub>, G<sub>1</sub>, and G<sub>2</sub> (µg mL<sup>-1</sup>) in each Aspergillus flavus strain.<sup>a</sup>

																												9
G2	4.41		ē		1	,	1.12	48.18	•	,	8.34	18.08	4.13		ı	16.81	1	11.03	1	,	23.68	•	1	ľ	•			
G1	10.72	1	9	1	1		1	1	245.46	1	1		,	16.55	Ľ	60.52	1	r		7.60			,	•	118.78			
B2		•	ï		1	148.52	45.69	,	·			·	,		ı		,	274.15		þ	ě	•	•		150.58			
B1	412.54	82.77	413.51	687.65	292.61	18483.5	263.48	921.93	715.64	180.15	473.30	569.11	94.88	124.00	30.31	21.79	1	Ü	ī	ì	í	ı	1321.9	238.82	22.93			
No.	85	98	87	88	68	06	91	92	93	94	95	96	26	86	66	100	101	102	103	104	105	106	107	108	109			
G2	2.80	1	e	1	1	00.9	3.06		90.95	1	1	E			r		2.43	ľ	71.50		78.87	147.16	24.26		8.36	,	4.41	10.28
Gl		1	6		116.47	9.91	18.56	1			1				ı		6.17	ı		1	Ü	11.79	,			09.9	ij.	24.94
B2	11.67	17.62	1	ı	1		Ţ	1	•	57.34	1	•	,	39.62	r	76.65		r		7	ľ	•	•	0	ŗ	•	C	ï
B1			562.88	ì	903.03	ī	33.31	,	í	ī	,	í	,	292.36	i	246.63		9.56	695.31	î	16.03	34.45	,	3.53	20.67	31.47	Ç	125.29
No.	57	58	59	09	61	62	63	64	65	99	29	89	69	70	71	72	73	74	75	9/	11	78	79	80	81	82	83	84
G2	3.77	24.09	11.01	11.63	1	E		40.71	r		1	E	,	1	r		1	E		1	E	,	1	Е		1	08.9	,
Gl				,	1	ı	ı	1		ı	1	1	1		ı			ľ	,	ì	ı		24.45		6.79		ı	ı
B2	at	2 <b>1</b> .	24.43	1	8 <b>1</b> 0	I.		1	ı	31	739.20	ı	1	.1	7.70		( <b>1</b> )	I,	,	31	E		1	I.S	4.78	5.84	15.44	ı
B1	90.84		i	i	1	8.18	12.00	1	ï	ı		ï	ī		ť	•		ť	i	j	Ē	,	ā	ī	ï	3.85	6389.1	
No.	59	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	20	51	52	53	54	55	99
G2	26.65	19.24	9.29	0.67	4.04	6.75	0.67	0.67	6.45	5.57	2.39	2.52	7.35	8.94	2.82	5.25	25.94	r	ı	3	ē	ı	1	1.96	3.42	,	Ĺ	3.29
Gl	8.07		ı	1	14.09	ı	ı	1	1	ı	6.73	1	ı	1	ï	,	1	ı	ī	9	ě	ì	1	80.6	ì	1	Ü	7.15
B2	1	1	ť	1	1	1	1	1	ı	1	•	ı	1	1	r	1	1	r	,	1	į.	1	12.85	1	í	1	ı	ı
BI	244.04	245.76	12.80	,	1	129.03	,	38.18	5.64	ı	1	ı	47.47	10.19	7.14		,	ř	29.79	1	ı		14.85	37.70		,	ı	ı
No.	-	7	3	4	5	9	_	<b>%</b>	6	10	Ξ	12	13	4	15	16	17	18	19	20	21	22	23	24	25	26	27	28

" No: Refers to the strain number included in the present study. B1: AFB\_1 is in pink; G1: AFG\_1 is in grey; B2: AFB\_2 is in green; G2: AFG\_2 is in cyan. The mean $\pm$ SD of the toxins: B1: 728.20 $\pm$ 2750.61; B2: 102.05 $\pm$ 185.15; G1: 36.52 $\pm$ 59.7; G2: 17.61 $\pm$ 27.96.

JAST



strains (Table 1), with an average concentration of 3,561.9 µg kg<sup>-1</sup>. This AFB<sub>1</sub> level was considerably higher than the allowable limit (~30 µg kg<sup>-1</sup>) (Nguyen et al., 2007). Our results are consistent with studies indicating that AFB<sub>1</sub> is the most prevalent AF contaminant and has the highest concentration among rice-infecting A. flavus isolates (Lai et al., 2015; Reddy et al., 2011; Sun et al., 2017). The high prevalence and concentration of AFB<sub>1</sub> makes the daily consumption of stored rice grains potentially dangerous. One limitation of this study was a lack of data on the storage duration of the collected rice grains. Therefore, further large-scale or national-level studies are required to avoid foodborne intoxication. Yazdanpanah et al. (2013)also recommended regular screening for AFB<sub>1</sub>infected rice. Such screening is critical because the consumption of a high quantity of AFB<sub>1</sub> is toxic to the liver; the cyclic nucleotide phosphodiesterase activity in the brain, liver, heart and kidney tissues can be inhibited by AFB<sub>1</sub>, resulting in severe impairment of the metabolism of proteins, carbohydrates, and lipids in the liver (Bonsi et al., 1999). Chronic intake of such toxins could act synergistically with other factors, such as the hepatitis B virus, to promote liver cancer (Chawanthayatham et al., 2017). In this study, AFB<sub>2</sub> had the second highest concentration among the 109 isolates (1,632 µg kg<sup>-1</sup>). Previous studies have shown that rice is susceptible to AFB<sub>1</sub> and AFB<sub>2</sub> accumulation (Lai et al., 2015; Reddy et al., 2011). Both the present observations and previous results indicate that rice grains are a major source of AF contamination. These findings reaffirm that the risk of AF poisoning in humans via rice contaminated by toxigenic A. flavus strains cannot be eliminated unless proper measures are taken. Consumption of rice is high in many countries; therefore, monitoring concentrations of AFB<sub>1</sub> in stored rice grains is strongly recommended as a preventive measure. The occurrence of AFs in most samples analyzed herein indicates that there is a need for regular, national-level programs

to monitor the concentrations of AFs in stored rice grains. The concentration of AFs should be reduced by adopting proper storage practices, such as Good Agricultural Practices (GAP) and Good Handling Practices (GHP) in Iraq.

# **CONCLUSIONS**

AF contamination by rice-infecting *A. flavus* strains is a serious concern. AFB<sub>1</sub> was the most prevalent of the four AFs analyzed in *A. flavus* strains isolated from rice grains. The results clearly indicated that the conditions and methods of rice storage used in Iraq were favourable for the growth of *A. flavus* and AF contamination. Therefore, future studies aiming to elucidate any possible etiological influence of AFs on the incidence of liver cancer in Iraq are strongly recommended. Strict regulations should also be devised and implemented to prevent further synthesis of AFs on rice grains.

# **ACKNOWLEDGEMENTS**

The authors thank Dr. Tahreer M. Al-Thuwaini, Department of Animal Production, College of Agriculture, Al-Qasim Green University for the kind scientific support.

# REFERECNES

- Al-Shuhaib, M. B. S., Albakri, A. H., Alwan, S. H., Almandil, N. B., Abdul Azeez, S. and Borgio, J. F. 2018. Optimal PCR Primers for Rapid and Accurate Detection of Aspergillus flavus Isolates. Microb. Pathog., 116: 351-355.
- Al-Zoreky, N. S. and Saleh, F. A. 2017. Limited Survey on Aflatoxin Contamination in Rice. Saudi J. Boil. Sci., 26: 225-231.
- 3. Aydin, A., Erkan, M. E., Başkaya, R. and Ciftcioglu, G. 2007. Determination of Aflatoxin B1 Levels in Powdered Red Pepper. *Food Control*, **18**: 1015-1018.
- 4. Bars, L. J. and Bars, L. P. 1992. Fungal Contamination of Aromatic Herbs,

- Aflatoxinogenesis and Residues in Infusions. *Microbiol. Aliment. Nutr.*, **10**: 267-271.
- Bonsi, P., Augusti-Tocco, G., Palmery, M. and Giorgi, M. 1999. Aflatoxin B1 is an Inhibitor of Cyclic Nucleotide Phosphodiesterase Activity. Gen. Pharmacol., 32: 615-619.
- Chawanthayatham, S., Valentine, C. C., Fedeles, B. I., Fox, E. J., Loeb, L. A., Levine, S. S., Slocum, S. L., Wogan, G. N., Croy, R. G. and Essigmann, J. M. 2017. Mutational Spectra of Aflatoxin B1 in Vivo Establish Biomarkers of Exposure for Human Hepatocellular Carcinoma. *PNAS*, 114: E3101-E3109.
- Cho, S. H., Lee, C. H., Jang, M. R., Son, Y. W., Lee, S. M., Choi, I. S., Kim, S. H. and Kim, D. B. 2008. Aflatoxins Contamination in Spices and Processed Spice Products Commercialized in Korea. *Food Chem.*, 107: 1283-1288.
- Dhanasekaran, D., Shanmugapriya, S., Thajuddin, N. and Panneerselvam, A. 2011. Aflatoxins and Aflatoxicosis in Human and Animals. In: "Aflatoxins-Biochemistry and Molecular Biology". Ramón Gerardo Guevara-González, InTech.
- Ehrlich, K. C., Chang, P. K., Yu, J. and Cotty, P. J. 2004. Aflatoxin Biosynthesis Cluster Gene *cypA* is Required for G Aflatoxin Formation. *Appl. Environ. Microb.*, 70: 6518-6524.
- Erkmen, O. and Bozoglu, T. F. 2008. Food Microbiology. I. "Microorganisms in Foods, Microbial Growth, Foodborne Diseases and Detection of Microorganisms and Their Toxins". İlke Publishing Company, Ankara, 336 PP.
- 11. Frisvad, J. C., Skouboe, P. and Samson, R. A. 2005. Taxonomic Comparison of Three Different Groups of Aflatoxin Producers and a New Efficient Producer of Aflatoxin B1, Sterigmatocystin and 3-Omethylsterigmatocystin, Aspergillus rambellii sp. nov. Syst. Appl. Microbiol., 28: 442-453.
- 12. Frisvad, J. C., Thrane, U., Samson, R. A. and Pitt, J. I. 2006. Important Mycotoxins and the Fungi which Produce Them. *Adv. Exp. Med. Biol.*, **571**: 3-31.
- Geiser, D. M., Dorner, J. W., Horn, B. W. and Taylor, J. W. 2000. The Phylogenetics of Mycotoxin and Sclerotium Production in Aspergillus flavus and Aspergillus oryzae. Fungal Genet. Biol., 31: 169-179.

- 14. Giray, B., Girgin, G., Engin, A. B., Aydın, S. and Sahin, G. 2007. Aflatoxin Levels in Wheat Samples Consumed in Some Regions of Turkey. *Food Control*, **18**: 23-29.
- González-Salgado, A., González-Jaén, T., Vázquez, C. and Patiño, B. 2008. Highly Sensitive PCR-Based Detection Method Specific for Aspergillus flavus in Wheat Flour. Food Addit. Contam. A, 25: 758-764.
- 16. Hasanvand, H., Moshtaghi, H., Heshmati, A., Boniadian, M. and Abbasvali, M. 2016. Inhibitory Effect of *Echinophora platyloba* Essential Oil on *Aspergillus flavus* in Culture Media and Cheese. *J. Food Qual. Hazards Control*, 3: 122-127.
- 17. Heshmati, A., Ghadimi, S., Ranjbar, A. and Khaneghah, A. M. 2019. Changes in Aflatoxins Content during Processing of Pekmez as a Traditional Product of Grape. *LWT*, **103**: 178-185.
- 18. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. 2010. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Ingested Nitrate and Nitrite, and Cyanobacterial Peptide Toxins. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, 94: 1-412. https://www.ncbi.nlm.nih.gov/pubmed/2114 1240
- Karaaslan, M. and Arslanğray, Y. 2015.
   Aflatoxins B1, B2, G1, and G2
   Contamination in Ground Red Peppers
   Commercialized in Sanliurfa,
   Turkey. Environ. Monit. Assess., 187:184.
- Klich, M. A. 2002. Identification of Common Aspergillus Species, Centraalbureau voor Schimmelcultures. Utrecht, The Netherlands.
- 21. Lai, X., Zhang, H., Liu, R. and Liu, C. 2015. Potential for Aflatoxin B1 and B2 Production by *Aspergillus flavus* Strains Isolated from Rice Samples. *Saudi J. Boil. Sci.*, 22: 176-180.
- Li, R., Wang, X., Zhou, T., Yang, D., Wang, Q. and Zhou, Y. 2014. Occurrence of Four Mycotoxins in Cereal and Oil Products in Yangtze Delta Region of China and Their Food Safety Risks. Food Control, 35: 117-122.
- 23. Majeed, S., De Boevre, M., De Saeger, S., Rauf, W., Tawab, A., Rahman, M. and Iqbal, M. 2018. Multiple Mycotoxins in Rice: Occurrence and Health Risk



- Assessment in Children and Adults of Punjab, Pakistan. *Toxins*, **10**: 77.
- 24. Mashak, Z., Sohi, H. J., Heshmati, A., and Nejad, A. S. M. 2016. Assessment of AflatoxinM1 Contamination in UHT Flavored Milk Samples in Karaj. *Iran. J. Pharm. Res.*, 15: 407-411.
- Nguyen, M. T., Tozlovanu, M., Tran, T. L. and Pfohl-Leszkowicz, A. 2007. Occurrence of Aflatoxin B1, Citrinin and Ochratoxin A in Rice in Five Provinces of the Central Region of Vietnam. Food Chem., 105: 42-47
- Paterson, R. R. M. and Lima, N. 2009. Mutagens Manufactured in Fungal Culture May Affect DNA/RNA of Producing Fungi. J. App. Microbiol., 106: 1070-1080.
- Pildain, M.B., Frisvad, J.C., Vaamonde, G., Cabral, D., Varga, J. and Samson, R.A. 2008. Two Novel Aflatoxin-Producing Aspergillus Species from Argentinean Peanuts. Int. J. Syst. Evol. Micr., 58: 725-735.
- Rahman, M.A., Kang, S., Nagabhatla, N. and Macnee, R. 2017. Impacts of Temperature and Rainfall Variation on Rice Productivity in Major Ecosystems of Bangladesh. Agric. Food Secur., 6: 10.
- Reddy, K. R. N., Raghavender, C. R., Salleh, B., Reddy, C. S. and Reddy, B. N. 2011. Potential of Aflatoxin B1 Production by Aspergillus flavus Strains on Commercially Important Food Grains. Int. J. Food Sci. Technol., 46: 161-165.
- Rustom, I. Y. S. 1997. Aflatoxin in Food and Feed: Occurrence, Legislation and Inactivation by Physical Methods. Food Chem., 59: 57-67.
- Sirhan, A. Y., Tan, G. H., Al-Shunnaq, A., Abdulra'uf, L. and Wong, R. C. S. 2014. QuEChERS-HPLC Method for Aflatoxin Detection of Domestic and Imported Food in Jordan. J. Liq. Chromatogr. Relat. Technol., 37: 321–342.
- 32. Sun, X. D., Su, P. and Shan, H. 2017. Mycotoxin Contamination of Rice in China. *J. Food Sci.*, **82:** 573–584.

- 33. Tanaka, K., Sago, Y., Zheng, Y., Nakagawa, H. and Kushiro, M. 2007. Mycotoxins in Rice. *Int. J. Food Microbiol.*, **119**: 59-66.
- 34. Tournas, V. H. and Niazi, N. S. 2018. Potentially Toxigenic Fungi from Selected Grains and Grain Products. *J. Food Safety*, 38: e12422.
- 35. Towner, R. A., Hashimoto, H. and Summers, P. M. 2000. Non-Invasive *in Vivo* Magnetic Resonance Imaging Assessment of Acute Aflatoxin B1 Hepatotoxicity in Rats. *Biochim. Biophys. Acta.*, **1475**: 314-320.
- Vaamonde, G., Patriarca, A., Pinto, V. F., Comerio, R. and Degrossi, C. 2003. Variability of Aflatoxin and Cyclopiazonic Acid Production by *Aspergillus* Section *flavi* from Different Substrates in Argentina. *Int. J. Food Microbiol.*, 88: 79-84.
- 37. Villers, P. 2014. Aflatoxins and Safe Storage. *Front. Microbiol.*, **5**: 158.
- 38. Wen, J., Kong, W., Wang, J. and Yang, M. 2013. Simultaneous Determination of Four Aflatoxins and Ochratoxin A in Ginger and Related Products by HPLC with Fluorescence Detection after Immunoaffinity Column Clean-up and Postcolumn Photochemical Derivatization. *J. Sep. Sci.*, **36**: 3709-3716.
- 39. Xiao-Han, Y., Guo-Jie, X., Xin-Yue, Z., Da, L., Hong-Xia, L., Yu-Feng, S., Fan, Z. and Chun-Sheng, L. 2017. A Fluorescence Polarization Immunoassay for the Detection of Aflatoxins in Herbal Teas. *Acta Pharm. Sin.*, **52**: 620-624.
- 40. Yazdanpanah, H., Zarghi, A., Shafaati, A. R., Foroutan, S. M., Aboul-Fathi, F., Khoddam, A., Nazari, F. and Shaki, F. 2013. Analysis of Aflatoxin B1 in Iranian Foods Using HPLC and a Monolithic Column and Estimation of Its Dietary Intake. *J. Pharm. Res.*, 12: 83-89.
- Zheng, W., Teng, J., Cheng, L., Ye, Y., Pan, D., Wu, J., Xue, F., Liu, G., and Chen, W. 2016. Hetero-Enzyme-Based Two-Round Signal Amplification Strategy for Trace Detection of Aflatoxin B1 Using an Electrochemical Aptasensor. *Biosens. Bioelectron.*, 80: 570–574.

شیوع بالای افلاتاکسین  $\mathbf{B}_1$  در  $\mathbf{A}$  در انبار آلوده کننده دانه برنج درانبار

م. ب. س. الشهایب، ا. ه. البکری، ه. و. هاشیم، س. ل. الوان، ن. ب. المندیل، پ. سلواراج، ر. جرمی، س. عبدالعزیز، و ج. فرانسیس بورجیو

# چكىدە

یکی از ریزجانداران آلوده کننده برنج که به خوبی شناخته شده، Aspergillus flavus است که متابولیت های سمی به نام افلاتاکسین (AFs) تولید می کند. هدف این پژوهش بررسی پتانسیل بیوسنتز همزمان چهار AFG1 ، AFB2 ،) AFB1 AFs و AFG2 ) در سویه های آلوده کننده بود. شیوع یا فراوانی افلاتاکسین در ۱۰۹ سویه A. flavus که از دانه های برنج هندی انبار شده در ۳۰۰ مکان مختلف واقع در منطقه فرات میانی در عراق و در طی سالهای ۲۰۱۵ و ۲۰۱۶ جمع آوری شده بود بررسی شد. افلاتاکسین ها عصاره گیری شد و مقدارشان با استفاده همزمان از کروماتوگرافی مایع با كارايي بالا (HPLC) مجهز به ردياب آرايه فتو ديود (photodiode array detector) تعيين گردید. نتایج آشکار ساخت که ۲۹٪ (n=32) از سویه ها غیر-افلاتاکسیونی ( nonaflatoxigenic) و مابقی یعنی ۷۱٪ (n=77) به طور تایید شده ای افلاتا کسینی بودند که توانایی های متغييري براي توليد , mono-, bi-, و tri-Afs داشتند. همچنين پراي توليد , AFB2 AFG2, AFG1,،AFB1 به ترتیب توسط ۴۹ (۴۵٪)، ۴۴ (۴۰٪)، ۲۰ (۱۸٪)، و ۱۷ (۱۶٪) ازسویه ها در غلظت های مختلف تولید شد. در میان همه سویه های  $AFB_1$  نظت،  $AFB_1$  با میانگین  $AFB_1$  ازهمه بیشتر بود. نتیجه اینکه ،  $AFB_1$  فراوان ترین افلاتاکسین تولید شده سویه A. flavus بود که آلوده کننده برنج است. آلودگی با افلاتاکسین ها همچنان خطرات بالقوه سلامتی را برای حیوانات و انسانها ایجاد می کند. این نتایج به روشنی نشان می دهد که شرایط نامناسب انبارداری برنج در عراق برای رشد A. flavus و آلوده کرن با افلاتاکسین مساعد است. بنا براین مطالعاتی در سطح ملی برای جلوگیری از مسمومیت های ناشی از غذا واجب است. مقررات سختی باید وضع و اجرا گردد تا از تولید افلاتاکسین روی دانه های برنج جلو گیری شود .