

Root Architectural Diversity in Wheat Cultivars and Its Role in Response to Terminal Water Stress

Afshin Zamani¹, Yahya Emam^{1*}, and Aref Nowrouzi¹

Abstract

Root architecture plays a key role in optimizing water uptake and sustaining yield under drought stress. The aim of this study was to investigate the role of vertical root distribution in determining the performance of three irrigated wheat cultivars under terminal water stress (TWS). A field experiment was conducted during the 2021–2022 growing season at Shiraz University (School of Agriculture), evaluating three bread wheat cultivars: Pishgam, Torabi, and Sirvan. The low-cost "pasta strainer" method, originally developed for evaluating rice root systems using plastic baskets, was adapted and employed to assess wheat mature root architecture in this context. To assess vertical root distribution, roots protruding through the side perforations of strainers were counted at three defined depth intervals: shallow (2–8 cm; SRN), middle (8–10 cm; MRN), and deep (10–13 cm; DRN). The results revealed significant differences in root architecture among cultivars. Compared to Sirvan and Torabi, Pishgam exhibited a more uniform root distribution across SRN, MRN, and particularly DRN. A significant interaction between irrigation and cultivar was observed for root weight, root volume, shoot biomass, and grain yield (GY). Pishgam appeared to be superior in most of measured traits. GY showed a positive association with MRN and DRN, which was further supported by principal component analysis aligning GY with root weight, root volume, MRN, and DRN. Despite the advantages and limitations of the pasta strainer method, our results demonstrated that cultivar with deeper rooting type, exhibited better drought performance.

Keywords: Pasta strainer, Root phenotyping, Root architecture, Mature root, Vertical root distribution.

1 Introduction

Bread wheat (*Triticum aestivum* L., $2n = 6x = 42$, AABBDD) is the most important cereal crop with significant increase in contribution of human food particularly in providing both calories and protein (Amini et al., 2023; Arzani & Ashraf, 2017). With widespread cultivation across more than

¹ Department of Plant Production and Genetics, School of Agriculture, Shiraz University, Shiraz, Islamic Republic of Iran.

* Corresponding author; e-mail: yaemam@shirazu.ac.ir or yaemam@gmail.com

219 million hectares, wheat contributes significantly to global agricultural production. Annually, wheat production exceeds 800 million tons of grain (FAO, 2024), making it a critical staple crop for sustaining populations worldwide. Climate change, particularly drought, reduces wheat production by limiting water availability, impairing growth, and decreasing yield potential. Rising temperatures and erratic rainfall further stress crops, leading to shorter growing seasons, lower grain quality, and reduced productivity (Pirnajmedin et al., 2024).

Drought, one of the foremost abiotic stresses, poses a considerable challenge to crop production, including wheat, and is further exacerbated by the depletion of water resources and climate change (Haghpanah et al., 2024). The severity, duration, and timing of drought stress occurrence during the growth stages of wheat can significantly reduce productivity, with potential losses ranging from 40% to 92% (Mahdavi et al., 2023; Senapati et al., 2019; Zamani et al., 2024; Zampieri et al., 2017). The primary effects of drought stress include dehydration, osmotic imbalance, dysfunction of endosomes and plasma membranes, loss of cellular turgor, inhibition of metabolite synthesis, depletion of cellular energy, oxidative stress, nutrient imbalance, reducing starch and protein content, and impaired photosynthesis (Haghpanah et al., 2024; Mokhtari et al., 2025). Water deficits manifest after the flowering stage, a phenomenon known as terminal drought, characterized by a decrease in precipitation and an increase in evaporation rates (Turner & Nicolas, 1987). However, the development of a profound water deficit can be mitigated by the presence of a deeper root system, enabling the wheat plants to access water stored at deeper soil layers, even during terminal drought stress (Figueroa-bustos et al., 2020). Khodaei et al. (2021) reported that the most root characteristics are under genetic control and significant associations were found for the deep rooting component. They identified candidate loci having significant effect on different aspects of the studied population particularly the deep rooting system and the root development under water-stress environment.

While both above and belowground traits play critical roles in plant responses to drought, most studies have primarily focused on aboveground traits, neglecting the importance of underground characteristics (Asefa et al., 2022). This oversight persists because root studies are inherently challenging due to the hidden nature of roots, soil heterogeneity, difficulties in non-destructive sampling, and the complex interactions between roots and their surrounding environment (Pirnajmedin et al., 2015, 2016). Therefore, it is crucial to shift attention towards investigating and comprehending the belowground characteristics of wheat plants.

Most studies on terminal drought tolerance in wheat have focused on evaluating morphological and physiological traits (Haghshenas et al., 2021; M. Mohammadi et al., 2021; Mokhtari et al., 2024, 2025; Raheison et al., 2020; Zamani et al., 2024). While these traits serve as important indicators for drought selection, a holistic understanding requires deeper exploration of root-system dynamics. By elucidating the intricate interplay between above- and belowground traits, we can unravel the plant's full adaptive mechanisms to drought stress. This integrated approach will enable the development of more effective breeding strategies to enhance drought resistance in wheat cultivars. Roots, as one of the primary organs of plants, play a pivotal role in detecting soil dehydration cues, highlighting the critical importance of elucidating optimal root system architecture, particularly with respect to its capacity for deep soil penetration and moisture extraction (X. Li et al., 2019). The responsiveness of the subterranean organ, especially under conditions of limited soil moisture, is fundamental for efficient water uptake and sustained physiological activity (Ranjan et al., 2022). Modeling studies have demonstrated that for each additional millimeter of soil water extracted by wheat roots after anthesis, grain yield may increase by approximately 55 kg per hectare (Manschadi et al., 2006), underscoring the substantial contribution of root traits to yield formation, particularly in environments where evaporative demand often exceeds precipitation during grain filling. These observations emphasize the need for comprehensive investigations into the functional relationships between root architectural traits and drought tolerance mechanisms.

The direct selection of root traits in breeding programs remains challenging, primarily due to the high costs, labor intensity, and time-consuming nature of root system studies. To overcome these limitations, several studies have explored root system architecture using diverse phenotyping approaches. These include small-scale controlled-environment studies (Basirat et al., 2019; Chen et al., 2020; Gioia et al., 2017; Hosseini et al., 2024; Khodaei et al., 2021; Nagel et al., 2020; Richard et al., 2015), field-based root phenotyping (Crocker et al., 2003; Fradgley et al., 2020; Maccaferri et al., 2016; Mehrabi et al., 2021; A. Wasson et al., 2016), as well as indirect root trait assessment through remote sensing technologies (Deery et al., 2019; Gutierrez et al., 2010; X. Li et al., 2019; Pinto & Reynolds, 2015). Additionally, many of the existing root phenotyping techniques lack sufficient accuracy and reproducibility (El Hassouni et al., 2018; Ober et al., 2021; A. P. Wasson et al., 2012). To address these limitations, we employed the “pasta strainer (basket)” method in our field experiment. This technique has previously been validated as a robust and

practical approach for capturing genotypic variation in root system architecture, as demonstrated in both rice (Uga et al., 2009) and durum wheat under controlled conditions (El Hassouni et al., 2018), with significant implications for agronomic performance. The pasta strainer method offers multiple advantages, including operational simplicity, cost-effectiveness, and ease of deployment in field settings. The aim of this study was to gain insights into how root traits could contribute to the performance of three Iranian wheat cultivars under terminal water stress conditions.

2 Materials and Methods

2.1 Growth conditions

During the 2021-2022 growing season, a field experiment was carried out at the research field of the School of Agriculture, Shiraz University, located at 29°43' N latitude and 52°35' E longitude in Iran. The experimental site had an elevation of 1788 m above sea level. The study involved three irrigated winter wheat cultivars, which were manually sown into pasta strainer baskets on November 9, 2021. Following sowing, drip irrigation was immediately initiated, with individual drip tapes installed along each row to ensure uniform water delivery. Standard integrated pest, weed, and disease management protocols were implemented throughout the growing season to maintain optimal crop health. The physico-chemical characteristics of the experimental soil are provided in Table 1. Based on soil test recommendations, nitrogen fertilizer was applied at a total rate of 150 kg N ha⁻¹ in the form of urea, distributed equally in three split applications at sowing, early tillering, and anthesis stages. Detailed meteorological data were recorded throughout the cropping season (Table 2).

Table 1. Physico-chemical characterization of the soil (0–30 cm depth) at the experimental site during the 2021–2022 growing season.

Electrical conductivity (dS.m ⁻¹)	Clay (%)	Silt (%)	Sand (%)	Total Nitrogen (%)	Organic matter (%)	Phosphorus (mg.kg ⁻¹)	Potassium (mg.kg ⁻¹)
0.97	39	40	21	0.12	0.50	21.5	561

Table 2. Climatic data of the experimental site at School of Agriculture during the 2020–2021 cropping season.

Month	Tmax (°c)	Tmin (°c)	Tavg (°c)	RHavg (%)	Pavg (mm)	SD (h/day)	Evap (mm)	Wind (m/s)
November	17.7	2.3	10	51.8	2.2	6.6	2.08	0.75
December	12.7	-1.3	5.7	55.3	2.0	5.9	1.0	0.92
January	13.8	-7.1	3.4	45.3	0.0	8.0	1.5	1.24
February	16.9	-2.7	7.1	52.2	1.7	8.4	8.3	1.75
March	21.2	2.7	12.0	46.0	0.3	8.3	9.3	1.76
April	26.4	4.7	15.5	46.0	0.2	8.7	11.9	1.45
May	29.9	9.0	19.5	41.4	0.0	10.3	13.7	1.65
June	36.6	13.6	25.1	25.9	0.0	12.0	20.3	1.1

Abbreviation. Tmax: Maximum Temperature; Tmin: Minimum Temperature; Tavg: Average Temperature; RHavg: Average Relative Humidity; Pavg: Precipitation Average; SD: Sunshine Duration; Evap: Evaporation.

2.2 Experimental design and treatments

The experiment was conducted following a split-plot design based on a randomized complete block design with three replicates. The main plots were assigned to two irrigation regimes: well-watered (WW) and terminal water stress (TWS). The subplots consisted of three irrigated winter wheat cultivars i. e. Pishgam, Sirvan, and Torabi (Table 3). In the WW treatment, irrigation continued until maturity, ensuring non-limiting soil moisture conditions. In contrast, the TWS treatment involved withholding irrigation from the 50% anthesis stage onwards to simulate terminal drought stress conditions representative of semi-arid environments such as the Persian Plateau (Najafi & Alizadeh, 2023).

Table 3. Characteristics of bread wheat cultivars evaluated in the experiment.

Wheat cultivars	Release year	Growth period	Climate	Growth habit	Pedigree	Origin	References
Pishgam	2002	Middle-ripening	Cold regions with drought-stressed conditions	Spring	Bkt/90-Zhong87	Iran	(Mahfoozi et al., 2009)
Sirvan	2011	Relative ly early-ripening	Temperate regions with drought-stressed conditions	Spring	PRL/2*PASTOR	Mexico (CIMMYT)	(Najafian et al., 2012)
Torabi	2019	Early-ripening	Temperate regions with drought-stressed conditions	Spring	CHEN/AEGILOPS SQUARROSA (TAUS)//BCN/3/BAV92/4/BERKUT	Mexico (CIMMYT)	(Bakhtiar et al., 2021)

2.3 Evaluation of mature root system characteristics

The pasta strainer method employed in this study was adapted from the methodology described by Uga et al. (2009), which was originally developed for assessing rice root systems using plastic baskets. This approach was subsequently modified for durum wheat under controlled conditions by El hassouni et al. (2018). In the present study, we further refined and applied this method under field conditions to evaluate root traits in irrigated bread wheat cultivars (Fig. 1). The plastic strainers used measured 13 cm in height and 25 cm in diameter. Each strainer was filled with field soil and buried to a depth of 13 cm to ensure alignment with the surrounding soil profile. To allow for adequate plant spacing and unrestricted three-dimensional root development, strainers were placed 50 cm apart, resulting in a final plant spacing of 75 cm between individual plants (Fig. 1a). Five seeds were sown at the center of each basket, and seedlings were later thinned to one plant per basket at the early tillering stage (Fig. 2b) (Zadoks et al., 1974). At physiological maturity, the aboveground plant parts were harvested at the soil surface, and shoot biomass (SB) was determined after oven-drying. Spikes were subsequently harvested and threshed to quantify grain yield (GY) using a precision balance. Following the harvest of aboveground parts, the plastic strainers containing the root systems were carefully extracted from the soil through manual excavation with a spade to allow for detailed root trait evaluation. Each strainer was then marked and sectioned into three vertical soil layers: shallow (2–8 cm), intermediate (8–10 cm), and deep (10–13 cm),

corresponding to angular sectors of 0–30°, 30–60°, and 60–90° from the vertical axis, respectively (A schematic illustration is shown in Fig. 2c). For each section, the number of roots protruding through the perforations in the side walls of the containers was counted (SRN, MRN, and DRN, respectively), enabling quantitative evaluation of root distribution across soil depths. Following root counting, all visible roots protruding from the container walls were carefully clipped to isolate a fixed soil-root volume equivalent to that of the pasta strainer. The baskets were then submerged in buckets of water to facilitate gradual disintegration of the soil matrix while minimizing mechanical damage to the root structures. The root samples were subsequently rinsed thoroughly with clean water to remove any adhering soil particles. Thereafter, the cleaned roots were oven-dried at 40 °C for 48 hours to a constant weight, and the resulting dry mass was recorded as root weight (RW).

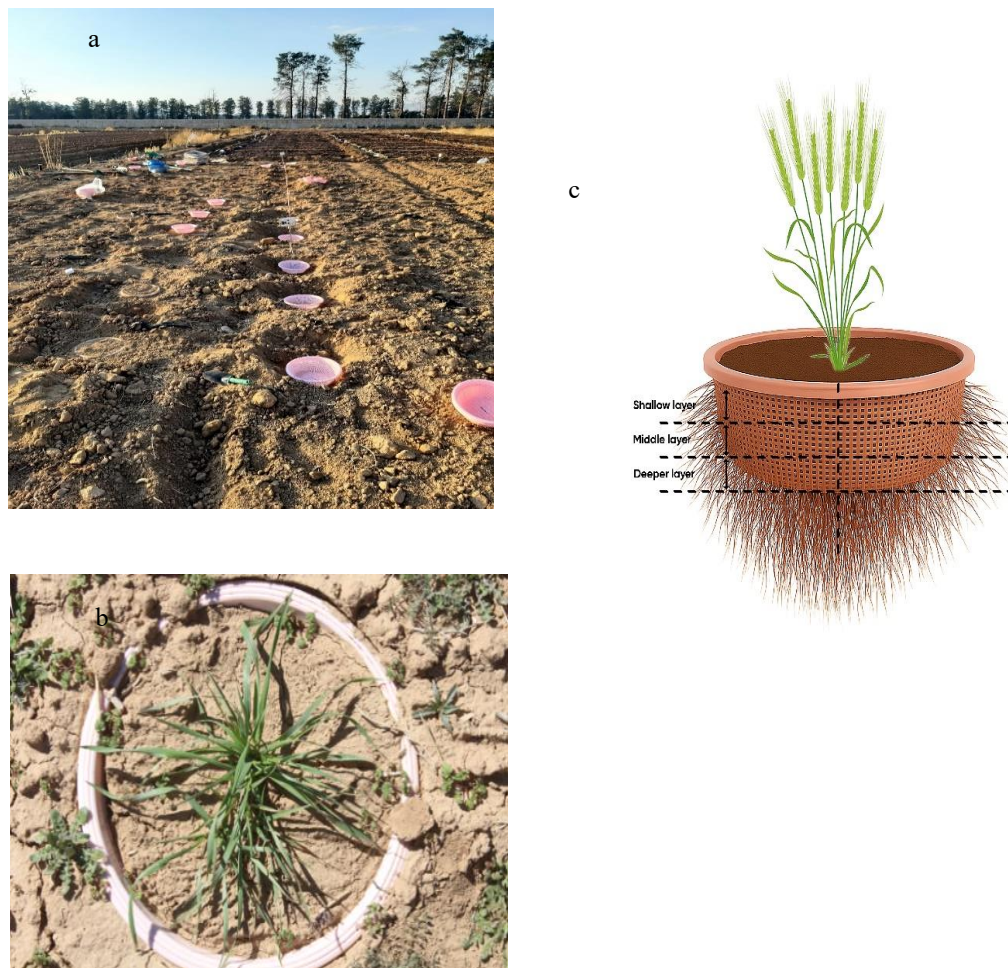


Fig. 1. Application of the pasta strainer technique to evaluate root system architecture in three irrigated bread wheat cultivars under terminal water stress in field conditions. (a) Bed preparation for basket burial and seed sowing at the center of each basket. (b) Wheat plants grown in the pasta strainers. (c) Schematic illustration of a pasta strainer removed from the soil, showing depth-based segmentation and root counting. The strainer was divided into three depth intervals: 2–8 cm (shallow root number, SRN), 8–10 cm (middle root number, MRN), and 10–13 cm (deep root number, DRN). Roots protruding through the side perforations were counted at each depth to quantify vertical root distribution.

2.4 Statistical analyses

The collected data were subjected to statistical analyses using SAS software (version 9.3; SAS Institute, Cary, NC, USA) to assess the significance of treatment effects. The Shapiro–Wilk test was employed to evaluate the normality assumption of the residuals, while Bartlett’s test was used to verify the homogeneity of variances. Analysis of variance (ANOVA) was performed to detect

significant differences among treatment groups, and mean comparisons were conducted using the least significant difference (LSD) test at a significance level of $p < 0.01$. In addition, Pearson correlation coefficients were calculated using SRplot software (Tang et al., 2023) to examine the associations among measured traits. Principal component analysis (PCA) was carried out in R software (version 4.4.2; R Core Team, Vienna, Austria) utilizing the *ggplot2* and *factoextra* packages for visualization and multivariate data interpretation.

3 Results & discussion

The ANOVA revealed significant main effects of irrigation on RW, and RV, SB, and GY. In contrast, the cultivar factor had a significant effect on all measured traits, including root number in the shallow, middle, and deeper soil layers (SRN, MRN, and DRN, respectively), total root number (TRN), RW, RV, SB, and GY. Moreover, the interaction between irrigation and cultivar was also significant for RW, RV, SB, and GY (Table 4).

Table 4. Analysis of variance (ANOVA) for above- and belowground traits in three wheat cultivars under two irrigation regimes including well-watered and terminal water stress conditions.

	df	SRN	MRN	DRN	TRN	RW	RV	SB	GY
Block	2	7.39	20.22	33.50	142.17	3.73	37.50	45.37	2.55
Irrigation	1	20.05	26.88	9.388	162	78.33***	272.22**	3411.65*	555**
							*	**	*
Irrigation *Block	2	2.06	6.99	22.72	33.17	4.561	26.39	62.81	3.77
Cultivar	2	48.39**	49.55**	236.17*	852.66**	20.11***	304.16**	745.50**	136.28
					*		*	*	**
Irrigation*Cultivar	2	0.72	2.89	35.72	26	19.97***	134.72*	149.42**	50.48*
								*	
Error	8	5.88	6.72	48.36	80.75	2.25	21.52	15.81	10.50
CV		9.19	9.22	9.50	7.04	21.31	1.44	7.24	14.27

*, **, *** Significant at the 0.05, 0.01, and 0.001 probability levels, respectively.

Abbreviations: WW, well-watered; TWS, terminal water stress; SRN, shallow layer root number; MRN, middle layer root number; DRN, deeper layer root number; TRN, total root number; RW, root weight; RV, root volume; SB, shoot biomass; GY, grain yield.

3.1 Variability in rooting pattern

The cultivars exhibited distinct variability in root distribution patterns (Fig. 2). In the shallow soil layer (Fig. 2a), Pishgam had the highest SRN (29.33), followed by Sirvan (26.17) and Torabi (23.67). Similarly, in the intermediate layer (Fig. 2b), Pishgam and Sirvan displayed higher MRN (30.67 and 28.67, respectively) compared to Torabi (25.00). In the deepest layer (Fig. 2c), Pishgam again showed the highest DRN (78.67), followed by Sirvan (74.50), while Torabi had the lowest DRN (66.33). Overall, Pishgam consistently exhibited a higher TRN across soil depths (Fig. 2d),

particularly in the deeper layers, suggests that Pishgam may possess a more extensive and developed root system, potentially enabling greater access to deeper soil moisture. Moreover, significant variation among cultivars in the assessed belowground traits highlights the utility of the pasta basket method as a valuable tool for genetic studies on root distribution. El Hassouni et al. (2018) stated that genotypes that allocate a larger proportion of their root at depth demonstrated a significant increase in GY, ranging from 28 to 42% on average. Similarly, Severini et al. (2020) found a positive correlation between deeper root systems and yield advantages. A narrow root system architecture facilitates deeper soil penetration and enhances the plant's ability to extract moisture reserves during the critical grain-filling phase (Botwright Acuña & Wade, 2012; Ober et al., 2021). In bread wheat (X. Li et al., 2019), barley (Robinson et al., 2018), durum wheat (El Hassouni et al., 2018), triticale (Severini et al., 2020), deeper root have been linked to improved yields. The orientation of root growth plays a crucial role in shaping root system architecture, influencing how roots are distributed within the soil profile (Borrell et al., 2014; Boudiar et al., 2021). In climates where terminal water stress often coincides with critical growth stages, deeper root growth becomes essential, as it allows plants to access water from deeper soil layers and sustain growth despite surface water scarcity. Conversely, a wider root angle promotes exploration of the upper soil layers, which can be advantageous for capturing moisture from the surface and from frequent, low-intensity rainfall events (Alahmad et al., 2019; Nakhforoosh et al., 2021; Ober et al., 2021).

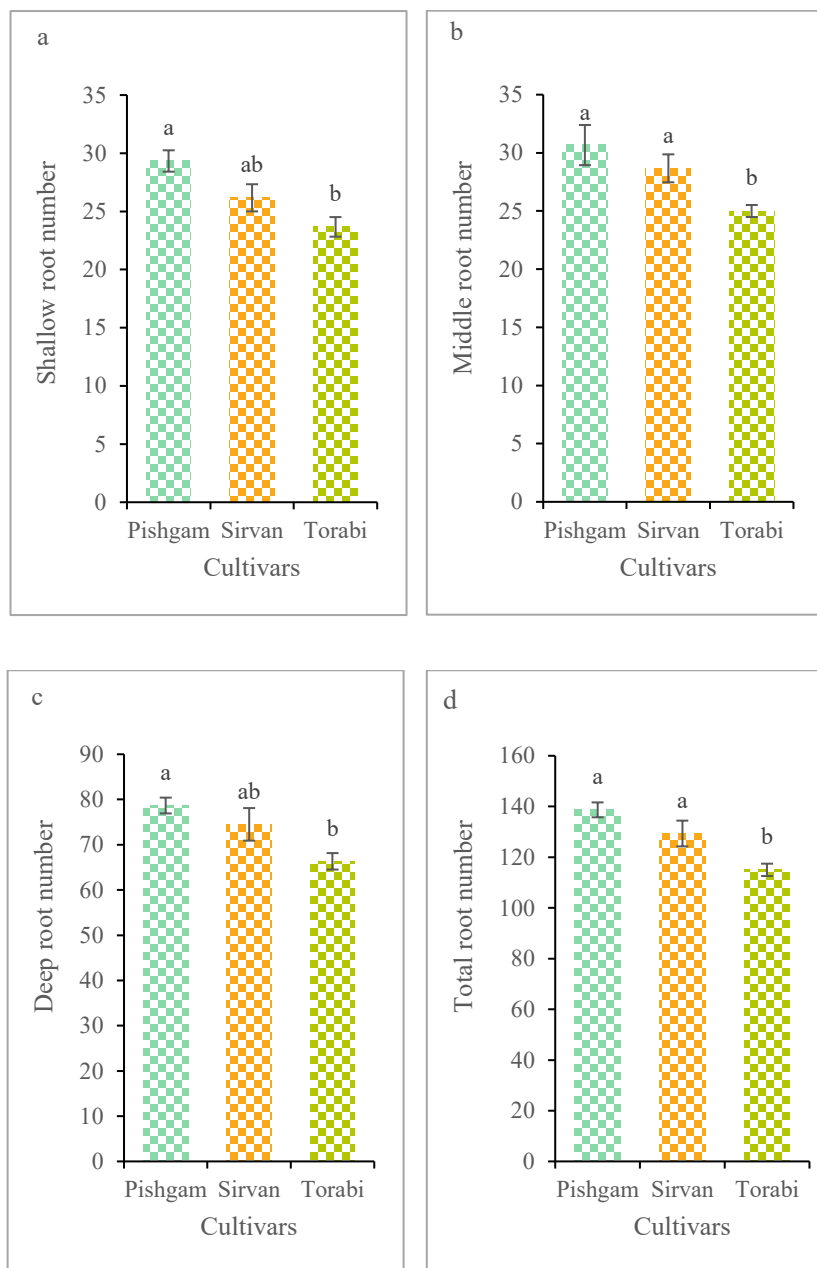


Fig. 2. Comparison of mean root numbers for three irrigated bread wheat cultivars across soil layers assessed using the pasta strainer technique. Root numbers were recorded at (a) shallow (2–8 cm), (b) middle (8–10 cm), (c) deep (10–13 cm) soil depths, and (d) total root number. The same letters in each column suggest a 5% probability of non-significance for the least significant difference (LSD) test.

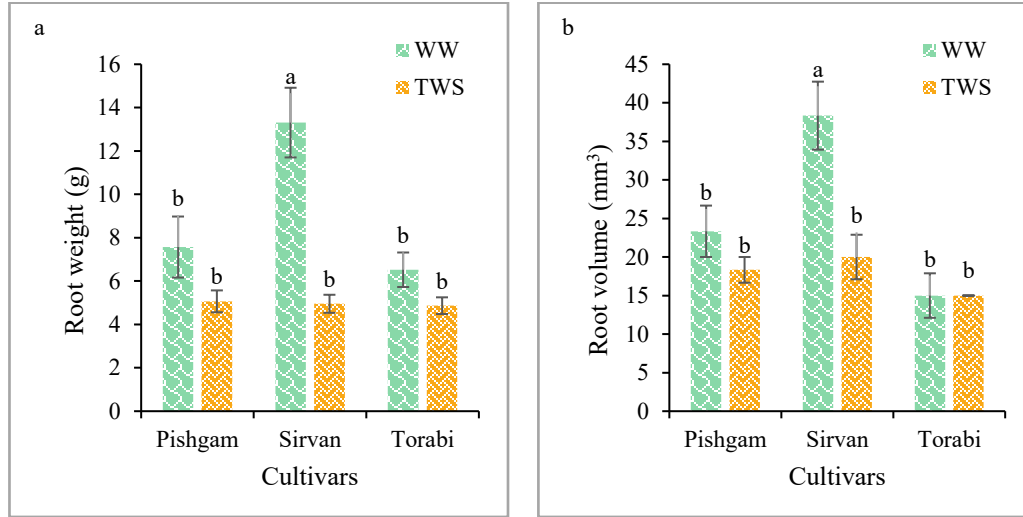


Fig. 3. Comparison of mean (a) root weight and (b) root volume for three irrigated bread wheat cultivars assessed using the pasta strainer technique under contrasting irrigation treatments: well-watered (WW) and terminal water stress (TWS). The same letters in each column suggest a 5% probability of non-significance for the least significant difference (LSD) test.

According to Fig. 3, Sirvan displayed a significantly higher RW and RV under WW conditions than other cultivars (13.31 g, 38.33 mm³, respectively). The elevated RW observed in Sirvan may be attributed to its greater RV, which is determined by multiplying root diameter by root length. In wheat, RW is influenced by various components, including root number, length, and diameter (Ehdaie et al., 2012). Therefore, the increase in RW in Sirvan could be the result of an increase in one or a combination of these root system traits. As the number of mature roots is typically established prior to the onset of TWS, the higher RW in Sirvan is likely due to thicker roots, which aligns with observations made during the experiment. Although Sirvan exhibited higher RW than Pishgam under WW conditions, it is noteworthy that Sirvan produced lower GY than Pishgam. This disparity suggests that factors beyond RW and RV may play a critical role in determining the overall performance of these cultivars.

3.2 Above-ground features influenced by irrigation and cultivar interaction

Under WW conditions, the Pishgam had significantly higher GY (35.08 g plant⁻¹) compared to the other cultivars (Fig. 4a). Sirvan followed with a mean GY of 28.65 g plant⁻¹, which, although slightly lower than Pishgam, was still significantly different. In contrast, Torabi showed a mean GY of 21.07 g plant⁻¹, significantly lower than Sirvan under WW conditions. These results indicate

that Pishgam possesses a strong ability to perform under optimal water availability, underscoring its potential as a high-yielding cultivar.

However, the situation changed markedly under TWS conditions (Fig. 4a). Pishgam, which had demonstrated exceptional performance under WW conditions, experienced a significant reduction in GY, with a mean of 21.00 g plant⁻¹. Similarly, Sirvan showed a notable decrease in GY under TWS, recording a mean value of 16.65 g plant⁻¹. There was no significant difference between Pishgam and Sirvan under stress conditions. Torabi also experienced a reduction in GY under TWS, with a mean of 13.84 g plant⁻¹, reflecting a significant decline compared to its performance under WW conditions. TWS led to a yield reduction of 40% in Pishgam, 42% in Sirvan, and 34% in Torabi. Despite the reductions, Pishgam consistently maintained the highest GY under both WW and TWS conditions, outperforming both Torabi and Sirvan. Nonetheless, Torabi demonstrated greater yield stability under stress compared to the other two cultivars.

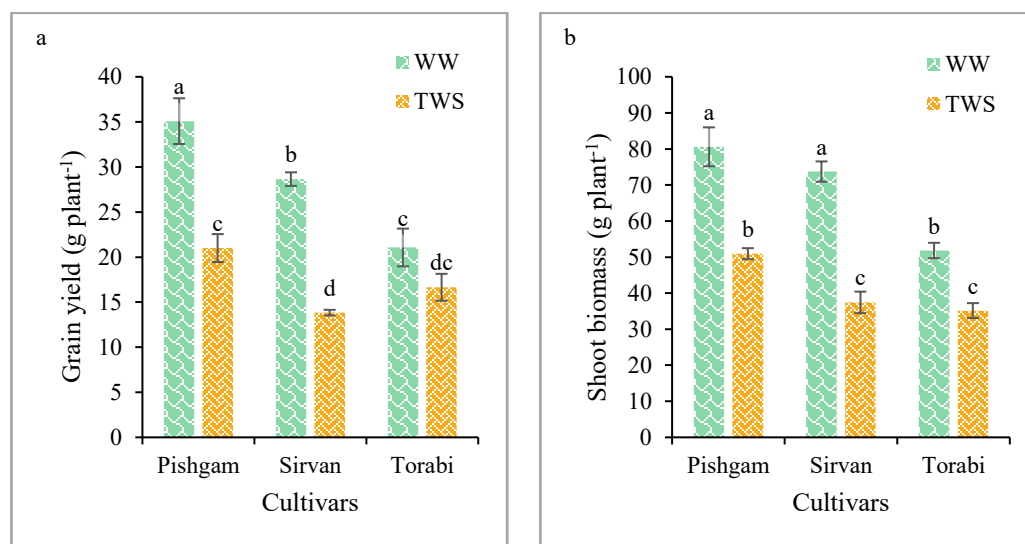


Fig. 4. Comparison of mean (a) grain yield and (b) shoot biomass for three irrigated bread wheat cultivars assessed using the pasta strainer technique under contrasting irrigation treatments: well-watered (WW) and terminal water stress (TWS). The same letters in each column suggest a 5% probability of non-significance for the least significant difference (LSD) test.

A similar trend was observed for SB among the cultivars (Fig. 4b). The imposition of TWS can substantially impair photosynthetic capacity, primarily through the disruption of carbon assimilation processes and the subsequent translocation of photoassimilates to developing grains. Such impairments compromise the grain filling process, ultimately resulting in decreased grain

size and GY (Senapati et al., 2019; Xie et al., 2015). Consistent with these observations, Zamani et al. (2024) using the same cultivars, reported that plants exposed to TWS exhibited a reduction in grain size accompanied by a more rapid depletion of grain moisture content relative to those grown under WW conditions. Extracting each millimeter of water from the soil throughout the crucial grain filling phase could lead to an increase in wheat yield by around 55 kg per hectare (Manschadi et al., 2006, 2008). Physiological responses to water deficit are largely governed by the functionality and efficiency of the root system. Nevertheless, under certain conditions, these adverse effects can be mitigated by root systems possessing enhanced capacity for water foraging, even under restricted soil moisture availability (Awad et al., 2018; Shazadi et al., 2024).

As shown in Fig. 5, strong and statistically significant positive correlations were observed between GY, SB, MRN, and DRN. In contrast, the correlation between GY and SRN was weaker and not statistically significant. Also, the principal component analysis (PCA) biplot (Fig. 6) illustrates the multivariate relationships among traits, functional attributes, and cultivar responses under contrasting water regimes. The first two principal components (Dim1 and Dim2) explained jointly 92.1% of the total variance, with Dim1 accounting for 71.6% and Dim2 for 20.5%, thereby capturing the predominant patterns within the dataset. Dim1, which explained the majority of the variation, was predominantly driven by root system architecture traits, including TRN, MRN, and DRN, as indicated by their high squared cosine values ($\cos^2 > 0.9$) and extended vector lengths. This highlights the substantial contribution of these root traits to the overall variability observed among treatments. Similarly, RV and RW also exhibited strong associations with Dim1, further emphasizing the integral role of root biomass in differentiating cultivar performance under varying water availability. GY and SB were associated with both Dim1 and Dim2, reflecting the combined influence of root architectural traits and environmental conditions on these productivity parameters. The alignment of GY and SB vectors with those of RV, RW, and deeper root distribution traits such as DRN and MRN suggests a positive association between increased root biomass, enhanced root penetration into deeper soil layers, and aboveground productivity. This reinforces the pivotal role of belowground resource capture in driving SB accumulation and yield formation. **These findings suggest that selecting for a narrower root distribution (i.e., higher MRN and DRN) may represent a more effective breeding strategy for enhancing yield under water-limited conditions.** From a physiological standpoint, the development of a deep root system enables plants to exploit water reserves located in deeper soil horizons, particularly during critical

reproductive phases such as grain filling, when transpirational demand is heightened. This trait confers a substantial adaptive advantage under terminal drought conditions, which are typified by the rapid depletion of surface soil moisture (C. Li et al., 2021; Palta et al., 2011; Ranjan et al., 2022; Shazadi et al., 2024). Deep rooting thus enhances drought avoidance mechanisms and contributes to yield stability under water-limited environments. In contrast, while shallow root systems may facilitate early-season nutrient acquisition, their limited penetration depth restricts water uptake during prolonged soil moisture deficits, ultimately compromising yield sustainability (R. Mohammadi, 2024; Vadez, 2014). Clearly, each root architectural configuration exhibits differential adaptive suitability depending on specific environmental conditions.

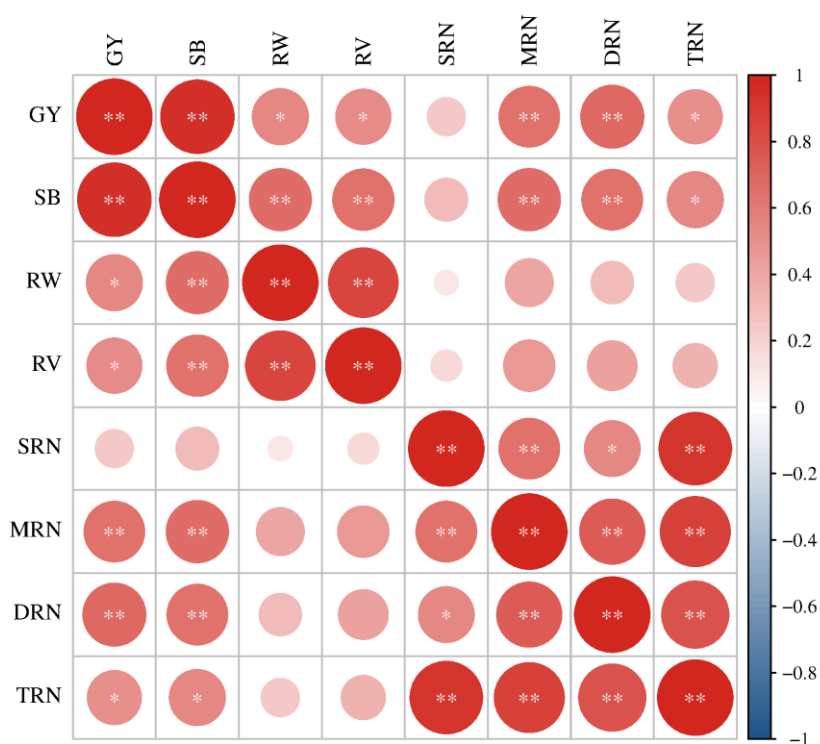


Fig. 5. Correlation coefficient for above- and belowground traits in bread wheat cultivars under irrigation regimes. The size of each circle indicates the correlation intensity between two features. *, ** Significant at the 0.05 and 0.01 probability levels, respectively. Abbreviations: TRN, total root number; DRN, deeper layer root number; MRN, middle layer root number; SRN, shallow layer root number; RW, root weight; RV, root volume; SB, shoot biomass; GY, grain yield.

Recent studies on wheat root have primarily focused on characterizing the root system during the seedling stage, which may not provide a complete understanding of the root system in adult plants. Additionally, these studies have often overlooked examining of the relationship between root and shoot traits, including the crucial trait of GY. In this study, we sought to address these limitations by investigating the root system and its behavior at plant maturity. By doing so, we aimed to gain a more comprehensive understanding of the adult plant root system and its association with GY under varying conditions. Our approach allowed us to explore the intricate connections between root distribution in different soil layers and GY. Notably, root distribution across the evaluated depths remained unaffected under TWS condition, further supporting the notion of strong genetic control over these traits. However, it should be acknowledged that the absence of significant effects observed in this study does not preclude the possibility that alternative water regimes could exert more pronounced influences on rooting patterns. By adopting this approach, the efficiency, consistency, and scalability of root trait evaluation can be substantially improved while minimizing resource and time requirements. However as root features are challenging to enhance through conventional selection strategies, and therefore, molecular approaches such as genome-wide association studies (GWAS) may help overcome some of the limitations (Khodaei et al., 2021; Pirnajmedin et al., 2016).

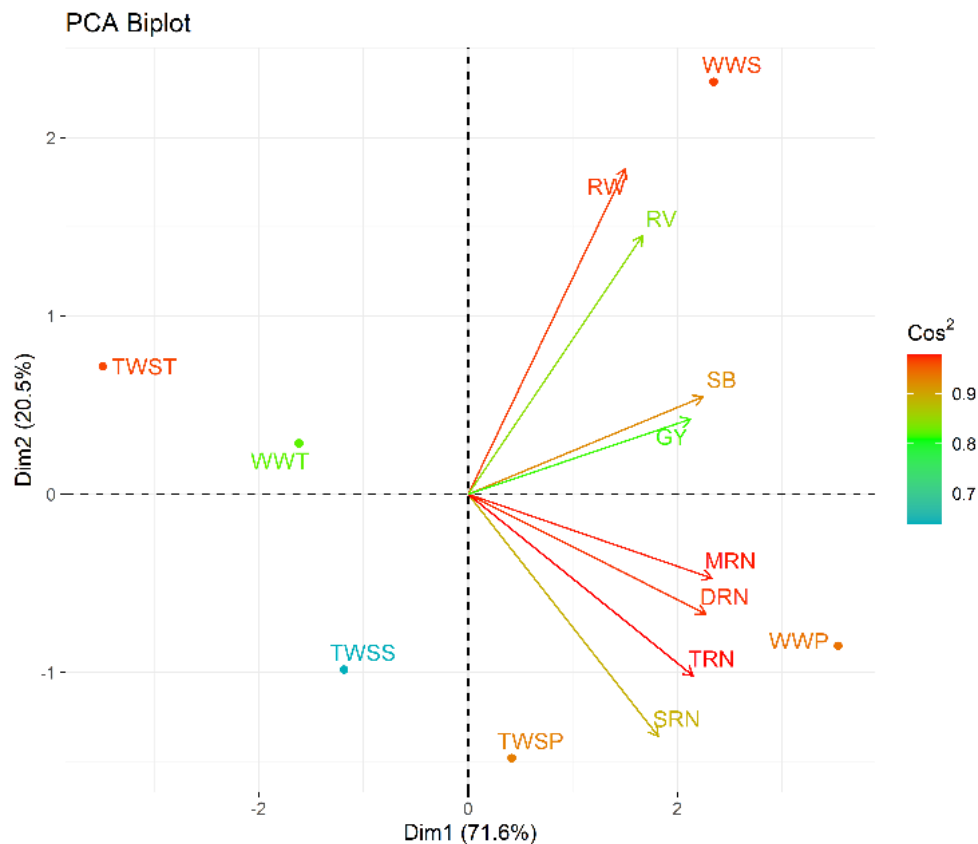


Fig. 6. Principal component analysis (PCA) biplot analysis between above and underground, functional traits treatments. RW, root weight; RV, root volume; SRN, shallow root number; MRN, middle root number; DRN, deeper root number; TRN, total root number; SB, shoot biomass; GY, grain yield; WWP, well-watered Pishgam; WWS, well-watered Sirvan; WWT, well-watered Torabi; TWSP, terminal water stress Pishgam; TWSS, terminal water stress Sirvan; TWST, terminal water stress Torabi.

4 Conclusions

Phenotyping root traits in wheat has long been constrained by inadequate methodologies. Despite the advantages and limitations of the pasta strainer method, our results demonstrated that cultivar with deeper rooting type, exhibited better drought performance. Based on the PCA results and the significant positive correlations observed between GY with both MRN and DRN underscore the critical contribution of root system architecture to crop productivity. These findings suggest that selecting for enhanced root traits can aid in identifying high-yielding cultivars under both TWS and WW conditions. As climate variability intensifies, especially in regions reliant on stored soil moisture, the promotion of vertical root development will be essential for improving wheat

resilience. The present study provides usable knowledge to inform breeding programs toward more adaptive, drought-resilient cultivars especially in semi-arid and arid regions.

References

- Alahmad, S., El Hassouni, K., Bassi, F. M., Dinglasan, E., Youssef, C., Quarry, G., Aksoy, A., Mazzucotelli, E., Juhász, A., Able, J. A., Christopher, J., Voss-Fels, K. P., & Hickey, L. T. (2019). A major root architecture QTL responding to water limitation in durum wheat. *Frontiers in Plant Science*, 10(April), 1–18. <https://doi.org/10.3389/fpls.2019.00436>
- Amini, A., Majidi, M. M., Mokhtari, N., & Ghanavati, M. (2023). Drought stress memory in a germplasm of synthetic and common wheat: antioxidant system, physiological and morphological consequences. *Scientific Reports*, 13(1), 8569. <https://doi.org/10.1038/s41598-023-35642-2>
- Arzani, A., & Ashraf, M. (2017). Cultivated ancient wheats (*Triticum* spp.): a potential source of health-beneficial food products. *Comprehensive Reviews in Food Science and Food Safety*, 16(3), 477–488. <https://doi.org/10.1111/1541-4337.12262>
- Asefa, M., Worthy, S. J., Cao, M., Song, X., Lozano, Y. M., & Yang, J. (2022). Above- and below-ground plant traits are not consistent in response to drought and competition treatments. *Annals of Botany*, 130(7), 939–950. <https://doi.org/10.1093/aob/mcac108>
- Awad, W., Byrne, P. F., Reid, S. D., Comas, L. H., & Haley, S. D. (2018). Great plains winter wheat varies for root length and diameter under drought stress. *Agronomy Journal*, 110(1), 226–235. <https://doi.org/10.2134/agronj2017.07.0377>
- Bakhtiar, F., Najafian, G., Khodarahmi, M., Ahmadi, G. H., Jafar-Nejad, A., Sarikhani Khorami, S., Amin Azarm, D., Ghandi, A., Nabati, E., Zareh Faiz Abadi, A., Nikzad, A., Abdi, H., Afshari, F., Zakeri, A., Yasaei, M., Ata Hoseini, S. M., Tabatabai, N., Dalvand, M., Ebrahim Nejad, S., ... Tabatabaei, S. A. (2021). Torabi, new wheat cultivar suitable for water-stressed conditions of temperate regions of Iran. *Research Achievements for Field and Horticulture Crops*, 9(2), 101–113. <https://doi.org/10.22092/rafhc.2021.343116.1247>
- Basirat, M., Mousavi, S. M., Abbaszadeh, S., Ebrahimi, M., & Zarebanadkouki, M. (2019). The rhizosheath: a potential root trait helping plants to tolerate drought stress. *Plant and Soil*, 445(1), 565–575. <https://doi.org/10.1007/s11104-019-04334-0>
- Borrell, A. K., Mullet, J. E., George-Jaeggli, B., van Oosterom, E. J., Hammer, G. L., Klein, P. E.,

- 368 & Jordan, D. R. (2014). Drought adaptation of stay-green sorghum is associated with canopy
369 development, leaf anatomy, root growth, and water uptake. *Journal of Experimental Botany*,
370 65(21), 6251–6263. <https://doi.org/10.1093/jxb/eru232>
- 371 Botwright Acuña, T. L., & Wade, L. J. (2012). Genotype×environment interactions for root depth
372 of wheat. *Field Crops Research*, 137, 117–125.
373 <https://doi.org/10.1016/j.fcr.2012.08.004>
- 374 Boudiar, R., Cabeza, A., Fernández-Calleja, M., Pérez-Torres, A., Casas, A. M., González, J. M.,
375 Mekhlouf, A., & Igartua, E. (2021). Root trait diversity in field grown durum wheat and
376 comparison with seedlings. *Agronomy*, 11(12), 2545.
377 <https://doi.org/10.3390/agronomy11122545>
- 378 Chen, Y., Palta, J., Prasad, P. V. V., & Siddique, K. H. M. (2020). Phenotypic variability in bread
379 wheat root systems at the early vegetative stage. *BMC Plant Biology*, 20(1), 185.
380 <https://doi.org/10.1186/s12870-020-02390-8>
- 381 Crocker, T. L., Hendrick, R. L., Ruess, R. W., Pregitzer, K. S., Burton, A. J., Allen, M. F., Shan,
382 J., & Morris, L. A. (2003). Substituting root numbers for length: improving the use of
383 minirhizotrons to study fine root dynamics. *Applied Soil Ecology*, 23(2), 127–135.
384 [https://doi.org/10.1016/S0929-1393\(03\)00024-6](https://doi.org/10.1016/S0929-1393(03)00024-6)
- 385 Deery, D. M., Rebetzke, G. J., Jimenez-Berni, J. A., Bovill, W. D., James, R. A., Condon, A. G.,
386 Furbank, R. T., Chapman, S. C., & Fischer, R. A. (2019). Evaluation of the phenotypic
387 repeatability of canopy temperature in wheat using continuous-terrestrial and airborne
388 measurements. *Frontiers in Plant Science*, 10, 875. <https://doi.org/10.3389/fpls.2019.00875>
- 389 Ehdaie, B., Layne, A. P., & Waines, J. G. (2012). Root system plasticity to drought influences
390 grain yield in bread wheat. *Euphytica*, 186(1), 219–232. [https://doi.org/10.1007/s10681-011-](https://doi.org/10.1007/s10681-011-0585-9)
391 0585-9
- 392 El Hassouni, K., Alahmad, S., Belkadi, B., Filali-Maltouf, A., Hickey, L. T., & Bassi, F. M. (2018).
393 Root system architecture and its association with yield under different water regimes in
394 Durum wheat. *Crop Science*, 58(6), 2331–2346. <https://doi.org/10.2135/cropsci2018.01.0076>
- 395 FAOSTAT (2024), accessed at <https://www.fao.org/faostat/en/#data/QCL>
- 396 Figueroa-bustos, V., Palta, J. A., Chen, Y., Stefanova, K., Siddique, K. H. M., & Armstrong, R.
397 D. (2020). Wheat cultivars with contrasting root system size responded differently to terminal
398 drought. *Frontiers in Plant Science*, 11, 1–12. <https://doi.org/10.3389/fpls.2020.01285>

- 399 Fradgley, N., Evans, G., Biernaskie, J. M., Cockram, J., Marr, E. C., Oliver, A. G., Ober, E., &
400 Jones, H. (2020). Effects of breeding history and crop management on the root architecture
401 of wheat. *Plant and Soil*, 452(1), 587–600. <https://doi.org/10.1007/s11104-020-04585-2>
- 402 Gioia, T., Galinski, A., Lenz, H., Müller, C., Lentz, J., Heinz, K., Briese, C., Putz, A., Fiorani, F.,
403 Watt, M., Schurr, U., & Nagel, K. A. (2017). *GrowScreen-PaGe*, a non-invasive, high-
404 throughput phenotyping system based on germination paper to quantify crop phenotypic
405 diversity and plasticity of root traits under varying nutrient supply. *Functional Plant Biology*,
406 44(1), 76–93. <https://doi.org/10.1071/FP16128>
- 407 Gutierrez, M., Reynolds, M. P., & Klatt, A. R. (2010). Association of water spectral indices with
408 plant and soil water relations in contrasting wheat genotypes. *Journal of Experimental*
409 *Botany*, 61(12), 3291–3303. <https://doi.org/10.1093/jxb/erq156>
- 410 Haghpanah, M., Hashemipetroudi, S., Arzani, A., & Araniti, F. (2024). Drought Tolerance in
411 Plants: Physiological and Molecular Responses. *Plants*, 13(21), 2962.
412 <https://doi.org/10.3390/plants13212962>
- 413 Haghshenas, A., Emam, Y., Sepaskhah, A. R., & Edalat, M. (2021). Can extended phenology in
414 wheat cultivar mixtures mitigate post-anthesis water stress? *European Journal of Agronomy*,
415 122, 126188. <https://doi.org/10.1016/j.eja.2020.126188>
- 416 Hosseini, S. S., Razavi, B. S., & Lakzian, A. (2024). Drought tolerance of wheat genotypes is
417 associated with rhizosphere size and enzyme system. *Plant and Soil*, 502(1), 671–685.
418 <https://doi.org/10.1007/s11104-024-06576-z>
- 419 Khodaei, S. M. M., Hashemi, M., Mirlohi, A., Majidi, M. M., Sukumaran, S., Esmaelzaeh
420 Moghaddam, M., & Abdollahi, M. (2021). Root characteristics of an elite spring wheat panel
421 under contrasting water treatments and their genome-wide association study. *Rhizosphere*,
422 19, 100413. <https://doi.org/10.1016/j.rhisph.2021.100413>
- 423 Li, C., Li, L., Reynolds, M. P., Wang, J., Chang, X., Mao, X., & Jing, R. (2021). Recognizing the
424 hidden half in wheat: root system attributes associated with drought tolerance. *Journal of*
425 *Experimental Botany*, 72(14), 5117–5133. <https://doi.org/10.1093/jxb/erab124>
- 426 Li, X., Ingvordsen, C. H., Weiss, M., Rebetzke, G. J., Condon, A. G., James, R. A., & Richards,
427 R. A. (2019). Deeper roots associated with cooler canopies, higher normalized difference
428 vegetation index, and greater yield in three wheat populations grown on stored soil water.
429 *Journal of Experimental Botany*, 70(18), 4963–4974. <https://doi.org/10.1093/jxb/erz232>

- 430 Maccaferri, M., El-Feki, W., Nazemi, G., Salvi, S., Canè, M. A., Colalongo, M. C., Stefanelli, S.,
431 & Tuberosa, R. (2016). Prioritizing quantitative trait loci for root system architecture in
432 tetraploid wheat. *Journal of Experimental Botany*, 67(4), 1161–1178.
433 <https://doi.org/10.1093/jxb/erw039>
- 434 Mahdavi, Z., Rashidi, V., Yarnia, M., Aharizad, S., & Roustaii, M. (2023). Evaluation of yield
435 traits and tolerance indices of different wheat genotypes under drought stress conditions.
436 *Cereal Research Communications*, 51(3), 659–669. [https://doi.org/10.1007/s42976-022-](https://doi.org/10.1007/s42976-022-00322-w)
437 [00322-w](https://doi.org/10.1007/s42976-022-00322-w)
- 438 Mahfoozi, S., Akbari, A., Chaichi, M., Sanjari, A. G., Nazeri, S. M., Abedi-Oskooee, S.,
439 Aminzadeh, G., & Rezaie, M. (2009). Pishgam, a new bread wheat cultivar for normal
440 irrigation and terminal stage deficit irrigation conditions of cold regions of Iran. *Seed and*
441 *Plant Journal*, 25(3), 513–517. <https://doi.org/10.22092/spij.2017.110998>
- 442 Manschadi, A. M., Christopher, J., deVoil, P., & Hammer, G. L. (2006). The role of root
443 architectural traits in adaptation of wheat to water-limited environments. *Functional Plant*
444 *Biology*, 33(9), 823–837. <https://doi.org/10.1071/FP06055>
- 445 Manschadi, A. M., Hammer, G. L., Christopher, J. T., & deVoil, P. (2008). Genotypic variation in
446 seedling root architectural traits and implications for drought adaptation in wheat (*Triticum*
447 *aestivum* L.). *Plant and Soil*, 303(1), 115–129. <https://doi.org/10.1007/s11104-007-9492-1>
- 448 Mehrabi, F., Sepaskhah, A. R., & Ahmadi, S. H. (2021). Winter wheat root distribution with
449 irrigation, planting methods, and nitrogen application. *Nutrient Cycling in Agroecosystems*,
450 119(2), 231–245. <https://doi.org/10.1007/s10705-021-10120-1>
- 451 Mohammadi, M., Mirlohi, A., Majidi, M. M., & Rabbani, A. (2021). Exploring the breeding
452 potential of Iranian emmer wheats to increase durum wheat tolerance to drought. *Plant*
453 *Genetic Resources: Characterization and Utilization*, 19(4), 363–374. [https://doi.org/DOI:](https://doi.org/DOI:10.1017/S1479262121000435)
454 [10.1017/S1479262121000435](https://doi.org/DOI:10.1017/S1479262121000435)
- 455 Mohammadi, R. (2024). Effects of post-flowering drought and supplemental irrigation on grain
456 yield and agro-phenological traits in durum wheat. *European Journal of Agronomy*, 156,
457 127180. <https://doi.org/https://doi.org/10.1016/j.eja.2024.127180>
- 458 Mokhtari, N., Majidi, M. M., & Mirlohi, A. (2024). Physiological and antioxidant responses of
459 synthetic hexaploid wheat germplasm under drought. *BMC Plant Biology*, 24(1), 747.
460 <https://doi.org/10.1186/s12870-024-05445-2>

- 461 Mokhtari, N., Majidi, M. M., & Mirlohi, A. (2025). Synthetic wheat as a new source of flour
462 quality under drought conditions: Associations with solvent retention capacity. *PLOS ONE*,
463 20(2), e0316945. <https://doi.org/10.1371/journal.pone.0316945>
- 464 Nagel, K. A., Lenz, H., Kastenholz, B., Gilmer, F., Averagesch, A., Putz, A., Heinz, K., Fischbach,
465 A., Scharr, H., Fiorani, F., Walter, A., & Schurr, U. (2020). The platform GrowScreen-Agar
466 enables identification of phenotypic diversity in root and shoot growth traits of agar grown
467 plants. *Plant Methods*, 16(1), 89. <https://doi.org/10.1186/s13007-020-00631-3>
- 468 Najafi, M. S., & Alizadeh, O. (2023). Climate zones in Iran. *Meteorological Applications*, 30(5),
469 e2147. <https://doi.org/10.1002/met.2147>
- 470 Najafian, G., Khodarahmi, M., Amini, A., Afshari, F., Malihipour, A., Ahmadi, Gh. H.,
471 Nikooseresht, R., Kafashi, A. K., Amin, H., Zakari, A., Nikzad, A. R., Jafarnezhad, A., Afuni,
472 D., Hassanpour, J., Mohammadi, A. R., Atahossaini, S. M., Nazeri, A., Mirzaie, A. Allah,
473 Shourabi, A. A., ... Sadeghi, N. (2012). Sirvan, new bread wheat cultivar, tolerant to terminal
474 drought with good bread making quality adapted to irrigate conditions of temperate regions
475 of Iran. *Research Achievements for Field and Horticulture Crops*, 1(1), 1–10.
476 <https://doi.org/10.22092/rafhc.2012.100115>
- 477 Nakhforoosh, A., Nagel, K. A., Fiorani, F., & Bodner, G. (2021). Deep soil exploration vs. topsoil
478 exploitation: distinctive rooting strategies between wheat landraces and wild relatives. *Plant*
479 *and Soil*, 459(1), 397–421. <https://doi.org/10.1007/s11104-020-04794-9>
- 480 Ober, E. S., Alahmad, S., Cockram, J., Forestan, C., Hickey, L. T., Kant, J., Maccaferri, M., Marr,
481 E., Milner, M., Pinto, F., Rambla, C., Reynolds, M., Salvi, S., Sciara, G., Snowdon, R. J.,
482 Thomelin, P., Tuberosa, R., Uauy, C., Voss-Fels, K. P., ... Watt, M. (2021). Wheat root
483 systems as a breeding target for climate resilience. *Theoretical and Applied Genetics*, 134(6),
484 1645–1662. <https://doi.org/10.1007/s00122-021-03819-w>
- 485 Palta, J. A., Chen, X., Milroy, S. P., Rebetzke, G. J., Dreccer, M. F., & Watt, M. (2011). Large
486 root systems: are they useful in adapting wheat to dry environments? *Functional Plant*
487 *Biology*, 38(5), 347–354. <https://doi.org/10.1071/FP11031>
- 488 Pinto, R. S., & Reynolds, M. P. (2015). Common genetic basis for canopy temperature depression
489 under heat and drought stress associated with optimized root distribution in bread wheat.
490 *Theoretical and Applied Genetics*, 128(4), 575–585. [https://doi.org/10.1007/s00122-015-](https://doi.org/10.1007/s00122-015-2453-9)
491 2453-9

- 492 Pirnajmedin, F., Majidi, M., & Gheysari, M. (2015). Root and physiological characteristics
493 associated with drought tolerance in Iranian tall fescue. *Euphytica*, 202(1), 141–155.
494 <https://doi.org/10.1007/s10681-014-1239-5>
- 495 Pirnajmedin, F., Majidi, M., & Jaškūnė, K. (2024). Adaptive strategies to drought stress in grasses
496 of the poaceae family under climate change: Physiological, genetic and molecular
497 perspectives: A review. *Plant Physiology and Biochemistry*, 213, 108814.
498 <https://doi.org/10.1016/j.plaphy.2024.108814>
- 499 Pirnajmedin, F., Majidi, M. M., & Gheysari, M. (2016). Survival and recovery of tall fescue
500 genotypes: association with root characteristics and drought tolerance. *Grass and Forage*
501 *Science*, 71(4), 632–640. <https://doi.org/10.1111/gfs.12231>
- 502 Raherison, E., Majidi, M. M., Goessen, R., Hughes, N., Cuthbert, R., Knox, R., & Lukens, L.
503 (2020). Evidence for the Accumulation of Nonsynonymous Mutations and Favorable
504 Pleiotropic Alleles During Wheat Breeding. *G3 Genes|Genomes|Genetics*, 10(11), 4001–
505 4011. <https://doi.org/10.1534/g3.120.401269>
- 506 Ranjan, A., Sinha, R., Singla-Pareek, S. L., Pareek, A., & Singh, A. K. (2022). Shaping the root
507 system architecture in plants for adaptation to drought stress. *Physiologia Plantarum*, 174(2),
508 e13651. <https://doi.org/10.1111/ppl.13651>
- 509 Richard, C., Hickey, L. T., Fletcher, S., Jennings, R., Chenu, K., & Christopher, J. T. (2015). High-
510 throughput phenotyping of seminal root traits in wheat. *Plant Methods*, 11(1), 1–11.
511 <https://doi.org/10.1186/S13007-015-0055-9>
- 512 Robinson, H., Kelly, A., Fox, G., Franckowiak, J., Borrell, A., & Hickey, L. (2018). Root
513 architectural traits and yield: exploring the relationship in barley breeding trials. *Euphytica*,
514 214(9), 151. <https://doi.org/10.1007/s10681-018-2219-y>
- 515 Senapati, N., Stratonovitch, P., Paul, M. J., & Semenov, M. A. (2019). Drought tolerance during
516 reproductive development is important for increasing wheat yield potential under climate
517 change in Europe. *Journal of Experimental Botany*, 70(9), 2549–2560.
518 <https://doi.org/10.1093/jxb/ery226>
- 519 Severini, A. D., Wasson, A. P., Evans, J. R., Richards, R. A., & Watt, M. (2020). Root phenotypes
520 at maturity in diverse wheat and triticale genotypes grown in three field experiments:
521 Relationships to shoot selection, biomass, grain yield, flowering time, and environment. *Field*
522 *Crops Research*, 255, 107870. <https://doi.org/10.1016/j.fcr.2020.107870>

- Shazadi, K., Christopher, J. T., & Chenu, K. (2024). Does late water deficit induce root growth or senescence in wheat? *Frontiers in Plant Science*, 15, 1–14. <https://doi.org/10.3389/fpls.2024.1351436>
- Turner, N. C., & Nicolas, M. E. (1987). Drought resistance of wheat for light- textured soils in the mediterranean climate. In J. P. Srivastava, E. Porceddu, E. Acevedo, & S. Varma (Eds.), *Drought Tolerance in Winter Cereals* (pp. 203–216). John Wiley & Sons, Ltd.
- Uga, Y., Ebana, K., Abe, J., Morita, S., Okuno, K., & Yano, M. (2009). Variation in root morphology and anatomy among accessions of cultivated rice (*Oryza sativa* L.) with different genetic backgrounds. *Breeding Science*, 59(1), 87–93. <https://doi.org/10.1270/jsbbs.59.87>
- Vadez, V. (2014). Root hydraulics: The forgotten side of roots in drought adaptation. *Field Crops Research*, 165, 15–24. <https://doi.org/10.1016/j.fcr.2014.03.017>
- Wasson, A., Bischof, L., Zwart, A., & Watt, M. (2016). A portable fluorescence spectroscopy imaging system for automated root phenotyping in soil cores in the field. *Journal of Experimental Botany*, 67(4), 1033–1043. <https://doi.org/10.1093/jxb/erv570>
- Wasson, A. P., Richards, R. A., Chatrath, R., Misra, S. C., Prasad, S. V. S., Rebetzke, G. J., Kirkegaard, J. A., Christopher, J., & Watt, M. (2012). Traits and selection strategies to improve root systems and water uptake in water-limited wheat crops. *Journal of Experimental Botany*, 63(9), 3485–3498. <https://doi.org/10.1093/jxb/ers111>
- Xie, Q., Mayes, S., & Sparkes, D. L. (2015). Carpel size, grain filling, and morphology determine individual grain weight in wheat. *Journal of Experimental Botany*, 66(21), 6715–6730. <https://doi.org/10.1093/jxb/erv378>
- Zadoks, J. C., Chang, T. T., & Konzak, C. F. (1974). A decimal code for the growth stages of cereals. *Weed Research*, 14(6), 415–421. <https://doi.org/10.1111/j.1365-3180.1974.tb01084.x>
- Zamani, A., Emam, Y., & Edalat, M. (2024). Response of bread wheat cultivars to terminal water stress and cytokinin application from a grain phenotyping perspective. *Agronomy*, 14(1), 182. <https://doi.org/10.3390/agronomy14010182>
- Zampieri, M., Ceglar, A., Dentener, F., & Toreti, A. (2017). Wheat yield loss attributable to heat waves, drought and water excess at the global, national and subnational scales. *Environmental Research Letters*, 12(6), 64008. <https://doi.org/10.1088/1748-9326/aa723b>