

Effect of Storage Time and Concentration of Aflatoxin M₁ on Toxin Binding Capacity of *L. acidophilus* in Fermented milk Product

N. Adibpour¹, S. Soleimanian-Zad^{1,2*}, M. Sarabi-Jamab³, and F. Tajalli⁴

ABSTRACT

Aflatoxins are potent carcinogenic and immunosuppressive agents. Acute exposure to high level of aflatoxins leads to aflatoxicosis, which cause rapid death due to liver failure. Immune modulating effects of probiotic bacteria have good prospects to detoxification of natural foods. This study was aimed to investigate the ability of *Lactobacillus acidophilus* strain LA-5 in the presence and absence of yoghurt starter culture for removing Aflatoxin M₁ (AFM₁) in comparison with yoghurt starter cultures (10⁸ CFU ml⁻¹). AFM₁ detoxification was evaluated for 21 days of yoghurt storage at 4°C at different concentrations of Aflatoxin (0.1, 0.5 and 0.75 µg L⁻¹). The amounts of unbound AFM₁ were determined using competitive Enzyme-Linked Immunosorbent Assay (ELISA). *L. acidophilus* combined with yoghurt starter culture and alone could significantly (P ≤ 0.05) remove AFM₁ compared to control group. The results indicated that increasing initial AFM₁ concentration in the yoghurt samples and storage time affected the capacity of AFM₁ binding.

Keywords: Aflatoxin M₁, Biological detoxification, Enzyme-linked immunosorbent assay, Lactic acid bacteria, Yoghurt.

INTRODUCTION

Mycotoxins are secondary metabolites produced by mycelia or spores of filamentous fungi (González *et al.*, 2001). Aflatoxins are one of the most carcinogenic substances known until now (Nierman *et al.*, 2008). Various food resources may be contaminated by aflatoxins such as corn, peanuts, cotton seeds, rice, pistachio, almonds, chestnuts, pumpkin seeds, as well as other oily seeds and sorghum (Chu, 1991; Tajkarimi *et al.*, 2007). The changing rate of

ingested Aflatoxin B₁ (AFB₁) to AFM₁ is highly variable, ranging from 0.3 to 6.2%. There is a linear relationship between the AFM₁ concentration in milk and of AFB₁ in contaminated feeds consumed by the livestock (Bakirci, 2001; Creppy, 2002; Mohamadi and Alizadeh, 2010). Chronic exposure to low levels of aflatoxins may threaten the public health followed by serious economic burdens (Oliveira and Germano, 1997). The International Agency for Research on Cancer (IARC, 2002) classifies AFM₁ as Group 1 that leads to human cancer; however, AFM₁ is about 10

¹ Department of Food Science and Technology, College of Agriculture, Isfahan University of Technology, Isfahan, 84156-83111, Islamic Republic of Iran.

² Research Center, Institute of Biotechnology and Bioengineering, Isfahan University of Technology, Isfahan, Islamic Republic of Iran.

* Corresponding author; email: soleiman@cc.iut.ac.ir

³ Research Institute of Food Science and Technology (RIFST), Department of Food Biotechnology, Mashhad, Islamic Republic of Iran.

⁴ Food Quality and Safety Research Department, Iranian Academic Center for Education, Culture and Research (ACECR)-Mashhad Branch, Mashhad, Islamic Republic of Iran.



times less carcinogenic than AFB₁. Since AFM₁ is detected frequently in milk and other dairy products that are commonly consumed (especially in countries that advanced ranching industry does not exist), a wide variety of methods were in order to control and decrease AFM₁ contamination of foods and feeds. Elimination of aflatoxin with chemical and physical methods have some disadvantages which limit their use. For example, insufficiency of toxin elimination, high costs and losing the nutritional value of the product (Line and Brackett, 1995; El-Nezami *et al.*, 1998). Biodegradation of aflatoxins by microorganisms offers an attractive alternative for the control, reduction or elimination of aflatoxins to maintain their quality and safety (Alberts *et al.*, 2009). Among all types of available microorganisms that may be utilized to remove aflatoxins from a contaminated medium, Lactic Acid Bacteria (LAB) would be a suitable choice for reducing the bioavailability of aflatoxins because of their unique characteristics. They are Generally Recognized As Safe (GRAS) by USFDA, also some of them confer beneficial effects on health which are called probiotics (El-Nezami *et al.*, 2002; Fuchs *et al.*, 2008). This study was carried out to investigate the ability of yoghurt starter culture and *Lactobacillus acidophilus* LA-5 for removing AFM₁ from contaminated probiotic yoghurt.

MATERIALS AND METHODS

Culture Preparation

The Direct Vat Set (DVS) lyophilized pouches of yoghurt starter culture (YoFlex) which contain *Streptococcus thermophilus* and *Lactobacillus delbrueckii* ssp. *bulgaricus*, also, *L. acidophilus* LA-5[®] as a probiotic strain were originally obtained from Chr. Hansen's Lab (Denmark). The lyophilized cultures were maintained at -20°C until use. Both bacterial cultures (1 g

per 100 ml) were inoculated directly into MRS broth (MerckTM, Germany) and incubated at 37°C for 24 hours. After centrifuging (3,400×g at 4°C for 10 minutes), the bacterial pellets were collected and washed three times with 20 ml Phosphate Buffer Saline (PBS) (pH= 7.4). To achieve cell count of 10⁸ CFU ml⁻¹, the Optical Density (OD_{600 nm}) of bacterial suspension in PBS was adjusted to a McFarland standard (No. 1) by a spectrophotometer (Martin and Palomino, 2009). After centrifugation, 20 ml of PBS was drained gently from each bacterial suspension and 30 ml of contaminated milk was replaced.

Growth of Starter Culture and Probiotic Bacteria

The bacterial cell count was determined using traditional plate counting at MRS agar (MerckTM, Germany) for the starter culture alone (Y), a combination of starter culture and *L. acidophilus* LA-5 (AY), also *L. acidophilus* alone (A). To assess the viability of *L. acidophilus* in the presence of starter culture (AY), according to Ashraf and Shah (2011), Ox-bile (0.15% v/v) (Fluka, Sigma-AldrichTM, Germany) was added into the MRS medium. All samples were incubated at 37°C for 72 hours, aerobically.

Milk Contamination with AFM₁ and Yoghurt Manufacturing

Aflatoxin M₁ (from *Aspergillus flavus*, 10 µg) was purchased from Sigma-AldrichTM (Germany). AFM₁ stock solution (10 µg ml⁻¹) was prepared by dissolving the entire powder in 1 ml of high-performance liquid chromatography grade acetonitrile (SigmaTM Chemical Co. Ltd., USA). The concentration was verified by HPLC method according to the Institute of Standards and Industrial Research of Iran (ISIRI-7133, 2011). Working solution (100 µg L⁻¹) was prepared accurately by pure acetonitrile solution as a

diluent and samples were stored at -20°C until use. Reconstituted milk was prepared by diluting skimmed milk powder (Code-15363; Merck™, Germany) in distilled water. Before adding AFM₁ to reconstituted milk, a portion of that was set apart as negative control. For yoghurt preparation, reconstituted milk was heated at 90-95°C for 5 minutes then cooled to 42°C. The pasteurized milk was contaminated with AFM₁ working solution at three different levels (0.1, 0.5 and 0.75 µg L⁻¹). The bacterial pellets were then inoculated into the contaminated milk and incubated at 42-45°C for 4 hours in order for the yoghurt to be set by the starter cultures (control group), also, 6-7 hours for yoghurt made by *L. acidophilus* La-5 alone. The prepared yoghurts were stored at 4°C for 21 days and samples were taken at 7 days intervals to determine unbound AFM₁.

Measurement of pH

The pH value of yoghurt samples were measured throughout the experiment by pH meter (Jenway™, UK) during a 21 day period (at 1, 7, 14 and 21 days).

Analysis of AFM₁ in Samples by Competitive ELISA

The yoghurt samples were centrifuged (3,400×g at 4°C for 5 minutes) at the end of each storage period (at days 1, 7, 14 and 21) and unbound AFM₁ content of the supernatants were determined by ELISA method. ELISA procedure was performed according to instructions provided by EuroProxima. One-hundred microliters of standard solutions and prepared samples were added into separate microtiter wells (pre-coated with anti-aflatoxin M₁) and incubated at room temperature (25°C) for 60 minutes in a dark environment. Next, the liquid was poured out and the wells were washed three times with washing buffer (300 µl) by microplate strip washer (ELx50;

Bio-Tek Instruments, USA). Then, 100 µl of the diluted enzyme conjugate was added to the wells, mixed gently by shaking the plate manually and incubated at room temperature for 30 minutes. Again, the wells were washed three times with washing buffer. After that, 100 µl of substrate/chromogen was added, mixed gently by hand and incubated in a dark place at room temperature for 30 minutes. Finally, 100 µl of the stop reagent was mixed by the wells contents and the absorbance was measured at $\lambda_{max}=450$ nm using ELISA plate reader (ELX808; Bio-Tek Instruments, USA). According to 5121AFM guidelines, the limit of detection (LOD) for the milk is < 0.006 ng ml⁻¹ and < 10 pg ml⁻¹ for cheese.

Quantification of Residual AFM₁ in Supernatant Samples by High-Performance Liquid Chromatography (HPLC)

In this study, the number of each toxin concentration of all the samples tested by ELISA kit, were randomly selected to confirm by High Performance Liquid Chromatography method according to ISIRI (2011). The linearity was evaluated by linear regression analysis using the least squares method and expressed as correlation coefficient (R²).

The method is based on the immunoaffinity clean-up of the milk samples followed by the determination of the AFM₁ content by HPLC as follows:

At first, the fat of yoghurt samples were separated by centrifugation. Then immunoaffinity column that contains monoclonal antibodies to aflatoxin M₁ bound to a solid support, was applied for purification of defatted samples. The maximum volume of the affinity column shouldn't be less than 10 ng AFM₁ and the recovery rate shouldn't be lower than 70%. 50 µl of the reconstituted samples were injected in the HPLC using Waters 474 fluorescence detector at 360 and 440 nm for excitation and emission, respectively.



The chromatography was carried out with Water HPLC system with Waters Alliance 2695 HPLC pump. The column and guard column used were 4.6×200 mm reverse phase ODS-5 μm C18 column (Phenomenex, USA) and OnyxTM Monolithic C18 with 10×4.6 mm LC guard cartridge, respectively. The mobile phase was composed of methanol and water (40:60 v/v). The flow rate of the injected sample was 2.0 ml per minute to achieve the optimum resolution of aflatoxin. Based on aflatoxin standards injected into the device, the retention time of aflatoxin M₁ in samples was 3.69 minutes. The Limit Of Detection (LOD) is defined as the lowest amount reproducibly detected with at least 3:1 (signal to noise ratio) and in this method LOD was 0.01 ng ml⁻¹ and the limit of quantitation was 0.03 ng ml⁻¹.

Statistical Analysis

All experiments were performed quadruplicate and the presented data are their means. Statistical analysis was carried out with IBM SPSS StatisticsTM 20 software. Significant differences between the means

were estimated by ANOVA and Duncan's tests at $P \leq 0.05$. All graphs were generated using Microsoft ExcelTM software.

RESULTS AND DISCUSSION

Survival of Yoghurt Bacteria and *L. Acidophilus* La-5

Enumeration of *L. acidophilus* was done during 21 days by 7-day intervals during the refrigerated storage by standard plate counting on MRS-bile medium, when a combination of starter culture and probiotic strain were cultivated, and on MRS agar when probiotic strain grew alone. The number of lactic acid bacteria and yoghurt starter culture were showed in Figure 1. The initial viable cell counts of starter cultures and *L. acidophilus* La-5 both were 3×10^8 CFU g⁻¹ immediately after yoghurt manufacturing before keeping in refrigerator. Survival of *L. acidophilus* La-5 in yoghurt in the absence of starter cultures remained stable throughout the storage period until day 14. From day 14 to 21, just 1 log cycle of cell count reduction was observed ($P \leq 0.05$). Survival of La-5 in the

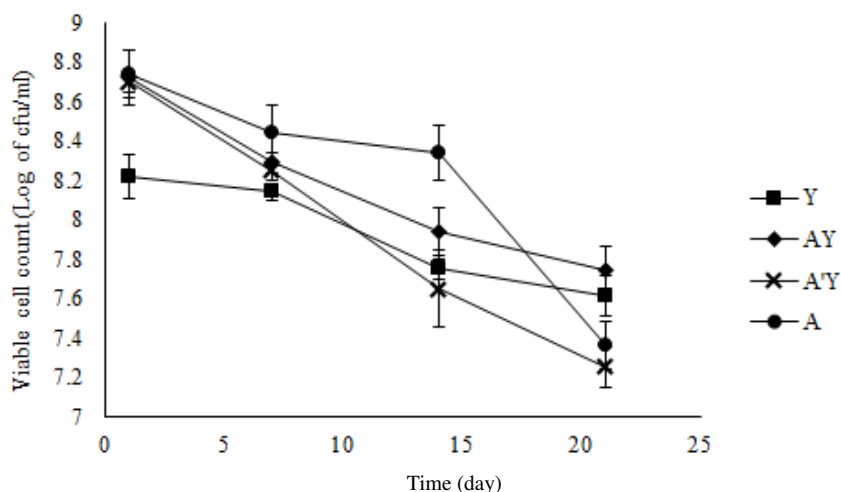


Figure 1. Enumeration of yoghurt starter culture and *L. acidophilus* La-5 survived, during 21 days of storage period at 7-day intervals. Data shown are representatives of quadruplicate. Starter culture alone (Y); combination of starter culture and *L. acidophilus* La-5 (AY); *L. acidophilus* La-5 separated from AY samples (A^Y), and *L. acidophilus* La-5 alone (A).

presence of starter culture showed 1.5 log cycle of cell count reduction throughout the 21 days of storage.

Acid and osmotic stress, as consequences of lactic acid production and application of food additives, are the most predominant stress factors during yoghurt manufacture and cold storage (Settachaimongkon *et al.*, 2015).

Many studies have reported low pH or the accumulation of organic acids, especially lactic acid which is mainly produced by *L. delbrueckii* ssp. *bulgaricus*, as one of the most influential factors which reduces the viability of probiotic cells in the products, (Kailasapathy and Chin, 2000; Lourens-Hattingh and Viljeon, 2001). Some scientists explained that acid can passively diffuse through the bacterial cell membrane and rapidly dissociate into protons inside the cytoplasm, then charge derivatives to which the cell membrane is impermeable causing an internal acidification that disorders the activity of acid sensitive enzymes, DNA and damages proteins. Thus, it is essential to monitor these qualifications during the storage time in order to favor bacterial growth (Bovo *et al.*, 2014). In this study, the pH of all three different yoghurt samples were declined to 4.5 after 21 days storage (unpublished data). It is reported that the least tolerable pH is 2.5 for *L. acidophilus* (Zhao *et al.*, 2012). The survival of *L. acidophilus* in acidic environments has been studied, and this species proved to be highly resistant to acid (Shah, 2000). Lorca and de Valdez (2001a; 2001b) expressed survival of *L. acidophilus* may be affected by physiological adaptation known as Acid Tolerance Response (ATR). Fundamental mechanisms of acid tolerance utilized by gram-positive bacteria include proton pumps, proteins involved in repair or degradation of damaged cell components, activity of arginine deaminase that cause increase of alkalinity of cytoplasm, urease and glutamine decarboxylase, and conversions in the composition of the cell envelope (Cotter *et al.*, 2001; Cotter and Hill, 2003; De Angelis and Gobbetti, 2004;

Ruiz *et al.*, 2011). In the presence of organic acids, the F_1F_0 -ATPase plays an important role in maintaining the intracellular pH (pHi). According to Tamime *et al.* (2005) and Demers-Mathieu *et al.* (2015), *L. delbrueckii* ssp. *bulgaricus* is known for the ability of post-acidification process and high production of hydrogen peroxide (H_2O_2) that have an impact on the growth of probiotic strains. As a result of lacking catalase, *L. acidophilus* is subjected to oxidative stress and this may damage the proteins and DNA of the cells and eventually kill them.

Changes of AFM₁ during Yoghurt Storage

The aflatoxin-binding capacity of different strains tested at 4°C during 21 days of storage are displayed in Figure 2. Yoghurt starter culture and *L. acidophilus* tested in this study were able to bind AFM₁. The significant difference ($P \leq 0.05$) between AFM₁ binding ability of *L. acidophilus* LA-5 and starter culture in yoghurt was demonstrated.

Preliminary investigations have expressed that yoghurt starter culture and probiotic bacteria could be used to remove AFM₁ from food and feed. Sarimehmetoğlu and Küplülü (2004) reported that *S. thermophilus* ST-36 (29.42–36.16%) has a great potential to bind a high percentage of AFM₁ in comparison with *L. delbrueckii* ssp. *bulgaricus* CH-2 (18.7–27.56%) in PBS and milk, respectively ($P < 0.01$). Elgerbi *et al.* (2006) assessed the ability of strains of *Lactobacillus* spp., *Lactococcus* spp. and *Bifidobacterium* spp. to bind the AFM₁ in buffered aqueous solution. They found that the percentage of AFM₁ bound by these strains ranged from 4.5-73.1% after 96 hours. El-Khoury *et al.* (2011) found that the yogurt bacteria, *L. bulgaricus*, *Str. thermophilus* and a combination of these two bacterium reduced AFM₁ content of milk to 58.5, 37.7 and 46.7% respectively, after incubation at 37°C for 6 hours. Bovo *et al.* (2012) evaluated the ability of some

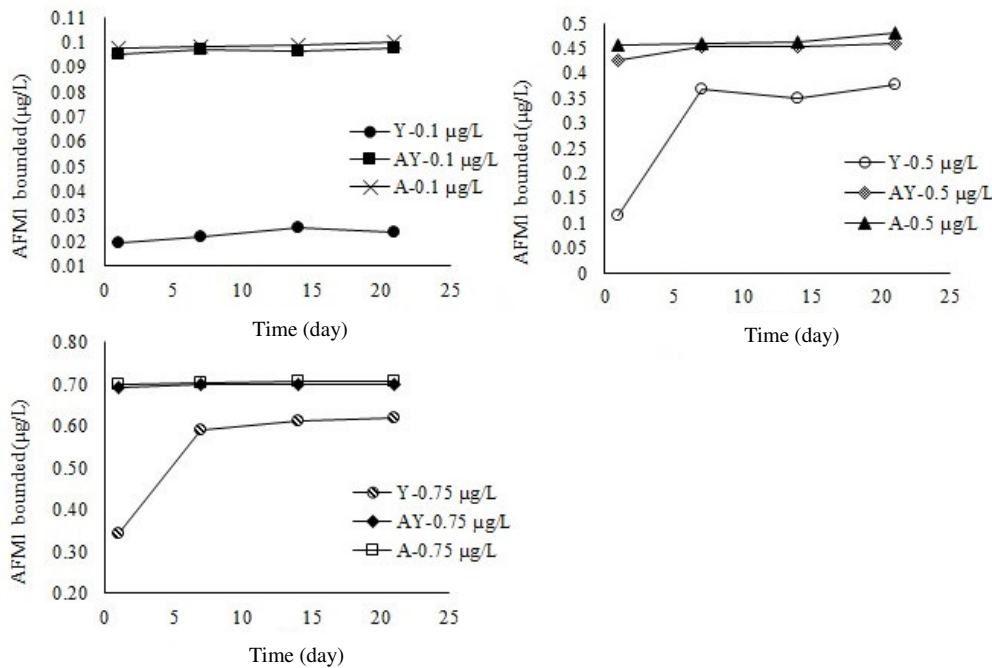


Figure 2. AFM₁ removal at three different levels of toxin in yoghurt during 21 days of storage in refrigerator with starter culture and *L. acidophilus*. Data shown are representatives of quadruplicate experiments. Means with the same letter for each types of yoghurt do not differ significantly at $P \leq 0.05$. Yoghurt made with starter culture (Y); probiotic yoghurt made with a combination of *L. acidophilus* LA-5 and starter culture (AY), and Fermented milk made with *L. acidophilus* La-5 alone (A).

probiotic strains to remove AFM₁ in skimmed milk and reported that the tested strains bound AFM₁ within a range from 13.51 to 37.75% for 15 minutes at 37°C. Serrano-Niño *et al.* (2013) assessed the ability of some species of probiotic bacteria and reported that the bioaccessibility of AFM₁ reduced in range of 22.72 to 45.17% in the presence of the tested strains.

Decrease of AFM₁ levels in yoghurt might be assigned to some factors such as low pH, formation of organic acids or other fermentation by-products (Govaris *et al.*, 2002). Reduction of pH during the fermentation alters the structure of caseins in milk proteins. These changes lead to the formation of a network like yoghurt gel which hold the aflatoxin inside the precipitate (Montazeri *et al.*, 2014).

The results of this research showed a significant reduction in unbound AFM₁ content through the storage time. Analysis of the data in Figure 2 indicated that the

binding of AFM₁ is a strain specific characteristic. At the first day of storage, *L. acidophilus* La-5 removed over 90% of the AFM₁ from the yoghurt samples. Then, until the end of the storage time a significant reduction ($P \leq 0.01$) in the amount of unbound aflatoxin was observed.

These results were in good agreement with previous reports (Elgerbi *et al.*, 2006; Biernasiak *et al.*, 2006). Motawee and El-Ghany (2011) evaluated the ability of eight dairy strain of lactic acid bacteria to remove aflatoxin M₁ and B₁ in yoghurt and noted that for all examined starters, the percentage of aflatoxins AFM₁ and AFB₁ reduction in yoghurt after 5 hours was considerably less than that at the end of storage period. Contrary to our observations, other authors found no reduction of AFM₁ in yoghurt during the cooled storage period Blanco *et al.*, 1993; Iha *et al.*, 2013). Factors such as toxin concentration, temperature of storage, time

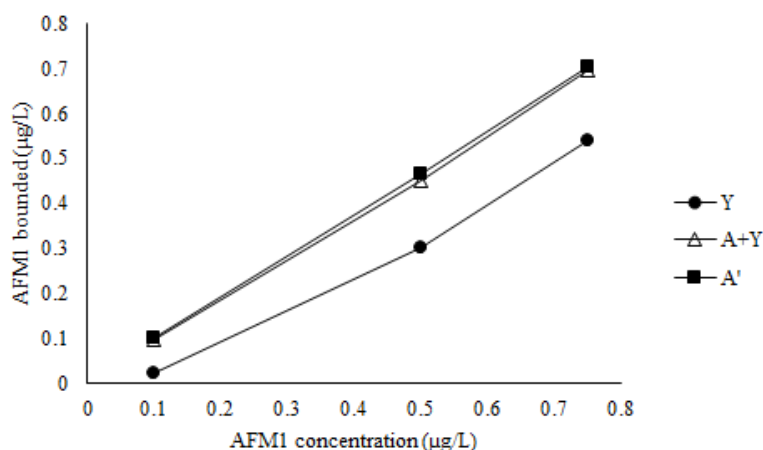


Figure 3. Effect of AFM₁ levels on toxin-binding capacity of yoghurt starter culture and *L. acidophilus* La-5. (Y) Yoghurt made with starter culture; (AY) Probiotic yoghurt made with a combination of *L. acidophilus* LA-5 and starter culture, and (A) Fermented milk made with *L. acidophilus* La-5 alone.

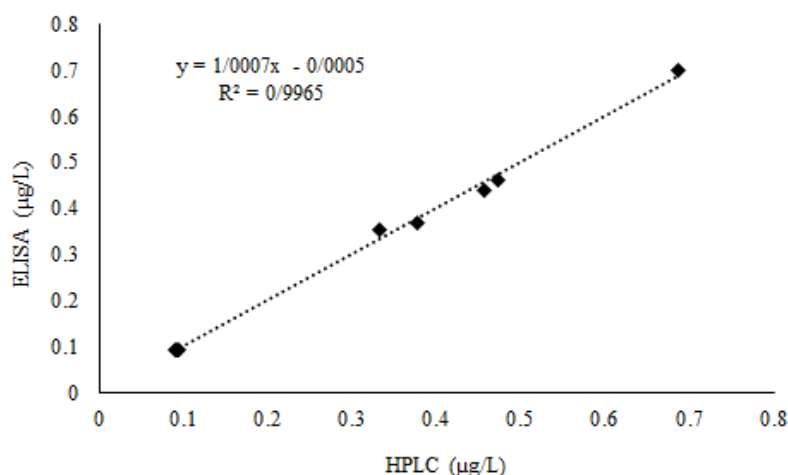


Figure 4. The correlation between HPLC and ELISA data for AFM₁ determination. Axes *x* and *y* show level of AFM₁ bounded which have been determined independently by ELISA and HPLC method, respectively.

elapsed before sample analysis, the difference in type of starter cultures used to make dairy product, variability in milk composition and milk contamination method may cause differences in results (Ismail *et al.*, 1989; Mohammadi *et al.*, 2009). The reduction of AFM₁ in yoghurt during storage period might be due to the oxidation of glucose that produces gluconolactone and hydrogen peroxide which will be distributed in yoghurt. H₂O₂ can form single reactive oxygen which may react with the double bond in the terminal of dihydrofuran moiety of the aflatoxin

molecule (Elsanhoty *et al.*, 2014). Some authors showed that non-viable cells could also remove higher amounts of aflatoxin from different media (Pierides *et al.*, 2000; Shahin, 2007; Kabak and Var, 2008; Bovo *et al.*, 2014). Therefore, increasing non-viable cell contents during the yoghurt storage as a result of pH value reduction or increase of bacterial second metabolite in a fatal overdose case, may help to remove more aflatoxin from media.

Analysis of our data indicated that *L. acidophilus* La-5 removed over 90% of the



AFM₁. This observation confirmed that La-5 could bind the AFM₁ quickly which was in agreement with El-Nezami *et al.* (1998a) for *L. rhamnosus* strains GG and LC705. At the time of adding LAB, both strains removed 80% of the AFB₁ approximately. Some authors suggested that the significant differences existing among aflatoxin binding ability of lactic acid bacteria depends on distinctive cell-wall structure (El-Nezami *et al.*, 1998b; Pierides *et al.*, 2000; Peltonen *et al.*, 2001; Lahtinen *et al.*, 2004; Zinedine *et al.*, 2005; Hernandez-Mendoza *et al.*, 2009). Strength of mycotoxin-LAB is a fast physicochemical interaction between the toxin and the functional groups of the cell surface (Bovo *et al.*, 2014; Zoghi *et al.*, 2014), and it is based on physical adsorption, ion exchange, and complexation, regardless of the bacterial metabolism. It was proposed that carbohydrate components of the bacteria cell walls particularly peptidoglycans were probably the compounds which were in charge of binding aflatoxin to the bacterial surface. Hernandez-Mendoza *et al.* (2009) indicated that except the peptidoglycans, teichoic acids were also an important part of the cell wall which could bind aflatoxin. Binding to macromolecules are functions of fibril network of teichoic acids and polysaccharides; and the stability and strength of binding of microorganism to toxins depends on strain, amino acid composition of peptidoglycan structure and environmental conditions (Zoghi *et al.*, 2014).

By increasing AFM₁ concentration, AFM₁ binding ability of yoghurt starter culture was increased (Figure 3). However, *L. acidophilus* La-5 in presence and also in the absence of yoghurt starter culture significantly showed further reduction in AFM₁ content at all concentrations tested compared to the control group ($P \leq 0.05$).

In this study, accuracy of the ELISA method for detecting AFM₁ in samples was verified by HPLC and the correlation between ELISA and HPLC methods were evaluated and are shown in Figure 4. The correlation coefficient (R^2) between these two methods was 0.9965. Therefore, the ELISA method can be used as a reliable and cheaper method to evaluate the

level of aflatoxin in milk and even in animal husbandry.

In our research, the AFM₁ binding ability of yoghurt starter culture and *L. acidophilus* La-5 were increased, by increasing initial AFM₁ concentration. Our results were supported by some studies showing that the amount of bound AFM₁ by bacteria in milk and PBS is raised with the increase of AFM₁ concentration (Rašić *et al.*, 1991; Kabak and Var, 2008).

CONCLUSIONS

Since milk and dairy products are an important part of people's daily food basket of the world, aflatoxin contamination of these products and human body's inability to reduce or eliminate these toxins, can endanger the health of a large number of people in the world. This study was an attempt to show the capability of *L. acidophilus* La-5 as a biological and safe method to reduce aflatoxin in dairy products. Also the assessment of storage time and increasing initial toxin level in products showed significant increase in toxin binding to bacterial cell wall and eliminating it from products. Favorable survival of *Lactobacillus acidophilus* La-5 in the product during storage indicated that this bacterium as a probiotic strain can not only reduce the level of aflatoxin contamination but also incorporate to the production of a probiotic product at the same time.

ACKNOWLEDGEMENTS

The authors would like to acknowledge the Research Institute of Food Science and Technology (RIFST), Department of Food Biotechnology, Mashhad, Iran and Food Quality and Safety Research Department, Iranian Academic Centre for Education, Culture and Research (ACECR)-Mashhad Branch for financial support of this research. The authors declare that there is no conflict of interests.

REFERENCES

1. Alberts, J. F., Gelderblomb, W. C. A., Botha, A. and Van Zyl, W. H. 2009. Degradation of Aflatoxin B₁ by Fungal Laccase Enzymes. *Int. J. Food Microbiol.*, **135(1)**: 47-52.
2. Ashraf, R., Shah, N. P. 2011. Selective and Differential Enumerations of *Lactobacillus delbrueckii* subsp. *bulgaricus*, *Streptococcus thermophilus*, *Lactobacillus acidophilus*, *Lactobacillus casei* and *Bifidobacterium* spp. in Yoghurt: A Review. *Int. J. Food Microbiol.*, **149(3)**: 194-208.
3. Bakirci, I. 2001. A Study on the Occurrence of Aflatoxin M₁ in Milk and Milk Products Produced in Van Province of Turkey. *Food Control.*, **12**: 47-51.
4. Biernasiak, J., Piotrowska, M. and Libudzisz, Z. 2006. Detoxification of Mycotoxins by Probiotic Preparation for Broiler Chickens. *Mycotoxin Res.*, **22(4)**: 230-235.
5. Blanco, J. L., Beatriz, A. C., Nuria, L., Susana, D., Marta, E., Garcia, L. and Suarez, G. 1993. Behavior of Aflatoxins during Manufacture and Storage of Yoghurt. *Milchwissenschaft*, **48(7)**: 385-387.
6. Bovo, F., Corassin, C. H., Rosim, R. E. and Oliveira, C. A. F. 2012. Efficiency of Lactic Acid Bacteria Strains for Decontamination of Aflatoxin M₁ in Phosphate Buffer Saline Solution and in Skim Milk. *Food Bioprocess Tech.*, **5(1)**: 1-5.
7. Bovo, F., Franco, L. T., Rosim, R. E. and Oliveria, C. A. F. 2014. Ability of a *Lactobacillus rhamnosus* Strain Cultured in Milk Whey Based Medium to Bind Aflatoxin B₁. *Food Sci. Technol. (Campinas)*, **34(3)**: 566-570.
8. Cotter, P. D. and Hill, C. 2003. Surviving the Acid Test: Responses of Gram-positive Bacteria to Low pH. *Microbiol. Mol. Biol. Rev.*, **67(3)**: 429-453.
9. Cotter, P. D., Gahan, C. G. and Hill, C. 2001. A Glutamate Decarboxylase System Protects *Listeria monocytogenes* in Gastric Fluid. *Mol. Microbiol.*, **40(2)**: 465-475.
10. Creppy, E. E. 2002. Update of Survey, Regulation and Toxic Effects of Mycotoxins in Europe. *Toxicol. Lett.*, **127(1-3)**: 19-28.
11. Chu, F. S. 1991. Mycotoxins: Food Contamination, Mechanism, Carcinogenic Potential and Preventive Measures. *Mutation Res.*, **259**: 291-306.
12. De Angelis, M. and Gobbetti, M. 2004. Environmental Stress Responses in *Lactobacillus*: a Review. *Proteomics*, **4(1)**: 106-122.
13. Demers-Mathieu, V., Audy, J., Laurin, E., Fliss, I. and St-Gelais, D. 2015. Impact of Commercial Mesophilic and Thermophilic Starters on the Growth of New Probiotic Isolates. *Int. Dairy J.*, **45**: 31-40.
14. Donkor, O. N., Henriksson, A., Vasiljevic, T. and Shah, N. P. 2006. Effect of Acidification on the Activity of Probiotics in Yogurt during Cold Storage. *Int. Dairy J.*, **16(10)**: 1181-1189.
15. Elgerbi, A. M., Aidoo, K. E., Candlish, A. A. G. and Williams, A. G. 2006. Effects of Lactic Acid Bacteria and Bifidobacteria on Levels of Aflatoxin M₁ in Milk and Phosphate Buffer. *Milchwissenschaft*, **61(2)**: 197-199.
16. El-Khoury, A., Atoui, A. and Yaghi, J. 2011. Analysis of Aflatoxin M₁ in Milk and Yoghurt and AFM₁ Reduction by Lactic Acid Bacteria Used in Lebanese Industry. *Food Control.*, **22(10)**: 1695-1699.
17. El-Nezami, H. S., Chrevatidis, A., Auriola, S., Salminen, S. and Mykkänen, H. 2002. Removal of Common Fusarium Toxins *In vitro* by Strains of *Lactobacillus* and *Propionibacterium*. *Food Addit. Contam.*, **19**: 680-686.
18. El-Nezami, H., Kankaanpaa, P., Salminen, S. and Ahokas, J. 1998a. Physicochemical Alterations Enhance the Ability of Dairy Strains of Lactic Acid Bacteria to Remove Aflatoxin from Contaminated Media. *J. Food Prot.*, **61(4)**: 466-468.
19. El-Nezami, H., Kankaanpaa, P., Salminen, S. and Ahokas, J. 1998b. Ability of Dairy Strains of Lactic Acid Bacteria to Bind Food Carcinogens. *Food Chem. Toxicol.*, **36(4)**: 321-326.
20. Elsanhoty, R. M., Salam, S. A., Ramadan, M. F. and Badr, F. H. 2014. Detoxification of Aflatoxin M₁ in Yoghurt Using



- Probiotics and Lactic Acid Bacteria. *Food Control.*, **43**: 129-134.
21. Fuchs, S., Sontag, G., Stidl, R., Ehrlich, V., Kundi, M. and Knasmüller, S. 2008. Detoxification of Patulin and Ochratoxin A, Two Abundant Mycotoxins, by Lactic Acid Bacteria. *Food Chem. Toxicol.*, **46(4)**: 1398-1407.
 22. Govaris, A., Roussi, V., Koidis, P. A. and Botsoglou, N. A. 2002. Distribution and Stability of Aflatoxin M₁ during Production and Storage of Yoghurt. *Food Addit. Contam.*, **19(11)**: 1043-1050.
 23. Gonçalves, E., Pinto, M. M. and Felício, J. D. 2001. Análise de Micotoxinas no Instituto Biológico de 1989 Biológico a 1999. *Divulgação Técnica*, **63(1/2)**: 15-19.
 24. Hernandez-Mendoza, A., Guzman-de-Pena, D. and Garcia, H. S. 2009. Key Role of Teichoic Acids on Aflatoxin B₁ Binding by Probiotic Bacteria. *J. Appl. Microbiol.*, **107(2)**: 395-403.
 25. IARC. 2002. *Monograph on the Evaluation of Carcinogenic Risk to Humans*. World Health Organization, IARC, Lyon, France, **82**: 171.
 26. Iha, M. H., Barbosa, C. B., Okada, I. A. and Trucksess, M. W. 2013. Aflatoxin M₁ in Milk and Distribution and Stability of Aflatoxin M₁ during Production and Storage of Yoghurt and Cheese. *Food Control.*, **29(1)**: 1-6.
 27. ISIRI. 2011. Milk and Milk Products: Determination of Aflatoxin M₁ by HPLC Method and Immunoaffinity Column Clean up Test Method. Iranian National Standard 7133. 1st Revision, Institute of standards and Industrial Research of Iran, April 2011, Karaj, Iran.
 28. Ismail, A. A., Tawfek, N. F., Abd-Alla, E. A., El-Dairouty, M. and Sharaf, R. K. 1989. Fate of Aflatoxin M₁ during Kefir Processing and Its Effect on the Microflora and the Chemical Structure. *Dtsch. Lebensmitt. Rundsch.*, **85(3)**: 76-78.
 29. Kabak, B. and Var, I. 2008. Factors Affecting the Removal of Aflatoxin M₁ from Food Model by *Lactobacillus* and *Bifidobacterium* Strains. *J. Environ. Sci. Health Part B*, **43(7)**: 617-624.
 30. Kailasapathy, K. and Chin, J. 2000. Survival and Therapeutic Potential of Probiotic Organisms with Reference to *Lactobacillus acidophilus* and *Bifidobacterium* spp. *Immunol. Cell Biol.*, **78**: 80-88.
 31. Lahtinen, S. J., Haskard, C. A., Ouwehand, A. C., Salminen, S. J. and Ahokas, J. T. 2004. Binding of Aflatoxin B₁ to Cell Wall Components of *Lactobacillus rhamnosus* Strain GG. *Food Addit. Contam.*, **21(2)**: 158-164.
 32. Line, J. E. and Brackett, R. E. 1995. Factors Affecting Aflatoxin B₁ Removal by *Flavobacterium aurantiacum*. *J. Food Prot.*, **58(1)**: 91-94.
 33. Lorca, G. L. and de Valdez, G. F. 2001a. A Low-pH Inducible, Stationary-phase Acid Tolerance Response in *Lactobacillus acidophilus* CRL639. *Curr. Microbiol.*, **42**: 21-25.
 34. Lorca, G. L. and de Valdez, G. F. 2001b. Acid Tolerance Mediated by Membrane ATPases in *Lactobacillus acidophilus*. *Biotech. Lett.* **23**: 777-780.
 35. Lourens-Hattingh, A. and Viljeon, C. B. 2001. Yoghurt as Probiotic Carrier Food. *Int. Dairy J.*, **11(1)**: 1-17.
 36. Martin, A. and Palomino, J. C. 2009. Nitrate Reductase Assay: Drug Susceptibility Testing for *Mycobacterium Tuberculosis*. Institute of Tropical Medicine, Mycobacteriology Unit, Antwerp, Belgium. Procedure Manual, version 2.
 37. Mohammadi, H., Alizadeh, M., Bari, M. R., Khosrowshahi, A. and Tadjik, H. 2009. Optimization of the Process Variable for Minimizing of the Aflatoxin M₁ Content in Iranian White Brine Cheese. *J. Agr. Sci. Tech.*, **11**: 181-190.
 38. Mohamadi, H. and Alizadeh, M. 2010. A Study of the Occurrence of Aflatoxin M₁ in Dairy Products Marketed in Urmia, Iran. *J. Agr. Sci. Tech.*, **12**: 579-583.
 39. Montazeri, H., Arjmandtalab, S., Dehghanzadeh, G., Karami, S., Razmjoo, M. M., Sayad, M. and Oryan, A. 2014. Effect of Production and Storage of Probiotic yogurt on aflatoxin M₁ residue. *J. Food Qual. Hazard. Control.*, **1(1)**: 7-14.
 40. Motawee, M. M. and Abd El-Ghany, M. A. 2011. Effect of Some Lactic Acid Bacteria Strains on Aflatoxins Reduction in Some Dairy Foods. *The 6th Arab and 3rd International Annual Scientific Conference on: Development of Higher Specific Education Programs in Egypt and*

- the Arab World in the Light of Knowledge Era Requirements.*
41. Nierman, W. C., Cleveland, T. E., Payne, G. A., Keller, N. P., Campbell, B. C., Bennett, J. W., Guo, B., Yu, J. and Robens, J. F. 2008. Mycotoxin Production and Prevention of Aflatoxin Contamination in Food and Feed. In: "The *Aspergilli: Genomics, Medical Aspects, Biotechnology and Research Methods*", (Eds.): Goldman G. H. and Osmani, S. A.. CRC Press, Boca Raton, PP. 457-472.
 42. Oliveira, C. A. F. and Germano, P. M. L. 1997. Aflatoxinas: Conceitos Sobre Mecanismos Detoxicidade e seu Envolvimento na Etiologia do Câncer Hepático Celular. *Rev. Saúde. Pública.*, **31(4)**: 417-424.
 43. Peltonen, K., El-Nezami, H., Haskard, C., Ahokas, J. and Salminen, S. 2001. Aflatoxin B₁ Binding by Dairy Strains of Lactic Acid Bacteria and Bifidobacteria. *J. Dairy Sci.*, **84(10)**: 2152-2156.
 44. Pierides, M., El-Nezami, H., Peltonen, K., Salminen, S. and Ahokas, J. 2000. Ability of Dairy Strains of Lactic Acid Bacteria to Bind Aflatoxin M₁ in a Food Model. *J. Food Prot.*, **63(5)**: 645-650.
 45. Rašić, J. L., Škrinjar, M. and Markov, S. 1991. Decrease of Aflatoxin B₁ in Yoghurt and Acidified Milks. *Mycopathologia*, **113(2)**: 117-119.
 46. Ruiz, L., Ruas-Madiedo, P., Gueimonde, M., De Los Reyes-Gavilan, C. G., Margolles, A. and Sanchez, B. 2011. How do Bifidobacteria Counteract Environmental Challenges? Mechanisms Involved and Physiological Consequences. *Gene. Nutr.*, **6(3)**: 307-318.
 47. Sarimehmetoğlu, B. and Küplülü, Ö. 2004. Binding Ability of Aflatoxin M₁ to Yoghurt Bacteria. *Ankara Üniv. Vet. Fak. Derg.*, **51(3)**: 195-198.
 48. Serrano-Niño, J. C., Cavazos-Garduño, A., Hernandez-Mendoza, A., Applegate, B., Ferruzzi, M. G., San Martin-González, M. F. and García, H. S. 2013. Assessment of Probiotic Strains Ability to Reduce the Bioaccessibility of Aflatoxin M₁ in Artificially Contaminated Milk Using an in Vitro Digestive Model. *Food Control.*, **31(1)**: 202-207.
 49. Settachaimongkon, S., Valenberg, van H. J. F., Winata, V., Wang, X., Nout, M. J. R., Hooijdonk, van A. C. M., Zwietering, M. H. and Smid, E. J. 2015. Effect of Sublethal Preculturing on the Survival of Probiotics and Metabolite Formation in Set-yoghurt. *Food Microbiol.*, **49**: 104-115.
 50. Shah, N. P. 2000. Probiotic Bacteria: Selective Enumeration and Survival in Dairy Foods. *J. Dairy Sci.*, **83(4)**: 894-907.
 51. Shahin, A. A. M. 2007. Removal of Aflatoxin B₁ from Contaminated Liquid Media by Dairy Lactic Acid Bacteria. *Int. J. Agri. Biol.*, **9(1)**: 71-75.
 52. Tajkarimi, M., Shojaee Aliabadi, F., Salah Nejad, M., Poursoltani, H., Motallebi, A. A. and Mahdavi, H. 2007. Seasonal Study of Aflatoxin M₁ Contamination in Milk in Five Regions in Iran. *Int. J. Food Microbiol.* **116(3)**: 346-349.
 53. Tamime, A. Y., Saarela, M., Sondergaard, A. K., Mistry, V. V. and Shah, N. P. 2005. Production and Maintenance of Viability of Probiotic Micro-organisms in Dairy Products. In: "Probiotic Dairy Products", (Ed.): Tamime, A. Y.. Blackwell Publishing, Oxford, PP 39-72.
 54. Zhao, R., Zhang, H., Niu, Sh. and Li, G. 2012. Tolerance of *Lactobacillus acidophilus* as Micro-ecological Strains by Simulating Gastrointestinal Environment. Future Communication, Computing, Control and Management. *The Series Lecture Notes in Electrical Engineering*. Springer, Berlin, Heidelberg, New York, **142**: 259-266.
 55. Zinedine, A., Farid, M. and Benlemlih, M. 2005. In Vitro Reduction of Aflatoxin B₁ by Strains of Lactic Acid Bacteria Isolated from Moroccan Sourdough Bread. *Int. J. Agri. Biol.* **7(1)**: 67-70.
 56. Zoghi, A., Khosravi-Darani, K. and Sohrabvandi, S. 2014. Surface Binding of Toxins and Heavy Metals by Probiotics. *Mini Rev. Med. Chem.*, **14(1)**: 84-98.



ارزیابی زمان انبارمانی و غلظت سم آفلاتوکسین M_1 بر ظرفیت اتصال به سم در باکتری لاکتوباسیلوس اسیدوفیلوس در محصول لبنی تخمیری

ن. ادیب پور، ص. سلیمانان زاد، م. سرابی جماب، و ف. تجلی

چکیده

آفلاتوکسین‌ها عوامل بالقوه سرطان‌زا و سرکوب‌کننده سیستم ایمنی بدن هستند. مواجهه شدید با سطح بالایی از آفلاتوکسین‌ها منجر به بروز آفلاتوکسیکوزیز شده، که مرگ سریع به دنبال نارسایی کبد را در پی خواهد داشت. اثرات تعدیل سیستم ایمنی مرتبط با باکتری‌های پروبیوتیک چشم‌انداز خوبی برای سم‌زدایی از مواد غذایی پیش روی ما قرار می‌دهد. هدف این مطالعه ارزیابی توانایی لاکتوباسیلوس اسیدوفیلوس سویه La-5 در حضور و عدم حضور آغازگرهای ماست جهت کاهش و یا حذف آفلاتوکسین M_1 در مقایسه با آغازگرهای ماست به عنوان تیمار شاهد بوده است (10^8 CFU/ml). سم‌زدایی AFM_1 در طول ۲۱ روز دوره انبارمانی ماست در دمای ۴ درجه سانتی‌گراد در غلظت‌های متفاوت آفلاتوکسین ($0/1$ ، $0/5$ ، $0/75 \mu\text{g/L}$) ارزیابی شد. مقدار آفلاتوکسین اتصال نیافته به سطح باکتری با استفاده از روش الیزای رقابتی اندازه‌گیری شد. با توجه به نتایج به دست آمده، لاکتوباسیلوس اسیدوفیلوس در ترکیب با آغازگرهای ماست و همچنین به تنهایی توانست به حد معنی‌داری ($P < 0/05$) در مقایسه با تیمار شاهد آفلاتوکسین را حذف کند. نتایج حاکی از آن بود که افزایش در مقدار اولیه غلظت آفلاتوکسین و مدت زمان انبارمانی بر ظرفیت اتصال به آفلاتوکسین موثر است.