

Genetic Evaluation of Spring Wheat (*Triticum aestivum* L.) Recombinant Inbred Lines for Spot Blotch (*Bipolaris Sorokiniana*) Resistance and Yield Components under Natural Conditions for South Asia

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ABSTRACT

The objectives of the present study were to evaluate spring wheat recombinant inbred lines (RILs) of diverse origin by estimating genetic parameters viz., variability, character association, cluster analysis, and principal component analysis (PCA) for spot blotch resistance and yield components at BHU Agricultural Research Farm during 2010-2011. Grain yield per plot was significantly and positively associated with biomass, 1,000-grain weight, harvest index, chlorophyll content, and grains per spike at genotypic level. The line 65 exhibited lowest mean of AUDPC value (632) indicating its potential as resistant parent. Cluster analysis grouped all the 324 spring wheat lines into 19 clusters using Ward's method. Extreme divergence was observed among clusters. By using D^2 -statistics, the highest inter cluster distance (584.72) was found between Clusters VIII and XIX. Cluster VIII recorded highest mean values for chlorophyll content, peduncle length, biomass, grains per spike, 1000-grain weight and grain yield. The major contributing trait towards genetic divergence was found to be AUDPC (60.36%). First 5 principal components (PC1, PC2, PC3, PC4 and PC5) accounted for proportionate values of 20.66, 17.96, 15.07, 8.28, and 7.38%, respectively, contributing 69.35% of the total variability. The second PCs had high positive PC value for plant height, biomass, and 1,000-grain weight. The breeding objectives of the present experiment was to identify genetically diverse wheat spot blotch resistant RILs for developing high yielding spot blotch resistant cultivars especially adopted to south Asia in future breeding programs.

Keywords: AUDPC, Cluster analysis, D2 analysis, PCA.

INTRODUCTION

Spot blotch disease caused by *Bipolaris sorokiniana* (sacc.) shoem syn. *Drechslera sorokiniana* (Sacc.) Subrm and Jain (syn. *Helminthosporium sativum*, teleomorph *Cochliobolus sativus*), is the most prominent disease of warmer, humid, and late-sown wheat growing regions of South Asia affecting livelihood of millions of farmers

(Saari, 1998; Joshi *et al.*, 2002). At present, spot blotch resistant potential in high yielding wheat varieties is poor and needs rigorous investigation, especially for warmer humid regions of South Asia (Sharma *et al.*, 2004; Joshi *et al.*, 2007). Several factors viz., time of sowing, sites, and moisture have adverse impact on crop yield. Severe infestation of spot blotch results in substantial yield losses ranging between 20-

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100% in South Asia by blighting of leaves and premature senescence (Duveiller and Gilchrist, 1994).

Continuous breeding results in narrow genetic diversity of the elite wheat germplasm pool and leads to problems relevant to biotic stresses, abiotic stresses, as well as adaptation (Zhang *et al.*, 2005). Maximum genetic dissimilarity among parents is essential to exploit transgressive segregation (Joshi *et al.*, 2004). Selection of genetically diverse parents upon hybridization results in higher heterosis in progenies. Therefore, there is an urgent need to exploit the existing genetic variability in wheat for evolving high yielding varieties that have wide adoptability and are highly productive under a changing climatic scenario (Baranwal *et al.*, 2012). Cluster and PC analyses are principal genetic diversity analysis tools having relative differences with each other. The cluster analysis is a robust approach for assessing family relationships (Mellingers, 1972). The objective of the present experiment was to identify genetically diverse spot blotch resistant wheat RILs for developing high yielding spot blotch resistant cultivars for South Asia.

MATERIALS AND METHODS

The present experiment was undertaken at Agricultural Research Farm, Institute of Agricultural Sciences, Banaras Hindu University (BHU), Varanasi, India, during Rabi season of 2010-11. A collection of 324 lines including 307 RILs (F8 generation) of 21 diverse crosses, seventeen distinct parents including one check i.e., Sonalika, and a highly susceptible cultivar to spot blotch disease was evaluated. Varanasi region is considered as hot spot for screening and evaluation against spot blotch disease. The genotypic set was developed by using promising spring wheat parental lines introduced from CIMMYT, Mexico and South Asia regional office, CIMMYT, Kathmandu (Nepal) including Chinese material, and other

germplasm collections. The experimental materials were sown in randomized block design with 3 replications (Table 1). The RILs were developed as per the methods described by Singh and Rajaram (1992) and Joshi *et al.* (2004). The experimental site is located in South- Eastern part of the Varanasi city at 25° 26' North latitude and 82° 99' East longitude at an elevation of 75.5 m above the mean sea level. Date of sowing was 31st of December 2010 and artificial inoculation was conducted during 23 February (4:30 pm). Growth observations were recorded for 15 yield components through random sampling method. Data on five plants of each line was averaged and mean data was used for statistical analysis. Agronomic practices recommended for irrigated and normal fertile soil (120 kg N; 60 kg P₂O₅ and 40 kg K₂O ha⁻¹) were followed to raise a good crop.

Area under Disease Progress Curve (AUDPC)

Spot blotch disease was recorded at different growth stages (Zadoks *et al.*, 1974). Disease severity (%) was recorded at different stages to calculate AUDPC (Van der Plank, 1963; Roelfs *et al.* 1992) using the following formula:

$$AUDPC = \sum_{i=1}^n \left[\left\{ \frac{Y_i + Y_{(i+1)}}{2} \right\} (t_{i+1} - t_i) \right]$$

Where, Y_i is the disease level at time t_i and $t_{(i+1)} - t_i$ the time (days) between two disease scores and n is the number of dates on which spot blotch was recorded.

Statistical Analysis

Character association was calculated following Robinson *et al.* (1951). Genetic divergence among different lines was assessed based on the estimated *inter-se* genetic distances among the lines using D^2 -statistics of Mahalanobis (1928), which is one of the most effective tools to measure

Table1. Collection of 324 spring wheat lines representing source of spot blotch resistance.

Line No.	Lines / Parents	Crosses/Details
1 – 9	Spot blotch RILs – 1 (9)	Yangmai-6×Monsu kunjer
10 – 25	Spot blotch RILs – 2 (16)	Monalds×Bonly
26 – 52	Spot blotch RILs – 3 (27)	Yangmai-6×Monalds
53 – 69	Spot blotch RILs – 4 (17)	Monalds×Songhi
70 – 86	Spot blotch RILs – 5 (17)	Songhi-4×IA 814-467
87 – 103	Spot blotch RILs – 6 (17)	Chirya 7×Tink
104 – 131	Spot blotch RILs – 7 (28)	Yangmai-6×Chirya-7
132 – 153	Spot blotch RILs – 8 (22)	Yangmai-6×Tink
154 – 168	Spot blotch RILs – 9 (15)	Songhi-4×BR-8-471
169 – 178	Spot blotch RILs – 10 (10)	Songhi-4×Chirya-7
179 – 198	Spot blotch RILs – 11 (20)	Monalds×Yangmai-6
199 – 210	Spot blotch RILs – 12 (12)	Yangmai-6×Ning-8027
211 – 224	Spot blotch RILs – 13 (14)	Songhi-4×Bonly
225 – 240	Spot blotch RILs – 14 (16)	IA-814-867×Tink
241 – 255	Spot blotch RILs – 15 (15)	Suzo-8×Monalds
256 – 275	Spot blotch RILs – 16 (20)	Monalds×Chirya-7
276 – 284	Spot blotch RILs – 17 (9)	Monalds×Ning-8127
285 – 304	Spot blotch RILs – 18 (20)	Songhi-4×Ning-8119
305	Spot blotch RILs – 19 (1)	Chirya-7×Longmai-10
306	Spot blotch RILs – 20 (1)	Songhi-4×Monsu Kunjer
307	Spot blotch RILs – 21 (1)	Monald×Ning- 8119
308	Parent - 1	Sonalika (Highly susceptible variety)
309	Parent - 2	HUW-234
310	Parent - 3	HUW-468
311	Parent - 4	HUW-510
312	Parent - 5	Sonalika
313	Parent - 6	IA-814-877
314	Parent - 7	Chirya-7
315	Parent - 8	Ning-8119
316	Parent - 9	Monalds
317	Parent - 10	Monsu Kunjer
318	Parent - 11	Tink
319	Parent - 12	Bonaly
320	Parent - 13	Songhi-4
321	Parent - 14	Suzo-8
322	Parent - 15	songhi-4
323	Parent - 16	Yangmai-6
324	Parent - 17	Lok-1

the genetic distance between lines as measured by allelic frequencies at a sample of loci. After arranging the D^2 -values of all combinations of one genotype with the others in ascending order of magnitudes, the lines were grouped into a number of clusters by Ward's method described by Rao (1952). The inter- and intra-cluster distances were calculated and their relationships were diagrammatically represented. The Statistical Analyses Software (SAS) and STATISTICA ver.10 were utilized for statistical analysis.

RESULTS AND DISCUSSION

In the present experiment, 324 spring wheat lines of diverse origin were analyzed for genetic parameters viz., character association, cluster analysis, and principal component analysis (PCA) for spot blotch resistant and yield components. The best ten lines based on the mean performance of few promising traits in desirable direction are represented in Table 2. The lines 239 and 308 showed lesser days to 50% flowering

**Table 2.** Promising ten lines for each of the promising traits under study.

Traits	1	2	3	4	5	6	7	8	9	10
DF (Early) ^a	Line 239 (60.33) ^b	Line 308 (60.33)	Line 240 (61.33)	Line 110 (62.00)	Line 243 (62.33)	Line 287 (62.33)	Line 215 (63.00)	Line 237 (63.00)	Line 282 (63.33)	Line 288 (63.33)
DPM1 (Early) ^b	Line 300 (93.33)	Line 302 (93.33)	Line 110 (94.00)	Line 261 (94.33)	Line 264 (94.33)	Line 270 (94.33)	Line 276 (94.33)	Line 280 (94.33)	Line 299 (94.33)	Line 111 (95.00)
AUDPC (Low) ^c	Line 65 (632.00)	Line 1 (644.32)	Line 41 (682.61)	Line 44 (744.34)	Line 91 (744.34)	Line 97 (744.34)	Line 98 (761.63)	Line 108 (762.86)	Line 92 (765.95)	Line 107 (778.91)
PH (cm) (Dwarf) ^d	Line 173 (48.33)	Line 171 (51.33)	Line 174 (55.83)	Line 172 (56.33)	Line 128 (58.33)	Line 176 (61.83)	Line 177 (61.83)	Line 175 (62.33)	Line 224 (63.33)	Line 109 (64.33)
BM (g) (High) ^e	Line 54 (232.50)	Line 107 (232.50)	Line 96 (217.50)	Line 46 (207.50)	Line 97 (207.50)	Line 108 (207.50)	Line 188 (203.33)	Line 9 (202.50)	Line 65 (192.50)	Line 249 (189.17)
GY (g) (High) ^f	Line 108 (69.60)	Line 41 (69.10)	Line 304 (66.17)	Line 107 (66.00)	Line 43 (65.60)	Line 258 (64.87)	Line 46 (64.60)	Line 78 (64.50)	Line 263 (63.97)	Line 54 (63.90)
TGW (g) (High) ^g	Line 12 (41.34)	Line 54 (40.74)	Line 34 (38.94)	Line 46 (38.94)	Line 296 (37.68)	Line 25 (37.54)	Line 175 (37.38)	Line 13 (36.94)	Line 207 (36.18)	Line 22 (36.14)
HI (%) (High) ^h	Line 78 (55.88)	Line 76 (54.08)	Line 41 (48.18)	Line 81 (47.68)	Line 43 (45.38)	Line 26 (45.18)	Line 85 (45.18)	Line 146 (42.65)	Line 47 (42.18)	Line 170 (41.95)

^a Days to 50% Flowering (Days); ^b Days to Physiological Maturity (Days); ^c Area Under Disease Progress Curve; ^d Plant Height (cm); ^e BioMass (g); ^f Grain Yield (g); ^g Thousand Grain Weight (g), and ^h Harvest Index (%). Bracketed values indicate the mean performance of the corresponding lines and bold line numbers are common for more than one trait.

(60.33 days) indicating early maturity. The line 65 exhibited lowest AUDPC value (632) indicating a resistant parent in consonance with Sharma *et al.* (1997). Low yield of few lines was indicating high susceptibility to spot blotch disease (Phadnawis *et al.*, 2002). Highest 1,000-grain weight was observed in the line 12 (41.34 g). The highest harvest index was observed in line 78 (55.88%) (Table 2). The observations suggested vast differences among the RILs in terms of spot blotch resistant and yield components.

Grain yield per plot was significantly and positively associated with biomass, 1000-grain weight, harvest index, chlorophyll content, and grains per spike at genotypic level (Table 3) as reported by Khodarahmpour *et al.* (2011) and Olfati *et al.* (2010). It suggests that the characters should be included in phenotyping for genetic improvement for wheat genotypes. Negatively significant correlation was observed between yield and AUDPC, indicating that spot blotch was the major problem for wheat yield at phenotypic level (Table 5) as reported by Gilchrist and Pfeiffer (1991), but non-significant (0.23) at

genotypic level, indicating major role of environment for the disease incidence. AUDPC showed negative and significant association with biomass, sheath length, days to physical maturity, chlorophyll content, days to physiological maturity, and days to 50% flowering at genotypic level (Table 3).

Genetic Divergence Analysis

All the 324 wheat lines were grouped into 19 clusters through Ward's method (Table 4; Figure 1). By using D^2 -statistics of Mahalanobis (1928), the D values (cluster distance) of this genotypic collection (square of D value represents D^2 value) are represented in Table 5 (Figure 2). In this method, highest intra cluster distance was found for Cluster VIII (53.07) followed by XIV. Similarly, lowest intra cluster distances have been obtained for Cluster I (32.16). The highest inter cluster distance (584.72) through this method has been found between Clusters VIII and XIX (Table 5).

Table 3. Character association between yield and its components for the 324 spring wheat lines.^a

Characters	CL	DPM1	DPM2	AUDPC	SL	PL	EL	PH	BM	GY	TGW	GPS	HI	SCM
DF	r_p 0.22*	0.67**	0.72**	-0.47**	0.13*	-0.23*	-0.04	-0.04	0.00	-0.30**	-0.30**	0.05	-0.38**	0.08
	r_g 0.19*	0.79**	0.62**	-0.35**	0.68**	-0.12*	0.09	0.14*	0.24*	-0.35**	-0.46**	0.97**	-0.67**	0.34**
CL		0.20*	0.17*	-0.36**	0.10	-0.08	0.01	-0.04	0.26**	0.18*	0.20*	0.17*	-0.05	0.06
	r_g	0.03	0.21*	-0.42**	0.43**	0.12*	0.29**	0.20*	0.54**	0.56**	0.23*	0.55**	-0.36**	-0.16*
DPM1			0.90**	-0.34**	0.09	-0.13*	-0.01	0.32**	-0.09	0.39**	-0.05	-0.08	-0.16*	0.16*
	r_g		0.66**	-0.37**	0.92**	0.28**	0.10	0.31**	0.01	-0.11*	-0.14*	0.92**	-0.21*	0.27**
DPM2				-0.33**	0.10	-0.12*	-0.02	0.32**	-0.09	0.34**	-0.10	-0.07	-0.22*	0.17*
	r_g			-0.50**	0.92**	0.39**	0.53**	0.44**	0.16	-0.27**	-0.25*	-0.63**	-0.51**	0.11*
AUDPC					-0.17*	-0.16*	-0.04	-0.01	-0.32**	-0.45**	-0.15*	-0.11*	-0.10	-0.09
	r_g				-0.77**	0.53**	-0.22*	-0.06	-0.82**	-0.23*	0.02	-0.47**	0.78**	-0.75**
SL						0.54**	0.31**	0.56**	0.26**	0.08	0.06	0.15*	-0.17*	0.10
	r_p					-0.60**	-0.19*	0.10	0.38**	-0.50**	0.10	-0.57**	-0.91**	0.00
	r_g						0.26**	0.77**	0.11*	0.00	0.04	0.00	-0.07	0.10
PL							-0.22*	0.78**	0.38**	-0.42**	0.02	-0.83**	-0.72**	0.29**
	r_p							0.34**	0.13*	0.07	0.09	-0.06	-0.05	0.14*
	r_g							0.22*	-0.01	-0.06	0.11*	0.43**	-0.14*	0.28**
EL									0.23*	0.01	0.14*	-0.01	-0.19*	0.14*
PH									0.41**	-0.14*	0.17*	-0.24*	-0.45*	0.35**
	r_p									0.59**	0.34**	0.32**	-0.31**	0.09
BM										0.85**	0.64**	-0.43**	0.11	-0.02
	r_g										0.66**	0.23*	0.56**	0.07
GY											0.78**	0.46**	0.66**	-0.04
	r_p											-0.09	0.41**	0.11*
TGW												-0.59**	0.51**	0.26**
	r_g												-0.06	-0.08
GPS													0.14*	-0.78**
	r_p													0.01
HI														0.04
	r_g													

*** Significant at 5 and 1% level of significance.

^a rp: Phenotypic correlation coefficient; rg= Genotypic correlation coefficient, (SCM): Seed Colour Mean, and Abbreviations: As in Table 2.

Table 4. Ward's clustering pattern (D²-statistics) representing 19 clusters for 324 spring wheat lines.

Cluster	Cluster members																															Total no. of lines	
I	1	2	3	4	5	14	102	111	121	157	201	203	204	246	248	289	317															17	
II	9	10	12	25	27	29	32	66	87	88	90	112	180	189	191	194	197	210	244	254	255	261	263	272	284	305							26
III	11	70	122	124	148	171	173	175	176	177	179	190	193	219	225	226	230	233	262	316	323											21	
IV	8	16	20	34	35	45	55	62	93	95	99	100	104	105	127	142	147	149	188	249	264	267	275									22	
V	19	24	132	133	134	136	137	143	153	174	181	183	186	218	265																	14	
VI	7	36	46	60	63	89	94	96	101	106	145	150	268																			12	
VII	6	21	44	129	146	151	198	205																								8	
VIII	13	41	54	65	91	92	97	98	107	108	144																					11	
IX	15	38	76	81	85	109	128	135	152	156	163	168	184	216	227	270	295	319														17	
X	18	30	42	48	49	117	118	119	120	164	182	185	192	199	202	209	234	245	250	253	266	274	276	278	280	283	285	286	306	314		30	
XI	86	113	139	158	235	236	251	256	257	303	307	310	315																			13	
XII	17	22	23	28	33	40	47	67	68	77	123	131	138	141	159	170	178	217	220	229	231	259	260	279	318							25	
XIII	43	103	125	126	130	140	208	240	247	269	273	277	281	321																		14	
XIV	26	31	72	73	74	79	115	116	169	172	206	207	214	223	271	291	292	293	298	302												20	
XV	50	53	61	64	69	78	84	110	114	224	228	239	241	242	252	258	282	313														17	
XVI	37	39	51	52	56	57	80	154	162	187	195	196	222	232	237	296	297	299	300	301	309											25	
XVII	58	71	82	155	160	161	166	211	213	221	287	288	294	311	312	324																16	
XVIII	59	75	83	200	212	215	243	290	304	320																						11	
XIX	165	167	238	308	322																											5	

Table 5. Average intra and inter-cluster *D* values (Ward's method) among 19 clusters of 324 spring wheat lines.^a

Clusters	I	II	III	IV	V	VI	VII	VIII	IX	X	XI	XII	XIII	XIV	XV	XVI	XVII	XVIII	XIX
I	32.18	46.79	55.99	49.73	61.43	83.24	99.70	142.02	135.00	100.93	128.12	80.37	72.01	213.68	171.38	263.97	372.96	319.06	453.01
II		39.30	61.70	71.19	88.57	109.02	128.97	168.48	110.57	75.78	100.50	60.57	49.80	187.22	143.89	235.78	344.90	290.75	424.33
III			46.52	81.27	73.69	114.91	120.07	169.69	120.41	95.04	126.61	70.32	83.04	197.69	161.25	251.49	357.79	305.30	439.59
IV				37.06	59.88	56.23	77.97	111.28	167.23	131.42	156.02	111.85	95.95	245.36	201.90	294.40	404.17	349.68	483.34
V					38.49	70.43	64.89	116.29	175.45	145.25	175.45	120.17	119.75	254.90	215.77	308.13	415.77	362.69	497.11
VI						37.24	55.25	74.86	208.58	172.45	196.28	152.64	135.28	287.15	243.19	335.85	445.78	391.24	524.51
VII							37.46	72.86	224.96	191.86	219.98	168.06	159.78	304.80	263.70	356.95	465.77	412.01	546.42
VIII								53.07	268.54	232.70	256.54	212.03	194.99	347.14	303.52	396.14	506.14	451.44	584.72
IX									41.37	53.99	58.58	68.44	90.63	92.08	60.61	141.39	246.65	194.16	327.75
X										33.73	48.78	44.48	54.80	122.94	80.85	170.38	278.76	224.84	358.57
XI											35.17	72.81	71.24	107.58	62.51	146.33	255.06	200.67	332.43
XII												33.96	53.42	143.74	104.91	195.10	302.68	249.28	383.69
XIII													33.72	162.04	116.60	206.92	316.57	261.93	394.71
XIV														51.52	66.01	75.32	170.39	120.52	250.86
XV															39.73	102.02	208.19	154.71	287.11
XVI																41.61	118.81	69.54	195.09
XVII																	47.11	71.05	95.63
XVIII																		44.19	142.47
XIX																			49.72

^a Figures in diagonal indicate intra-cluster distance.

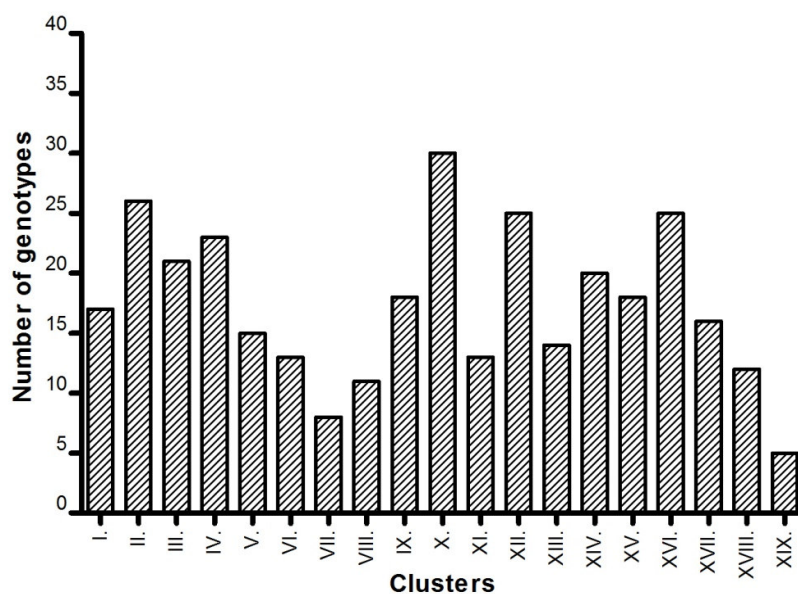


Figure 1. Graphical representation of 324 spring wheat lines into 19 clusters and number of respective genotypes within each cluster.

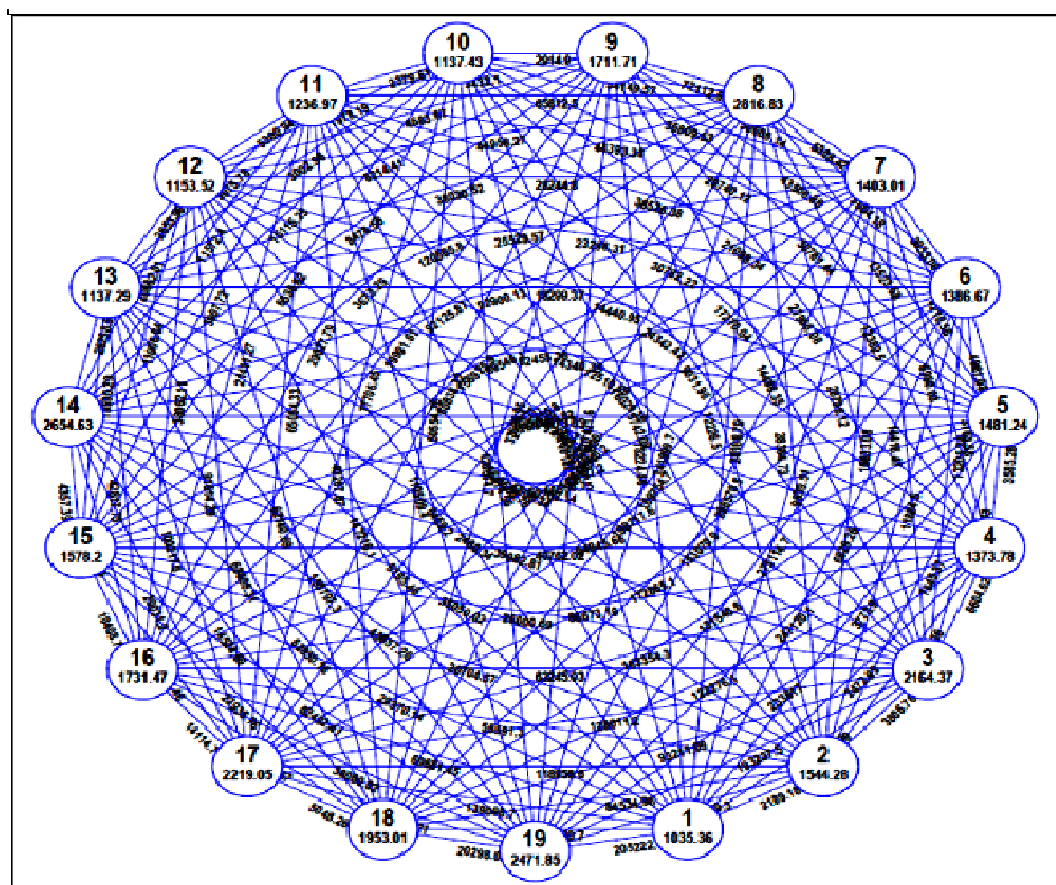


Figure 2. Ward's clustering pattern representing 19 clusters of D^2 -statistics for 324 spring wheat lines.



Hybridization between these cluster members revealed transgressive segregation for effective selection. Clusters VI, VII, VIII, XI, XII and XIX exhibited highest cluster mean values for most of the traits. Highest mean values for ear length and plant height are found in Cluster VI (Table 6). Cluster VII expressed highest values for days to 50% flowering, days to physical maturity, and seed colour mean. Cluster VIII recorded highest mean values for chlorophyll content, peduncle length, biomass, grains per spike, 1,000-grain weight, and grain yield as reported by Khodadadi *et al.* (2011). Highest cluster mean values for sheath length, harvest index and AUDPC have been exhibited by Clusters XI, XII and XIX, respectively (Table 6). The major contributing traits towards genetic divergence was found to be AUDPC (60.36%) followed by biomass (5.96%), plant height (0.58%) and grain yield (0.54%) similar to that reported by Goel *et al.* (2005) (Table 6). In the present experiment, AUDPC and biomass played major role in clustering wheat lines (Table 6; Figure 3-A).

Principal Components Analysis

PC analysis revealed the largest contributor to the total variation at each axis of differentiation. Seven PCs (PC1 to PC7) were considered from the original data explaining 79.85% of the total variation (Table.7) similar to that reported by Hailegiorgis *et al.* (2011),

Ravishanker *et al.* (2013), and Caliskan and Bayazit (2013). Out of the total 7 PCs, 5 principal components (PC1, PC2, PC3, PC4 and PC5) accounted with proportionate variance values of 20.66, 17.96, 15.07, 8.28, and 7.38%, respectively, and contributed 69.33 % of the cumulative variation having Eigen value greater than one (Table 7). Two dimensional ordinations of 324 spring wheat lines on PC axes 1 and 2 are represented for separation of the lines which reveal existence of extreme variability in the present wheat genotypic set (Figure 3-B). The first principal component has high positive component value for days to 50% flowering, chlorophyll content, days to physical maturity, and days to physiological maturity. PC1 has negative component value for AUDPC, 1,000-grain weight, grain yield, and harvest index. The second principal component had high positive component value for plant height, biomass, peduncle length, and sheath length and high negative component value for AUDPC. The abovementioned traits having high positive or negative component value reveal more genetic diversity and they play tremendous role in representing the clusters. The third principal component had high positive component value for grain yield, 1,000-grain weight, and harvest index and high negative component value for AUDPC, sheath length and peduncle length (Table 7) similar to that reported by Hailegiorgis *et al.* (2011). The projection of component traits on PC1 and PC2 revealed

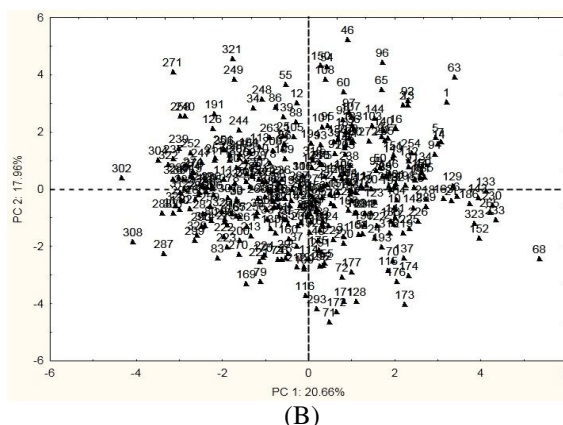
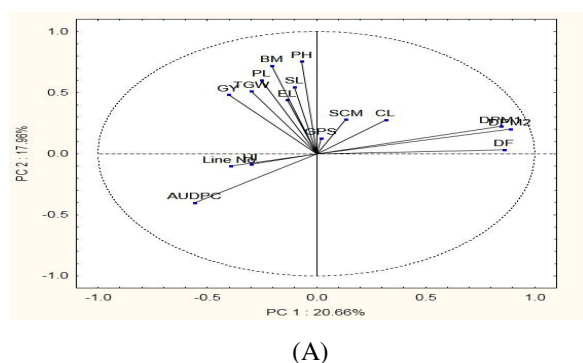


Figure 3- A) Principal component biplot for yield components in the spring wheat lines , B) Scattered diagram of the first two principal components for yield components in the spring wheat lines.

Table 6. Cluster mean values (Ward's method) and contribution of traits toward divergence for the studied traits in spring wheat lines.^a

Cluster	DF	CL	DPM1	DPM2	AUDPC	SL	PL	EL	PH	BM	GY	TGW	GPS	HI	SCM
I.	73.94	45.62	101.82	107.23	998.55	20.03	34.08	10.35	93.10	144.95	39.31	26.09	36.19	27.37	104.33
II.	71.12	46.96	99.73	105.34	1063.43	19.88	33.41	10.28	88.54	154.39	43.23	28.34	39.94	28.29	104.12
III.	75.86	47.91	102.00	107.13	1019.44	18.23	27.87	9.37	73.52	105.95	30.66	24.46	35.83	29.13	99.99
IV.	70.94	47.24	99.03	105.04	943.67	19.43	32.95	10.50	89.20	167.10	46.10	29.31	39.76	28.23	100.02
V.	79.00	46.62	102.27	108.37	899.44	19.75	31.23	9.20	84.50	114.67	29.43	23.20	40.76	26.30	104.55
VI.	75.00	47.87	102.77	108.07	869.72	19.80	34.62	11.10	97.79	178.14	45.86	30.47	41.41	25.38	103.01
VII.	79.13	46.40	103.50	108.64	809.39	19.07	31.68	9.59	83.90	138.65	40.71	26.29	38.09	28.95	106.05
VIII.	73.15	48.03	103.24	108.00	752.48	19.66	36.03	9.51	95.56	187.58	54.36	31.57	45.20	29.44	100.44
IX.	71.93	45.82	99.54	105.28	1251.17	18.16	32.39	9.59	80.94	119.40	33.55	23.10	39.83	30.48	100.08
X.	70.43	44.39	98.93	104.24	1193.83	19.39	32.88	10.05	84.95	144.25	43.32	27.45	39.08	30.09	102.90
XI.	68.90	45.98	98.13	103.02	1258.90	20.46	34.26	10.09	89.26	173.85	46.01	27.91	38.83	26.69	104.23
XII.	71.01	45.46	100.13	105.27	1136.84	18.50	31.42	9.48	81.13	125.47	38.79	25.94	39.87	31.82	99.72
XIII.	68.69	46.77	98.33	103.71	1132.81	19.77	34.13	10.81	90.90	173.27	50.78	31.32	37.49	29.62	101.31
XIV.	69.77	42.93	98.07	103.30	1411.80	18.28	31.39	8.96	79.33	114.38	34.73	24.93	37.54	30.58	96.27
XV.	68.48	44.03	97.98	103.56	1341.92	18.58	34.26	10.39	87.28	145.09	42.10	26.10	40.93	29.59	102.99
XVI.	68.20	44.21	97.39	103.17	1536.34	18.90	34.45	9.74	86.36	145.33	39.66	26.71	38.55	27.73	100.41
XVII.	68.08	43.73	97.71	103.96	1754.28	19.15	34.66	9.89	84.46	123.85	33.25	24.52	34.66	26.96	100.91
XVIII.	65.67	40.54	97.53	102.99	1645.82	18.88	33.69	10.47	84.44	140.21	40.07	24.40	41.22	29.51	100.10
XIX.	68.40	43.30	98.40	103.49	1937.86	19.84	35.86	10.01	92.23	154.83	36.59	25.36	38.06	24.24	103.62
% contribution of traits towards divergence															
0.01 0.02 0.00 0.00 0.00 0.00 0.01 0.00 0.00 0.58 5.96 0.54 0.02 0.27 0.04 0.03															

^a Abbreviations: As in Table 2

**Table 7.** Principal Component analysis (PCA) for spot blotch resistant and yield components in the spring wheat lines.

Traits	PC1	PC2	PC3	PC4	PC5	PC6	PC7
DF	0.47	0.02	-0.11	0.14	0.15	-0.06	0.13
CL	0.17	0.16	0.17	0.37	0.03	0.16	-0.26
DPM1	0.46	0.13	0.08	-0.10	0.10	-0.16	0.13
DPM2	0.49	0.12	0.08	-0.14	-0.01	-0.09	0.12
AUDPC	-0.31	-0.24	-0.19	-0.11	-0.02	0.06	-0.02
SL	-0.06	0.32	-0.34	0.18	0.27	-0.13	0.32
PL	-0.14	0.35	-0.29	-0.26	-0.27	0.16	0.20
EL	-0.07	0.26	-0.18	-0.09	0.32	-0.07	0.06
PH	-0.04	0.45	-0.25	-0.23	-0.22	0.03	0.07
BM	-0.11	0.43	0.11	0.24	-0.15	-0.18	-0.38
GY	-0.22	0.28	0.43	0.09	0.09	-0.05	0.17
TGW	-0.16	0.30	0.40	-0.09	0.09	-0.20	-0.15
GPS	0.01	0.07	0.00	0.61	-0.32	0.44	0.34
HI	-0.16	-0.05	0.45	-0.16	0.21	0.16	0.57
SCM	0.07	0.16	-0.01	-0.19	0.46	0.74	-0.31
Eigen value	3.31	2.87	2.41	1.32	1.18	0.85	0.83
Cumulative Eigen value	3.31	6.18	8.59	9.91	11.09	11.95	12.78
Individual variance (%)	20.66	17.96	15.07	8.28	7.38	5.34	5.18
Cumulative variance (%)	20.66	38.61	53.68	61.96	69.33	74.67	79.85

^a Abbreviations: As in Table 2.

that the ear length, 1,000-grain weight, biomass, and harvest index were positively associated with grain yield (Figure 3-A).

CONCLUSIONS

The present experiment was conducted to identify spot blotch resistant RILs with better yield potential for south Asia. Based on character association study, grain yield had strong positive association with chlorophyll content, 1,000-grain weight, biomass and harvest index and negative association with the disease. The present experimental materials revealed extreme genetic variability. The materials were classified under 19 clusters and its major proportion of variance depicted by principle components. The promising RILs will be evaluated under multi-location trial for their location suitability. Genetically diverse promising RILs will be exploited as potent donor against spot blotch disease.

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REFERENCES

1. Baranwal, D. K., Mishra, V. K., Vishwakarma, M. K., Yadav, P. S. and Arun, B. 2012. Studies on Genetic Variability, Correlation and Path Analysis for Yield and Yield Contributing Traits in Wheat (*T. aestivum* L. em Thell.). *Plant Arch.*, **12**: 99-104.
2. Caliskan, O. and Bayazit, S. 2013. Morphological and Chemical Diversity of Pomegranate Accessions Grown in Eastern Mediterranean Region of Turkey. *J. Agr. Sci. Tech.*, **15**: 1449-1460.
3. Duveiller, E. and Gilchrist, L. 1994. Productions Constraints Due to *Bipolaris*

- sorokiniana* in Wheat: Current Situation and Future Prospects. In: "Wheat in Heat-stressed Environments: Irrigated, Dry Areas and Rice-Wheat Farming Systems", (Eds.): Saunders, D. A. and Hettel, G. P.. ISBN-13: 9789686127874, UNDP/ARC/BARI/CIMMYT, DF, Mexico, PP: 343-352.
4. Gilchrist, L. I. and Pfeiffer, W. H. 1991. Resistance to *Helminthosporium sativum* in Bread Wheat: Relationship of Infected Plant Parts and Their Association of Agronomic Traits. In: "Wheat for the Non-traditional Warm Areas", (Ed.): Saunders, D. A.. CIMMYT, El Batán, DF, Mexico, PP.1-549.
 5. Goel, P., Pal, S. S. and Jaiswal, J. P. 2005. Genetic Analysis for Spot Blotch (*Bipolaris sorokiniana* Sacc. in Borok. Shoem.) Reaction in Wheat [*Triticum aestivum* (L.) Em. Thell.]. *Indian J. Genet. Plant Breed.*, **65**: 305 -306.
 6. Hailegiorgis, D., Mesfin, M. and Genet, T. 2011. Genetic Divergence Analysis on Some Bread Wheat Genotypes Grown in Ethiopia. *J. Central Eur. Agric.*, **12**: 344 -352.
 7. Joshi, A. K., Chand, R. and Arun, B. 2002. Relationship of Plant Height and Days to Maturity with Resistance to Spot Blotch in Wheat. *Euphytica*, **123**: 221 -228.
 8. Joshi, A. K., Kumar, S. Chand, R. and Ortiz-Ferrara, G. 2004. Inheritance of Resistance to Spot Blotch Caused by *Bipolaris sorokiniana* in Spring Wheat. *Plant Breed.*, **123**: 213 -219.
 9. Joshi, A. K., Ortiz-Ferrara, G. Crossa, J. Singh, G. and Alvarado, G. *et al.*, 2007. Associations of Environments in South Asia Based on Spot Blotch Disease of Wheat Caused by *Cochliobolus sativus*. *Crop Sci.*, **47**: 1071 -1081.
 10. Khodadadi, M., Fotokian M. H. and Miransari, M. 2011. Genetic Diversity of Wheat (*Triticum aestivum* L.) Genotypes Based on Cluster and Principal Component Analyses for Breeding Strategies. *Aust. J. Crop Sci.*, **5**: 17 -24.
 11. Khodarahmpour, Z., Choukan, R., Bihanta, M. R. and Hervan, E. M. 2011. Determination of the Best Heat Stress Tolerance Indices in Maize (*Zea mays* L.) Inbred Lines and Hybrids under Khuzestan Province Conditions. *J. Agr. Sci. Tech.*, **13**: 111-121.
 12. Mahalanobis, P. C. 1928. A Statistical Study at Chinese Head Measurement. *J. Asiatic Soc. Bengal*, **25**: 301 -377.
 13. Mellingers, J. S., 1972. Measures of Genetic Similarity and Genetic Distance. VII. In: "Studies in Genetics", (Ed.): Wheeler, M. R.. Publication No. 7213, University of Texas, Austin, PP. 145-153.
 14. Olfati, J. A., Peyvast, G., Shabani, H. and Nosratie-Rad, Z. 2010. An Estimation of Individual Leaf Area in Cabbage and Broccoli Using Non-destructive Methods. *J. Agr. Sci. Tech.*, **12**: 627-632.
 15. Phadnawis, B. N., Khatod, J. P., Vitkare, D. G., Shivankar, R. S. and Nagone, A. H. 2002. Genetic Variability and Correlation Coefficient Studies in Durum Wheat. *Ann. Plant Physiol.*, **6(2)**: 115-118.
 16. Rao, C. R. 1952. *Advanced Statistical Methods in Biometrical Research*. John Wiley and Sons, New York, PP. 361-367.
 17. Ravishanker, Kumar, S., Baranwal, D. K., Chatterjee, A. and Solankey, S. S. 2013. Genetic Diversity Based on Cluster and Principal Component Analyses for Yield and Quality Attributes in Ginger (*Zingiber officinale* Roscoe). *Int. J. Plant Breed. Gene.*, DOI:10.3923/ijpbg.
 18. Robinson, H. F., Comstock, R. E. and Harvey, P. H. 1955. Genotypic and Phenotypic Correlation's in Wheat and Their Implications in Selection. *Agrono. Jour.*, **43**: 282-287.
 19. Roelfs, A. P., Singh, R. P. and Saari, E. E. 1992. *Rust Diseases of Wheat: Concepts and Methods of Disease Management*. CIMMYT, Mexico, USA, PP: 37-38.
 20. Saari, E. E. 1998. Leaf Blight Diseases and Associated Soil Borne Fungal Pathogens of Wheat in North and South East Asia. In: "Helminthosporium Blights of Wheat: Spot Blotch and Tan Spot", (Eds.): Duveiller, E., Dubin, H. J., Reeves, J. and McNab, A.. CIMMYT, Mexico, USA, PP: 37-51.
 21. Sharma, R. C., Dubin, H. J., Bhatta, M. R. and Devkota, R. N. 1997. Selection for Spot Blotch Resistance in Four Spring Wheat Populations. *Crop Sci.*, **37(2)**: 432-435.
 22. Sharma R.C., Duveiller, E., Ahmed, F., Arun, B., Bhandari, D., Bhatta, M.R., Chand, R., Chaurasiya, P.C.P., Gharti, B., Hossain, M.H., Joshi, A.K., Mahto, B.N., Malaker, P.K., Reza, M.A., Rahman, M., Samad, M.A., Shaheed, M.A., Siddique, A.B., Singh, A.K., Singh, K.P., Singh, R.N. and Singh. S.P., 2004. Helminthosporium Leaf Blight Resistance and Agronomic Performance of Wheat Genotypes Across Warm Regions of South Asia. *Plant Breed.*, **123**: 520 -524.
 23. Singh, R. P. and Rajaram, S. 1992. Genetics of Adult-plant Resistance of Leaf Rust in



- 'Frontana' and Three CIMMYT Wheats. *Genome.*, **35**: 24-31.
24. Van der Plank, J. E. 1963. *Plant Diseases: Epidemics and Control*. 1st Edition, ISBN: 0127114505, Academic Press, New York, 349 PP.
25. Zadoks, J. C., Chang, T. T. and Konzak, C. F. 1974. A Decimal Code for the Growth Stages of Cereals. *Weed Res.*, **14**: 415-421.
26. Zhang, P., Dreisigacker, S., Melchinger, A.E., Reif, J.C., Ginkel, M. V., Kazi, A., Hoisington, D. and Warburton, M.L. 2005. Quantifying Novel Sequence Variation and Selective Advantage in Synthetic Hexaploid Wheats and Their Backcross-derived Lines Using SSR Markers. *Mol. Breed.*, **15**: 1-10.

ارزیابی ژنتیکی رگه های خویش آمیخته و نو ترکیب گندم بهاره (*Triticum aestivum* L.) برای مقاومت به Spot Blotch (*Bipolaris Sorokiniana*) و ارزیابی اجزای عملکرد گندم در شرایط طبیعی جنوب آسیا

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چکیده

هدف این پژوهش ارزیابی ژنتیکی رگه های خویش آمیخته و نو ترکیب گندم بهاره (RILs) با مبداء های متنوع بود و برای این منظور پارامترهای ژنتیکی شامل تغییرپذیری، همراهی شاخص ها، تجزیه خوشه ای، و تجزیه به مولفه های اصلی (PCA) برای مقاومت به Spot Blotch، و اجزای عملکرد در مزرعه تحقیقاتی دانشگاه هندوی بناراس طی ۱۱-۲۰۱۰ ارزیابی شد. عملکرد دانه در هر کرت به گونه ای معنی دار و مثبت با زیست توده، وزن هزار دانه، شاخص برداشت، محتوی کلروفیل، و تعداد دانه در خوشه در سطح ژنوتیپی همراهی (رابطه) داشت. رگه ۶۵ کمترین مقدار میانگین "مساحت زیرمنحنی پیشرفت مرض" (AUDPC) را که برابر ۶۳۲ بود نشان داد و این نشانگر استعداد و پتانسیل این رگه به عنوان والد مقاوم بود. تجزیه خوشه ای داده ها با استفاده از روش Ward، همه ۳۲۴ رگه گندم را در ۱۹ خوشه دسته بندی کرد. واگرایی شدیدی بین خوشه ها مشاهده شد. با استفاده از آماره D^2 بیشترین فاصله بین خوشه ای (۵۸۴/۷۲) بین خوشه VIII و XIX به دست آمد. بیشترین میانگین برای کلروفیل II و طول ساق گل، زیست توده، دانه در خوشه، وزن هزار دانه و عملکرد دانه برای خوشه VIII ثبت شد. صفت اصلی که به واگرایی ژنتیکی کمک می کرد AUDPC بود (60.36%). پنج مولفه اصلی (PC1, PC2, PC3, PC4, PC5) ارزش متناسبی برابر ۶۶/۲۰٪، ۱۵/۰۷٪، ۱۷/۹۶٪، ۸/۲۸٪، ۷/۳۸٪ داشتند و مجموعاً ۶۹/۳۵٪ از تغییرات را توجیه میکردند. مولفه های اصلی دوم مقادیر مثبت بالایی برای طول گیاه، زیست توده و وزن هزار دانه داشتند. هدف بهنژادی در این پژوهش شناسایی رگه های گندم مقاوم به Spot Blotch با تنوع ژنتیکی متنوع بود تا بتوان در برنامه های آینده بهنژادی کولتیوارهای پر محصول که به ویژه با جنوب آسیا سازگارند به وجود آورد.