Viability and Quality of Fermented Milk Made Using Local and Commercial Starters during Fermentation and Cold Storage

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ABSTRACT

Recent trends in food consumption and lifestyle show an increased demand for foods that are not only tasty and nutritious, but also provide additional benefits related to health, i.e. functional foods. One example of functional food is probiotic fermented milk produced with lactic acid bacteria. This has been shown to be beneficial to human gut health. The present study aimed to study the viability of Lactobacillus Casei subsp. casei R-68 (LCR-68) and Lactobacillus Casei strain Shirota (LCS) during fermentation and cold storage, as well as the quality of fermented milk produced from both strains. The research was conducted using a Completely Random Design. The data obtained was analyzed using ANOVA and DNMRT. The t-test was used to compare the growth and viability of LCR-68 and LCS. Fermentation time significantly affected the pH value, total lactic acid, total LAB and protein content, but did not significantly affect the fat and ash content of the fermented milk product. The best probiotic fermented milk in terms of viability and quality was produced via fermentation for 15 hours using strain LCR-68 as a starter. LCR-68 and LCS cultured in skimmed milk showed slightly different growth patterns. However, both strains showed similar viability. The total LAB after cold storage for a month was 6.64 and 6.68 log CFU mL−1 in the LCR-68 and LCS fermented milk, respectively. According to the results, LCR-68 can be used as a starter for making probiotic fermented milk.

Keywords: Lactobacillus casei strain Shirota, Lactobacillus casei subsp. casei R-68, Probiotic fermented milk.

INTRODUCTION

Probiotics are referred to as "live microorganisms that, when administered in adequate amounts, confer a health benefit to the host" (FAO/WHO, 2010). It is well-known that probiotics have many health benefits such as antimicrobial activity, diarrhea alleviation, anti-carcinogenic properties, improving lactose intolerance, strengthening the immune system (Cenci et al., 2002; Shah, 2007; FAO/WHO, 2010; Pato et al., 2017; Mahmoudi et al., 2019a), the ability to survive and adhere the gastrointestinal tract (Pato, 2003; Mahmoudi et al., 2019a; Mahmoudi et al., 2019b) and as a therapy against cold and flu pathogens (Leyer et al., 2009).

The demand for healthy foods has been one of the most important trends of food consumption in recent years (Bigliardi and Galati, 2013). Consumers are increasingly aware of their own health as well as the social and environmental impact of their food consumption (Falguera et al., 2012). Fermented milk is believed to have a good

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nutrition level, in addition to properties that are beneficial for human health, especially the digestive system, because it contains probiotics that can nourish the human digestive tract. The majority of commercial probiotics are *Lactobacillus* and *bifidobacteria* species used in products such as yogurt, fermented milk and frozen desserts (Tamin and Robinson, 2001; Shah, 2007). According to Maulidya (2007), fermented products that use skimmed milk as a starting ingredient are advantageous. This is because it is easier for digestion and contains a very low amount of fat, allowing the product to be stored longer compared to products made from whole milk.

One of the requirements for Lactic Acid Bacteria (LAB) being used as a probiotic is viability during the processing and storage stages. Storage time and temperature affect bacterial survival (Anggraini, 2016). No general agreement has been reached on the recommended intake level. The suggested levels range from $10^6$ CFU mL$^{-1}$ (Kurman and Rasic, 1991) to over $10^7$ and $10^8$ CFU mL$^{-1}$ (Korbekandi et al., 2011). However, it is generally recommended that the probiotic culture must be present in the product at a minimum number of $10^7$ CFU mL$^{-1}$ (Ishibashi and Shimamura, 1993). These suggestions have been made to compensate for the possible decline in the concentration of the probiotic organisms during the processing and storage of the product, as well as throughout the passage through the upper and lower parts of the gastrointestinal tract.

One example of a fermented milk product sold commercially is Yakult. Yakult uses *Lactobacillus casei* strain Shirota (LCS) as a starter. LCS has various beneficial health effects in humans (Spanhaak et al., 1998; Nagao et al., 2000; Matsumoto et al., 2006; Sutula et al., 2013). The shelf life of LCS is about 40 days with $10^6$ CFU mL$^{-1}$ total LAB during cold storage (Anonymous, 2015). Meanwhile, in West Sumatra, Indonesia, dadih is a traditional fermented food made from buffalo milk that is sold and consumed locally. Among the LAB isolates derived from dadih curd (Hosono et al., 1989) with the potential to be used as a probiotic, there is *Lactobacillus casei* subsp. *casei* R-68 (LCR-68) (Hosono et al., 1990; Pato, 2003, Pato and Hosono, 2004; Pato et al., 2017). Currently, there are no reports on the viability of LCR-68 during fermentation and cold storage. Therefore, the present study aimed to compare the viability of commercial strain LCS against local strain LCR-68 during cold storage, and the quality of the fermented milks produced from these strains.

**MATERIALS AND METHODS**

**Research Method**

The study was conducted experimentally. *Lactobacillus casei* subsp. *casei* R-68 (LCR-68) was isolated from dadih, traditional fermented buffalo milk, and identified by Hosono et al. (1989), *Lactobacillus Casei* strain Shirota (LCS) was obtained from the Indonesian Yakult Company. The viability of LCR-68 and LCS and the biochemical changes during fermentation were compared. Fermentation time varied between 6 to 26 hours at 37°C followed by cold storage at ±3-5°C. The cultures were observed every week starting weeks 0 to 6.

**Preparation of Active Culture**

Active culture was prepared according to Setioningsih et al. (2004), whereby 100 mL pure cultures of LCR-68 or LCS were inoculated into test tubes containing 5 mL of sterile MRS broth. The medium was stirred using an automatic mixer until homogeneous, then incubated at 37°C for 24 hours.

**Preparation of Starter**

The starter was prepared following methods described by Setioningsih et al.
(2004) using LCR-68 and LCS (Yakult). First, a 15% skimmed milk medium was prepared and stirred evenly using a mixer until homogeneous. Then, the medium was sterilized at 115 °C for 10 minutes. After the medium reached a temperature of ±40°C, the medium was inoculated with active 5% culture, then incubated at 37°C for 12 hours (starter I). Next, 5% of starter I was inoculated into 250 mL of 15% skimmed milk medium. This was incubated at 37°C for 12 hours to obtain starter II. Starter II was used to prepare the probiotic fermented milk.

**Preparation of Probiotic Fermented Milk**

Probiotic fermented milk was prepared according to Ginting (2016). First, the medium was prepared as follows: skimmed milk (157.5 g), CMC (0.525 g) and sucrose (52.5 g) were weighed accurately, and water was added to the mix to a volume of 1,050 mL. The mixture was stirred using a mixer. The homogeneous medium was distributed into 42 plastic bottles each of 25 mL, and pasteurized at 85°C for 15 minutes. The medium was then cooled to 40°C and inoculated with 5% LCR-68 starter II in 21 bottles, and LCS starter II in 21 bottles. The bottles were then incubated at 37°C for 6 to 22 hours to obtain the final probiotic fermented milk product. Fermentation times were optimized to produce the probiotic fermented milk for the study.

**Cold Storage Treatment of Probiotic Fermented Milk**

Cold storage of probiotic fermented milk was carried out according to methods described by Usman and Hosono (1999). Probiotic fermented milks made using LCR-68 or LCS were stored at ±3-5°C for six weeks, and observed every week starting from weeks 0 (first day) to 6.

**Parameters Observed**

The parameters measured in probiotic fermented milk were moisture, fat, protein, ash, carbohydrate, lactose, sucrose, and reducing sugar content as well as pH, total lactic acid, and LAB viability. Proximate analysis as well as lactose, sucrose, and reducing sugar contents were performed according to AOAC (2012). LAB viability was calculated according to the method described by Fardiaz (1998). pH was measured using a pH meter, and total lactic acid was determined by alkalimetric titration using 0.1N NaOH.

**Data Analysis**

The data were analyzed using analysis of variance (ANOVA). Data with F count equal to or greater than F table were further tested using the Duncan New Multiple Range Test (DNMRT) at 5% level to determine the differences between treatments. Data resulting from the comparison tests were further analyzed by t-test.

**RESULTS AND DISCUSSION**

**LCR-68 Fermented Milk Quality Parameters**

The results show that fermentation time significantly affected pH, total lactic acid and LAB viability in LCR-68 fermented milk (Figure 1).

Figure 1 shows that longer fermentation times decreased pH values significantly, and conversely increased total lactic acid. The longer the fermentation process, the more simple sugars (especially lactose and glucose) were broken down by LCR-68 into lactic acid. Similar results were reported in a traditional fermented milk product from Ghana called *Numu* (Akabanda et al., 2010) and in fermented sheep’s milk (Lasik et al., 2011).
Total lactic acid content of the probiotic fermented milks produced using different fermentation times in this study ranged from 0.57 to 0.97%. All the treatments produced fermented milks that met yogurt quality standards for total lactic acid content, which is between 0.5 to 2.0%; the minimum Codex standard for total lactic acid content is 0.3% and Yakult quality standards is 0.5%.

Longer fermentation times resulted in higher LAB viability. After fermentation for 15 to 18 hours, LAB viability increased significantly. After 21 hours of fermentation, LAB viability decreased significantly. Increased fermentation times led to an increased release of metabolic compounds into the fermented milk in the form of lactic acid. Increased lactic acid in the media inhibits the growth of LCR-68. The LAB viability of the probiotic fermented milk under treatments T4 and T5 (see Table 1) met the LAB viability of the yogurt quality standard equal to a minimum of $10^7$ CFU g$^{-1}$ (SNI 2981: 2009). It also met the Codex standards for fermented milk (Codex Stan 243-2003) and Yakult quality standards, while the commercial fermented milk products use LCS as a starter culture.

Analysis of the quality parameters of the fermented milk indicated that fermentation time significantly affected protein and carbohydrate levels, but did not significantly affect moisture, fat and ash contents of LCR-68 fermented milk (Table 1).

The data in Table 1 show that long fermentation times did not significantly affect moisture, fat and ash contents of fermented milk. Similar amounts of water, sucrose and skim milk were used as starting ingredients in the preparation of the fermented milk: 15% skimmed milk and 1% sucrose were used in the media. Across different fermentation times, it is possible that LCR-68 does not metabolize fat in the medium as an energy source during growth. Moreover, the moisture and ash components remained relatively constant in the media due to the constant water and mineral requirements by LAB during fermentation.

The data in Table 1 shows that the longer fermentation times increased the protein level in the fermented milk. Fermentation for 18 and 21 hours showed significantly increased protein levels. The increase in protein level can be attributed to the increase in the number of LCR-68, where protein
makes up a large part of the LAB cells. LAB cells containing protein were found in the cell wall, cell membrane, ribosome and cytoplasm (Moat et al., 2002; Hu et al., 2011). Thus, the higher the number of LABs, the higher level of protein obtained in the cocoghurt. Imam et al. (2015) reported an increase in protein levels in cocoghurt during fermentation from 3 to 15 hours. Conversely, longer fermentation times lowered the carbohydrate level of the fermented milk. The decrease in carbohydrate level is due to the metabolism of the lactose and sucrose sugars in the fermentation medium as a conversion into lactic acid by the LCR-68 strain. Strain LCR-68 mainly metabolizes lactose and sucrose to obtain energy and to form lactic acid as a by-product (Pato et al., 2017). This metabolic process is further indicated by a decrease in pH value and an increase in the total amount of lactic acid produced (Figure 1).

The moisture content of the fermented milks in the present study was slightly higher (85.14-85.51%) than that in commercial Yakult (81.9%). Fat content was also slightly higher (0.30-0.35%) than in Yakult (0.1%). Protein content in LCR-68 and LCS probiotic fermented milk (2.92-4.09%) was higher than that in Yakult (1.2%). Overall, protein levels in the final fermented milks met the standard protein content of at least 2.7% following the SNI and Codex standard. Meanwhile, ash content in the probiotic fermented milks (0.42-0.51%) was slightly higher than that in Yakult (0.3%). However, carbohydrate levels in Yakult were found to be higher than in our fermented milks following all treatment times. Yakult carbohydrate levels at 16.5% were much higher than those measured in our LCR-68 fermented milk (9.48-11.10%). The higher carbohydrate levels in Yakult could be attributed to the addition of sucrose syrup to the medium during the manufacturing process to produce a sweet-tasting product that is more preferred by consumers (Anonymous, 2015).

Based on the results of the first part of this study, it was found that a 15-hour fermentation time produced the best LCR-68 fermented milk in terms of quality and LAB viability. Therefore, part two of the study was carried out to compare the viability between strains LCR-68 and LCS over time. The results are shown in Figure 2. The longer the fermentation process, the more significant the decrease in pH value in both fermented milks (strain LCR-68 and strain LCS). Decreasing pH values in both types of fermented milk showed relatively similar patterns with no significant differences according to the t-test. The decrease in pH value in both probiotic fermented milks is a result of the metabolism of lactose and sucrose into lactic acid.

### Comparison of LCR-68 and LCS Fermented Milk

Longer fermentation times significantly increased total lactic acid levels produced in both LCR-68 and LCS probiotic fermented milk. The increase in total lactic acid

### Table 1. Quality parameters of probiotic fermented milk made using starter Lactobacillus casei subsp. casei R-68 (LCR-68) during the fermentation process.

<table>
<thead>
<tr>
<th>Fermentation time (h)</th>
<th>Moisture (%)</th>
<th>Fat (%)</th>
<th>Protein (%)</th>
<th>Ash (%)</th>
<th>Carbohydrate (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1 (Fermentation time for 6 h)</td>
<td>85.22 a</td>
<td>0.33 a</td>
<td>2.92 b</td>
<td>0.42 a</td>
<td>11.10 a</td>
</tr>
<tr>
<td>T2 (Fermentation time for 10 h)</td>
<td>85.14 a</td>
<td>0.35 a</td>
<td>3.21 ab</td>
<td>0.44 a</td>
<td>10.51 ab</td>
</tr>
<tr>
<td>T3 (Fermentation time for 14 h)</td>
<td>85.51 a</td>
<td>0.34 a</td>
<td>3.50 ab</td>
<td>0.46 a</td>
<td>10.25 abc</td>
</tr>
<tr>
<td>T4 (Fermentation time for 18 h)</td>
<td>85.05 a</td>
<td>0.34 a</td>
<td>3.79 ab</td>
<td>0.47 a</td>
<td>10.35 abc</td>
</tr>
<tr>
<td>T5 (Fermentation time for 22 h)</td>
<td>85.51 a</td>
<td>0.31 a</td>
<td>4.09 a</td>
<td>0.50 a</td>
<td>9.480 c</td>
</tr>
<tr>
<td>T6 (Fermentation time for 26 h)</td>
<td>85.36 a</td>
<td>0.30 a</td>
<td>4.09 a</td>
<td>0.51 a</td>
<td>9.750 bc</td>
</tr>
<tr>
<td>Commercial Yakult</td>
<td>81.90</td>
<td>0.10</td>
<td>1.20</td>
<td>0.30</td>
<td>16.50</td>
</tr>
</tbody>
</table>

* Means followed by the lowercase letters in the same column indicate significant difference (P< 0.05).
produced in both types of fermented milk showed relatively similar patterns. The amount of lactic acid produced by strain LCS was higher than that of strain LCR-68, but total lactic acid at similar fermentation times showed no significant difference, according to the t-test. Longer fermentation times increased the number of LAB significantly in both LCR-68 and LCS probiotic fermented milk. The increase was due to the availability of nutritional sources derived from skimmed milk and sucrose contained in the medium. Sucrose and lactose were used as energy sources for the relatively rapid growth of LAB at 10-hour fermentation time for strain LCS and 15 hours for LCR-68. In the production of kefir, during the first 24 hours of fermentation, the lactose content decreased from a mean value of 4.92 (w/w) to 4.02% (w/w); the concentration of L(+) lactic acid increased from 0.01% to 0.76% (w/w) and the pH decreased to 4.24 over the same period. After 24 hours of fermentation, the changes in the levels of lactose and L(+) lactic acid, and in pH, occurred more slowly (Fontan et al., 2006). Decrease in pH from 6.09 to 4.85 and lactose content from 45.80 to 32.40 mg mL⁻¹ occurred at the end of 6 to 24 hours fermentation time in Brazilian kefir (Leite et al., 2013).

LAB in both types of fermented milk increased, despite showing a somewhat different pattern (Figure 2). Both strains had the same growth pattern when fermented for five hours. After five hours, LCS continued growth in the logarithmic phase until 10 hours of fermentation before entering the stationary phase. On the other hand, LCR-68 growth remained in the logarithmic phase for up to 15 hours, before entering the stationary phase at 20 to 30 hours of fermentation. The t-test indicates significant differences between both strains after fermentation of 10 to 15 hours. At the end of single starter fermentation, LCR-68 and LCS were present at levels of 7.16 and 7.17 log units, respectively. In contrast, fermentation of kefir using mixed starters of lactic acid bacteria, acetic acid bacteria, and yeast presented lactic acid bacteria at 10 log units (Leite et al., 2013).

Viability of LCR-68 and LCS in Cold Storage

Yakult and probiotic fermented milk are generally distributed and sold in cold conditions. Therefore, it is necessary to study the viability of probiotics during cold storage.
Figure 3. LAB viability, pH and total lactic acid in probiotic fermented milk during cold storage.

Lactic acid bacteria viability in both strains LCR-68 and LCS decreased with length of cold storage. Total LCR-68 in fermented milk at weeks 0 to 6 decreased 20.21% from 7.42 to 5.92 log CFU mL$^{-1}$ (down 1.50 log CFU mL$^{-1}$), whereas total LCS from weeks 0 to 6 decreased 15.90% from 7.94 to 6.68 log CFU mL$^{-1}$ (down 1.26 log CFU mL$^{-1}$). Under the same conditions, LCR-68 showed greater decrease in LAB compared to LCS. A similar decrease in lactose levels in the fermented milk was observed; lactose levels in LCR-68 (42.27%) decreased more than LCS (35.0%) (Figure 3). According to Anggraini, 2016, the slow growth of LAB in fermented milk is caused by the limited amount of lactose in the fermented milk. As a result, activity of LAB was inhibited, resulting in decreased total LAB during cold storage. In contrast, the total lactic acid bacteria in Brazilian kefir remained constant during cold storage (Leita et al., 2013).

There are four phases of LAB growth: the lag phase (adaptation), the logarithmic (exponential) phase, the stationary phase, and the death phase (Volk and Wheeler, 1993). The data in Figure 3 shows a decrease in the number of both LAB strains from weeks 0 to 6. The total decrease in LAB is caused by a lack of additional nutrients for LAB growth during the cold storage process. The cells run out of energy, including reserved energy in their cells. After going through the logarithmic and stationary phases, the cells undergo the decline or death phase, which is characteristic of the low levels of LAB in the fermented milk after several weeks (Maulidya, 2007). Mulyani et al. (2008) reported that probiotic ice cream stored for 10, 20, and 30 days showed a decrease in the LAB viability from 1.5×10$^{10}$ to 1.3×10$^{9}$ CFU mL$^{-1}$, and then to 1.1×10$^{8}$ CFU mL$^{-1}$, respectively. Soyghurt probiotics used as praline chocolate fillers stored for four weeks showed a decrease in total LAB from weeks 0 to 4, i.e. 10.18, 10.15, 10.25, 10.34 and 9.97 CFU mL$^{-1}$, respectively (Rangkuti et al., 2013). The number of Lactobacilli and Streptococcus slightly decreased during cold storage at 5ºC for 15 days for both ewe and cow milk yoghurts (Karami, 2018).

The minimum number of probiotic strains in fermented milk products or food products is 10$^{8}$ CFU mL$^{-1}$. Meanwhile, the recommended daily intake for probiotics is around 10$^{8}$ CFU mL$^{-1}$ (Shah, 2007). Based on these results, LCR-68 fermented milk meets the criteria up to the fifth week; while LCS fermented milk meets the criteria up until the sixth week. At least 100 mL intake per day is required in
order for the fermented milk probiotics to benefit the human digestive tract.

Total lactic acid in the fermented milk during cold storage increased with length of storage time. Total lactic acid in LCR-68 fermented milk increased from weeks 0 to 6 from 0.34 to 0.56% (64.7% increase), while total lactic acid in LCS fermented milk from weeks 0 to 6, increased from 0.31 to 0.54% (74.2% increase) (Figure 3). The increase in total lactic acid in LCS fermented milk was greater than LCR-68. Lactic acid levels in fermented milks increase with increased storage time, as reported by Asaminew and Eyassu (2011). During cold storage, the fermentation process continues, whereby LAB metabolizes lactose and sucrose into lactic acid. An increase in total lactic acid correlates with an increase in total dissociated and non-dissociated acids (Usmiati et al., 2011). Lactic acid levels in yogurt that is accepted from the sensory standpoint ranges from 0.8 to 2.0%. The production of lactic acid in fermentation is closely related to the number of Lactobacillus casei in fermented products. In a study by Rangkuti et al. (2013), probiotic soyghurt used as praline chocolate filler had a shelf life storage period of four weeks. Total lactic acid in the soyghurt increased from week 0 by 0.32 to 0.55% at week 4. Another study by Ayuti et al. (2016) reports that Lactobacillus casei grown at 4-10°C over 90 days showed an increase in total lactic acid from 0.89% on the first day to 1.35% on Day 90.

pH during cold storage decreased with length of storage time (Figure 3). The pH of LCR-68 fermented milk from weeks 0 to 6 decreased from 4.70 to 3.47 (26.17% decrease), while pH in LCS decreased from 4.92 to 3.47 (35% decrease). The rate of decrease in LCR-68 fermented milk was greater than that of LCS fermented milk during cold storage. LCR-68 metabolizes lactose at a faster rate than LCS, as indicated by the lower amount of lactose remaining in LCS fermented milk at the end of storage time. Lactose is one of the simple sugars used by LAB for metabolic activity (Pramono et al., 2011). The more sugars metabolized, the more lactic acid produced, resulting in reduced pH of the fermented milk. Decrease in lactose levels during cold storage was also reported in soyoghurt (Muawanah, 2007) and in Brazilian kefir (Leita et al., 2013).

Sucrose levels during cold storage decreased with length of storage time (Figure 4). Sucrose levels in LCR-68 decreased from 4.70 to 0.67%, (85.75% decrease) from weeks 0 to 6. Sucrose levels in LCS increased from 4.92 to 5.32 (8.27% increase) from weeks 0 to 6. Sucrose levels decreased more rapidly in LCR-68 than in LCS during cold storage, which is consistent with the findings of Usmiati et al. (2011) where Dadih fermented for 21 days showed a decrease in pH from 4.51 to 3.95 and Leita et al. (2013) where Brazilian kefir fermented for 28 days in cold storage resulted in a decrease in pH from 4.75 to 4.32. Hence, storage time affects the pH in probiotic fermented milk. A decrease in pH in fermented milks coincides with lowered amounts of sucrose and lactose during cold storage (Figure 4).

Lactose and sucrose are used as an energy source by lactic acid bacteria; they are converted into lactic acid in the fermented milk (Figure 4). Lactose levels decreased during cold storage with increased length of storage time. Lactose levels in LCR-68 fermented milk decreased from weeks 0 to 6 from 5.03 to 4.92 (21.61% decrease), while lactose levels in LCS decreased from 4.70 to 4.17 (21.61% decrease). pH in both types of fermented milk decreased with increased storage time. LCR-68 fermented milk showed greater decrease in pH in cold storage than LCS. The fermentation process continues slowly in cold temperatures; sugars such as lactose and sucrose in fermented milk, which act as sources of energy, continue to be converted by LAB into organic acids, especially lactic acid. Lactic acid is dissociated into H+ and CH3CHOHCOO-. High levels of lactic acid allows the release of high levels of H+ ions into the medium, causing the pH of the fermented milk to drop (Khotimah and Kusnadi, 2014; Mirdalisa et al., 2016). The results of this study are in line with the findings of Usmiati et al. (2011) where Dadih fermented for 21 days showed a decrease in pH from 4.51 to 3.95 and Leita et al. (2013) where Brazilian kefir fermented for 28 days in cold storage resulted in a decrease in pH from 4.75 to 4.32. Hence, storage time affects the pH in probiotic fermented milk. A decrease in pH in fermented milks coincides with lowered amounts of sucrose and lactose during cold storage (Figure 4).
Lactic Acid Bacteria during Fermentation

Decrease over six weeks or 14.30% per week), while sucrose levels in LCS fermented milk from weeks 0 to 6 decreased from 4.80 to 0.84% (81.45% decrease over six weeks, or an average reduction of 14.57% per week). The rate of decrease in sucrose levels in fermented milk during cold storage was greater in LCR-68 than in LCS. According to Sobowale et al., 2011, sucrose is needed as an energy source to maintain cell survival in cold conditions. Therefore, longer storage periods led to higher levels of sucrose decline. Lactic acid bacteria use sugar as an energy source for growth, and produce metabolites in the form of lactic acid during fermentation (Salminen et al., 1998).

Similar decrease in sucrose levels from 1.56 to 1.28% was also reported in soy milk yoghurt by Muawanah (2007).

Reducing sugars are monosaccharides and disaccharides with reducing groups, such as glucose, fructose, galactose, maltose, and lactose. Reducing sugar levels in fermented milk during cold storage decreased with increase in length of storage time (Figure 4). Reducing sugar levels in LCR-68 fermented milk from weeks 0 to 6 decreased from 34.36 to 25.50% (25.78% decrease), while reducing sugar levels in LCS fermented milk from weeks 0 to 6 decreased from 34.51 to 25.32% (26.62% decrease). LCR-68 fermented milk showed lower decrease in reducing sugar levels compared to LCS fermented milk during cold storage. During fermentation, disaccharides are broken down into monosaccharides. Lactose in skimmed milk is broken down into glucose and galactose, and sucrose is converted into glucose and fructose. Reducing sugar levels correlate with total lactic acid (Figure 3). Higher levels of total lactic acid in fermented milk coincide with lower reducing sugar content. This is because, during fermentation, LAB uses simple sugars as an energy source for growth, and produces end metabolites in the form of lactic acid sugars to produce lactic acid (Salminen et al., 1998). In a study by Fithri et al. (2008) on fermented soy milk, reducing sugar levels decreased from 2.70 to 0.75%. The drop in reducing sugar levels was caused by utilization of the sugars by L. casei strains as an energy source for cell growth and the formation of metabolites such as lactic acid.

CONCLUSIONS

This study established that fermentation time significantly affected pH value, total lactic acid, total LAB, and protein content, but did not significantly influence fat and ash content in probiotic fermented milk produced with Lactobacillus Casei subsp. casei strain R-68 (LCR-68) and Lactobacillus Casei strain Shirota (LCS). High quality probiotic fermented milk was produced using strain LCR-68 as starter with 15-hours fermentation time. LCR-68 and LCS cultured in skimmed milk showed slightly different growth patterns, but similar viability in cold storage for six weeks. The quality parameters of LCR-68 fermented milk at weeks 0 and 6 contained total LAB of 7.42-5.92 log CFU mL⁻¹, total lactic acid 0.34-0.56%, pH 4.70-3.47, lactose 4.92-2.84%, sucrose 4.70-0.67%, and reducing sugar 34.36-25.50%. Whereas for LCS fermented milk, total LAB was 7.94-6.68 log CFU mL⁻¹, total lactic acid 0.31-0.54%, pH 5.32-4.17, lactose 5.03-3.27%, sucrose 4.80-0.84%, and reducing sugar 34.51-25.32%. In conclusion, LCR-68 can be used as a starter for making probiotic fermented milk.

ACKNOWLEDGEMENTS

We sincerely thank the Institute for Research and Community Service, Universitas Riau, and the Ministry for Research, Technology, and Higher Education of the Republic of Indonesia for providing the research grant for this study.

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ژیستایی و کیفیت شیر تخمری تولید شده با استفاده از مایه میکروبی محلی و تجاری در طی تخمر و در سرد خانه

چکیده

روند های اختیاری مصرف مواد غذایی و سیب زمینی افزایش تفاوت با برای غذاهای را نشان می‌دهد که فقط خوش مزه و مغذی ترین بسته به منافع دیگری نیز برای سلامت انسان دارند. بنابراین مواد غذایی زیست فعال (functional food) بکه نمونه از مواد غذایی زیست فعال، شیر تخمری پروپیوتیک تولید شده با بکتری‌های لاکتوژیک اسید است. چنین نشان داده شده که این مایه برای پزشکان معیث می‌باشد. Laetobacillus casei subsp. casei ر-68 (LCR-68) که بهبود ویژه کیفیت شیر تخمری تولید شده با در سویه. این پژوهش با استفاده از طرح کلی تصادفی انجام شد. داده‌های به دست آمده با استفاده از کار واژن داده شد. نیز از آزمون ١ برای مقایسه رشد و سرعت LCS-68 و LCR-68 ANOVA و DNMRT تحلیل استفاده شد. نتایج نشان داد که طول زمان تخمری pH که لاجیس اسید کلی و محتوای پروتئین به طور معناداری تاثیر داشت ولی اثر بر چربی و کاهش شیر تخمری تولید شده معنی دار نبود. از نظر زیستایی و کیفیت، بهترین شیر تخمری پروپیوتیک با ١٥ ساعت تخمری و استفاده از سویه Skimmed میکروئی تولید شد. اکتا های رشد LCS-68 و LCR-68 که در سویه Skimmed LCS-68 و LCR-68 که در سویه Skimmed LCS-68 و LCR-68

References:
Lactic Acid Bacteria during Fermentation

Lactic Acid Bacteria (LAB) are used in the production of milk. In one study, the LAB population was determined after fermentation. The LAB population was counted using the CFU/mL method. The LAB population was found to be 46/4 log CFU mL⁻¹. The LAB population in the control sample (LCR-68) was 68/6, while the population in the experimental sample (LCR-68) was 46/4. The LAB population in the control sample was significantly different from the experimental sample.