Expression Analysis of Gene Encoding 1-Deoxy-d-Xylulose 5-Phosphate Reductoisomerase (DXR) and Cardenolide and Digitoxin Production in Digitalis purpurea L. Using Polyamines and Methyl Jasmonate as Elicitors

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ABSTRACT

Digitoxin is a glycoside isolated from leaves of various species of Digitalis genus that are commonly called foxgloves. Digitoxin is used for chronic cardiac insufficiency, tachyarrhythmia form of atrial fibrillation, paroxysmal ciliary arrhythmia, and paroxysmal supraventricular tachycardia. 1-Deoxy-d-Xylulose 5-phosphate Reductoisomerase (DXR) is an important rate-limiting enzyme in 2-C-Methyl-d-Erythritol 4-Phosphate (MEP) pathway, and expression level of DXR plays a critical role in control of plant isoprenoid biosynthesis pathway. The effects of different elicitors including putrescine, spermine, and spermidine at concentrations of 50 and 100 mg L⁻¹ and also Methyl Jasmonate (MJ) at 50, 100, and 200 μM concentrations were explored on transcript levels of DXR gene in cell suspension culture of foxglove (Digitalis purpurea). DXR transcription levels were assessed by the semi-quantitative RT-PCR. Results showed that elicitors had significant effects on the expression level of DXR and contents of cardenolide and digitoxin. The highest digitoxin (61.3 μg g⁻¹ DW) and cardenolide (1.48% mg⁻¹ DW) content was observed in 100 mg L⁻¹ spermidine treatment. In all treatments, the expression of DXR gene was increased compared with the control sample. The highest transcription levels of DXR gene was observed in 50 μM MJ and 100 mg L⁻¹ putrescine elicitors, which was nine folds higher than the control condition, and the lowest levels were observed for putrescine treatments at concentrations of 50 and 100 mg L⁻¹, respectively.

Keywords: Elicitor, Foxglove, Gene expression, Tissue culture.

INTRODUCTION

Medicinal plants are the source of secondary metabolites and bioactive compounds that are used as food additives, natural pigments, insecticides, flavors, aromas, fragrances, bio-based fuels, plastics, enzymes, preservatives, cosmetics (cosmeceuticals), bioactive compounds, and fine chemicals (Mulabagal and Tsay, 2004). Digitalis purpurea (foxglove, common foxglove, purple foxglove) is a species of medicinal flowering plant in the family Plantaginaceae (Olmstead et al., 2001). The plants are well known as the original source of the heart medicine digoxin (Olmstead et al., 2001). Plant tissue culture is the

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technique of maintaining and growing plants on artificial medium under defined conditions in vitro (Bunsupa et al., 2018). Cell cultures have been established for many plants, but often they do not produce sufficient amounts of the required secondary metabolites. However, in many cases the biosynthesis of secondary metabolites can be enhanced by treatment of undifferentiated cells with elicitors such as methyl jasmonate (Poulev et al., 2003). An ‘elicitor’ may be defined as a substance which, when introduced in small concentrations to a living cell system, initiates or improves the biosynthesis of specific compounds (Gorelick and Bernstein, 2014). It is well known that the treatment of plants with elicitors causes accumulation of a range of plant defensive secondary metabolites in intact plants or in plant cell cultures (Poulev et al., 2003). There are many reports on Methyl Jasmonate (MJ) effect on secondary metabolite production such as Wang et al. (2015) who enhanced production of secondary metabolites by MJ elicitation in cell suspension culture of Hypericum perforatum. Khan et al. (2019) reported that elicitors had significant effects on the production of pharmacologically attractive secondary metabolites in callus cultures of Fagonia indica. It was reported that MJ had effects on the production of secondary metabolites in plant cell suspension cultures of Thevetia peruviana (Mendoza et al., 2018).

Terpenoids constitute one of the most diverse groups of natural products and have different roles in growth regulation, cell signaling, inducing defense mechanisms, membrane structures, and redox chemistry (Rodriguez-Concepcion et al., 2001). All isoprenoids are derived from a common building unit, Isopentenyl diPhosphate (IPP) and its isomer Dimethylallyl diPhosphate (DMAPP), and plastidial isoprenoids are derived from a mevalonate-independent pathway currently known as the 2-C-Methyl-d-Erythritol 4-Phosphate (MEP) pathway (Rodriguez-Concepcion et al., 2001). The initial reaction is formation of 1-Deoxy-d-Xylulose 5-Phosphate (DXP) from pyruvate and d-glyceraldehyde 3-phosphate, and it is catalyzed by DXP synthase (DXS) (Lange et al., 1998). 1-Deoxy-d-Xylulose-5-phosphate isomerase-Reductase gene (DXR), catalyzes the NADP-dependent rearrangement and reduction of 1-Deoxy-d-Xylulose-5-Phosphate (DXP) to 2-C-Methyl-d-Erythritol 4-Phosphate (MEP), which is the key element for MEP-pathway that supports the main C5 units for the formation of mono and diterpenes (Takahashi et al., 1998; Banerjee et al., 2013). Plastidial MEP pathway is considered as the main important source of precursors for essential plastid isoprenoids (Wright et al., 2014). Most isoprenoids including volatile compounds are involved in plant resistance responses to biotic and abiotic stresses and made from precursors produced by the MEP pathway (Gershenzon and Dudareva, 2007). Because of the pivotal role of DXR for providing the downstream products of the MEP pathways, in the past two decades, there has been increasing interest in exploring the DXR function for manipulating biosynthetic pathways to increase the yield of terpenoids for different purposes. By altering the expression of DXR, the beneficial metabolite compounds have increased in mint plant (Mahmoud and Croteau, 2002). Constitutive expression of DXR in Arabidopsis seedlings has resulted in the accumulation of high levels of isoprenoids such as chlorophylls, tocopherols, and carotenoids (Carretero-Paulet et al., 2006). Over-expression of DXR gene also resulted in high increase of diterpene yield in transgenic bacteria (Morrone et al., 2010). Biotic elicitors have significant role in production of pharmaceutical terpenoids by increasing the amount of the enzymes involved in biosynthetic pathways via regulating the expression of biosynthetic genes (Kang et al. 2009; Lu et al., 2016). Xing et al. (2010) reported a high expression of DXR under the drought as an abiotic stress. Application of MJ increased the expression level of DXR and increased triptophanolide content in cell
culture suspension of *Tripterygium wilfordii* (Hook. f.) (Tong *et al*., 2015). Because of the key role of DXR as an upstream gene involved in MEP pathways for producing secondary metabolites in plants, we evaluated the expression of the DXR in *Digitalis purpurea* L. as an important medicinal plant in responses to polyamines and MJ (biotic elicitors) treatments under *in vitro* condition.

**MATERIALS AND METHODS**

**Sterilization and Culture Medium**

The seeds of the *Digitalis purpurea* L. with code number 411Dp*MPISB were provided from Medicinal Plants Institute, Academic Center for Education, Culture and Research (ACECR). The seeds were disinfected under laminar flow hood with 70% ethanol for one minute, then sterilized with 1% sodium hypochlorite for 10 minutes and washed three times with autoclaved distilled water. Also, all instruments were sterilized by autoclave at 121°C and 1.5 atmospheres for 20 minutes. The macro and micro elements and vitamins were added to the medium according to Murashig and Skoog’s consideration.

The most effective treatments for callus induction was established based on our previously study (BAP 2 mg L⁻¹; 2,4-D 0.5 mg L⁻¹). The levels of elicitors included four levels of MJ (distilled water as control, 50, 100, and 200 μM, abbreviated as MJ0, MJ50, MJ100, MJ200) and 7 levels of polyamine (50 and 100 mg L⁻¹ of putrescine; 50 and 100 mg L⁻¹ of spermine; 50 and 100 mg L⁻¹ of spermidine; Treatments abbreviation: Ctrl- Control, Put50- 50 mg L⁻¹ Putrescine, Put100- 100 mg L⁻¹ Putrescine, SPM50- 50 mg L⁻¹ Spermine, SPM100- 100 mg L⁻¹ Spermine, SPD50- 50 mg L⁻¹ Spermidine, SPD100- 100 mg L⁻¹ spermidine). The research was performed based on factorial experiment in RCBD design with three replications.

**Cardenolide and Digitoxin**

Cardenolide and digitoxin concentration in treated plants were measured by HPLC as described by Desta and McErlane (1982).

**Measuring Cardenolide**

Methanol (MeOH) with 1.8 mL value was used to extract 50 mg of callus and the samples were homogenized at a speed of 6 m.s⁻¹ for 45 seconds using the Fast Prep Tool (MP Biomedicals, Solon, Ohio, USA). After centrifugation, the supernatant solution was collected and filtered through a pressure of 0.45 mµ. Fifteen μL of the extract was injected into an HPLC of the Agilent 1100 series and the compounds were discarded on the reverse phase column of Gemini C18. Extraction of cardenolides was performed with a constant flow of 0.7 mg min⁻¹ with phosphoric acid 0.25% of acetonitrile on a gradient of water: 0 to 5 minutes 20% of acetonitrile, 20 minutes 70% of acetonitrile, 20-25 minutes 70% of acetonitrile, 30 minutes 95% of acetonitrile, 30-35 minutes 95% of acetonitrile. The UV absorption spectrum from 200 to 400 nm was recorded by a diode array detector. Courtesy with a maximum absorption of 217 and 222 nm was recorded as Cardenolide (Bingham and Agrawal, 2010).

**Measuring Digitoxin**

Calli were dried in an oven at 40°C for 48 hours, then, 50 mg of dried calluses was treated with 1 milliliter of 70% methanol for 30 minutes in ultrasonic bath at 65-70-65°C and immediately placed on ice for 3 minutes. It was centrifuged at room temperature for 10 minutes at 13,000 rpm. Then, the supernatant was centrifuged with 0.25% of 15 m/v lead acetate solution, lead Acetate Isolated was discarded separately. Half a milliliter of 4 m / v monosodium phosphate was added and centrifuged.
Supernatant was mixed with 0.5 milliliters of chloroform/isopropanol solution in a ratio of 3 to 2 and centrifugation was performed for 5 minutes. The lower phase was transferred to the 2 mL tube as the first extract. The remaining methanolic solution was used for secondary extraction. Chloroform/isopropanol was added and then centrifuged at room temperature for 5 minutes at 13,000 rpm. Both extracts were combined under the laminar hood and injected to HPLC apparatus (Desta and McErlane, 1981).

**Semi-Quantitative RT-PCR**

Semi quantitative RT-PCR reaction was used to determine the relative expression of DXR in treated samples with different polyamine stimuli. Primer sequences for DXR gene and 18srRNA as reference gene are shown in Table 1.

Total RNA was extracted from 200 mg of upper leaves of treated and un-treated Digitalis lanata L. according to the manufacturer’s protocol (Sinaclon, Iran). DNase treatment was performed using the RNase-Free DNase Set (Promega, Promega UK, Southampton, UK) at the end of the extraction procedure. cDNA synthesis was performed using M-MLV Reverse Transcriptase (Fermentas, Germany) following the manufacturer’s protocol. The yield of cDNA was measured based on the PCR signal generated using 18SrRNA as a reference gene amplified from 30 to 35 cycles starting with 2 µL of the cDNA template. Semi-quantitative RT-PCR was carried out using published protocol (Spencer and Christensen, 1999) to measure the expression level of DXR in treated cells.

To ensure that no false-positive PCR fragments would be generated from pseudogenes in contaminating genomic DNA, primer sequences were designed to span intron regions whenever genomic sequence data were available (Table 1). All RT and PCR were performed with reagents from Fermentas; Inc. PCR in 30 and 35 different cycles was performed independently as follows: 1 cycle of 95 ºC for 3 minutes; 94 ºC for 1 minute, 55 ºC for 40 seconds and 72ºC for 60 seconds; 1 cycle of 72 ºC for 7 minutes. Each set of PCR reaction included cDNA from non-treated plant as template for negative control and cDNA template from the digitoxin treated sample as a positive control. PCR products were resolved in 1.5% agarose gel and stained in GelRed™ (Biotium). A 100 bp DNA ladder molecular weight marker (Fermentas, Germany) was used to confirm the expected molecular length of the amplified products.

Expression level of the DXR gene was assessed according to the published protocol (Antiabong et al., 2016). The amplicon images (PCR bands) in the gel were captured under Ultraviolet (UV) light and documented using a DigiDoc gel documentation system (Bio-rad; Australia). All the parameters and experimental conditions used were kept constant throughout the study. Image analysis was done using ImageJ software version 1.4.3u (Abramoff et al., 2004). The PCR band density was automatically generated by the ImageJ software using the specified “area”.

**Statistical Design and Analysis**

Factorial (methyl jasmonate and

### Table 1. Primer sequences for DXR and 18srRNA genes.

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequences</th>
<th>Amplicon size (bp)</th>
<th>Accession No.</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>DXR F</td>
<td>5'-GTGCCGGGACTAAAGCCTACG-3'</td>
<td>101</td>
<td>NM_001203671.1</td>
<td>Mohammadi Faresani (2015)</td>
</tr>
<tr>
<td>DXR R</td>
<td>5'-CGGAAGCAGAAAGGACCAC-3'</td>
<td>461</td>
<td>JQ612131.1</td>
<td>Nematollahi et al. (2012)</td>
</tr>
<tr>
<td>18srRNA F</td>
<td>5'-AACGGCTACCACATCCAAG-3'</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>18srRNA R</td>
<td>5'-TCATTACTCCGATCCCAG-3'</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Table 2. Means of square for digitoxin and cardenolide in response to experimental treatments.

<table>
<thead>
<tr>
<th>SOV</th>
<th>Digitoxin</th>
<th>Cardenolide</th>
<th>Dry weight</th>
<th>Fresh weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Polyamine</td>
<td>658.51**</td>
<td>0.49**</td>
<td>2199.37</td>
<td>548498.96</td>
</tr>
<tr>
<td>methyl jasmonate</td>
<td>727.48**</td>
<td>0.58**</td>
<td>3975.09**</td>
<td>1077929**</td>
</tr>
<tr>
<td>Polyamine×Methyl jasmonate</td>
<td>181.25**</td>
<td>0.18**</td>
<td>1462.21**</td>
<td>431637.24**</td>
</tr>
<tr>
<td>Error</td>
<td>2.79</td>
<td>0.001</td>
<td>24.55</td>
<td>8643.61</td>
</tr>
<tr>
<td>CV (%)</td>
<td>4.16</td>
<td>4.75</td>
<td>3.78</td>
<td>6.21</td>
</tr>
</tbody>
</table>

** significant effect at 1% statistically level.

polyamine) experiments were conducted based on completely randomized design with three replications. The SAS (9.1 v) was applied for data analysis, also means comparisons were done with Duncan test at 5% probability level. Image j software was used for analysis of molecular section in relation to electrophoresis product.

RESULTS AND DISCUSSION

Application of Elicitors

It was found that interaction effect of polyamine and methyl jasmonate had significant effect on fresh, dry, and fresh/dry weights of callus (Table 2). The highest means of fresh (2204 mg), dry (168.3 mg), and fresh/dry (13.10) weights was obtained by the application of 100 mg L\\(^{-1}\\) Spermidine+50 μM MJ treatments. In addition, lowest means of fresh (877.3 mg) and dry (86.27 mg) weights was obtained by the application of 200 μM MJ without polyamine. In this regard, El-Mekkawy et al. (2018) reported that application of elicitors led to induction of *Ecballium Elaterium* callus and resulted in increasing yields of the secondary metabolites.

Digitoxin and Cardenolide

According to analysis of variances, treatments had significant effects on Digitoxin and Cardenolide traits: the highest means of cardenolide and digitoxin was observed by 100 mg L\\(^{-1}\\) spermidine+50 μM MJ and this treatment showed 4.1 and 2.4 folds increase of cardenolide and digitoxin compared to the control, respectively. In addition, lowest means of cardenolide and digitoxin were obtained by 200 μM MJ without polyamine (Figure 1). Treatment of plant cells with biological elicitors (glycemic polymers, glycoproteins, small organic acids, or extracts from fungal cells) and non-toxic elicitors (US, UV, salts, heavy metals and substances such as salicylic acid, Methyl jasmonate, fungal eucalyptus, silver nitrate, arachidonic acid and vanadyl sulfate) is always one of the most effective tools that increases the synthesis of secondary metabolites. The elicitors, through induction of defensive responses, cause biosynthesis and accumulation of secondary metabolites (Perez-Alonso et al., 2016).

Gene Expression of DXR in Response to Different Elicitors

Grecian foxglove plants treated with any kind of polyamine compound expressed the DXR gene transcript levels more than the untreated control for 3 to 7 days after treatment (Figures 2 and 3). However, DXR gene transcribed in cells treated with 100 mg L\\(^{-1}\\) putrescine and methyl jasmonate 50 μM + putrescine 100 mg L\\(^{-1}\\) transcribed to lower and highest levels compared to the negative control. Samples treated with the 50 μM methyl jasmonate+100 mg L\\(^{-1}\\) putrescine or 50 μM methyl jasmonate+50 mg L\\(^{-1}\\)
Figure 1. Mean comparisons for interaction of polyamines and methyl jasmonate on digitoxin (a) and cardenolide (b).

Spermine mg accumulated DXR gene transcripts about 7.5 and 7 fold more than the negative control, respectively. DXR smilingly transcribed to a 2-fold level compared to the negative control in plants treated with the 50 μM methyl jasmonate+50 and 100 mg L⁻¹ of spermidine as well as 100 μM methyl jasmonate+100 mg L⁻¹ putrescine, polyamine compounds. Results showed that with increase in the concentration of polyamines in methyl jasmonate 200μM+putrescine 50 mg treatment, DXR transcript level did not significantly increase and it was about the level of the later treatments. Dabiri et al. (2017) showed that DXS was up-regulated in response to methyl jasmonate elicitor. Fan et al. (2016) reported that DXS and DXR were differentially expressed in roots, stems, leaves, and protocorms, with higher transcript levels in roots and stems compared with other organs in Dendrobium.
Figure 2. Semi-quantitative RT-PCR of RDX gene in digitoxin treated and un-treated grecian foxglove plants, 3 to 7 days after treatment. Tissue of grecian foxglove plants treated with digitoxin is the positive control and different concentrations of polyamines compounds. The levels of DXR messenger RNA accumulation are compared with that of the housekeeping gene 18S rRNA. Numbers indicate the different concentrations of treated polyamines as follows: putrescine (50 and 100 mg), spermidine (50 and 100 mg), methyl jasmonate 50 mg+ putrescine 100 mg, methyl jasmonate 50 mg+Spermin 50 mg, methyl jasmonate 50 mg+spermidin (50 and 100 mg), methyl jasmonate 100 mg+putrescine 100 mg, and methyl jasmonate 200 mg+putrescine 500 mg, respectively. C, positive control; N, negative control; L, 100 bp DNA ladder. Amplified products for DXR and 18S rRNA genes are 101 and 461 bp in size for 30 cycles of PCR, respectively.

Figure 3. Assessment by RT-PCR of DXR gene transcript levels in tissue of grecian foxglove plants treated with digitoxin as positive control and different concentrations of polyamines including: putrescine (50 and 10 mg), Spermidin (50 and 100 mg), methyl jasmonate 50 mg+ putrescine 100mg, Methyl Jasmonate 50 mg+Spermin 50 mg, Methyl Jasmonate 50 mg+Spermidin (50 and 100 mg), methyl jasmonate 100 mg+putrescine 100 mg, and methyl jasmonate 200mg+putrescine 500 mg. Densitometry analysis results are shown as bar charts whose values appear relative to the mean intensity of the positive controls (arbitrary value 1). Bars: Standard error.

Officinale. As a result, transcript levels of DXS and DXR were induced by abscisic acid, methyl jasmonate, and salicylic acid, whereas their transcript levels were not regulated by brassinolide. According to the fact that DXR plays a pivotal role in the control of plant isoprenoid biosynthesis, application of different elicitors may affect...
the plant isoprenoid quality and quantity. It has been reported that there are correlations between the expression of this gene and isoprenoid biosynthesis. Of course, this correlation depends on the type of plant, tissue, and stage of development (Schmiderer et al., 2010). Mahmoud and Croteau (2002) reported that in transgenic peppermint, the increasing of DXR expression led to the increase in the monotropenes compared to wild plants, and depletion caused a significant decrease in monotropenes. Such a correlation was reported by McConkey et al. (2000) on peppermint.

CONCLUSIONS

Results of the present study suggested that foxglove and its callus were a potential source of digitoxin and cardenolide content. Elicitors, namely, putrescine, spermine, spermidine, and methyl jasmonate significantly influenced the digitoxin and cardenolide. This research provides valuable insights into the potential manipulation of foxglove callus on the production of secondary metabolite for pharmacological, cosmetic, and agronomic industries. Therefore, the elicitation of cultured tissues is necessary to improve the production of phytochemical compounds. Likewise, DXR interestingly transcribed to a 7.5 to 8 fold levels compared to the negative control in plants treated with the methyl jasmonate 50 mg+ spermine 50 mg as well as methyl jasmonate 50 mg+ putrescine 100 mg, polyamine compounds.

REFERENCES

Xylulose-5-Phosphate Reductoisomerase in *Dendrobium officinale*. Plant Cell Tissue Organ Culture (PCTOC), 125: 381-385.


مورد بررسی قرار گرفت. تغییرات در سطح رونویسی زن DXR توسط روش RT-PCR تأیید گردید. نتایج نشان داد که محورهای زیستی اثر معنی‌دار بر مقدار بیان Zn DXR و بیوتنز کاردنولید و دیجیتکسین دارند. بالاترین میزان دیجیتکسین (1/3 میکرو‌گرم بر گرم وزن خشک) و کاردنولید (1/68 درصد در میلی‌گرم وزن خشک) در تیمار 100 میلی‌گرم در لیتر اسپره‌ایدیان مشاهده شدند. در کلیه تیمارها بیان زن DXR در مقایسه با نمونه‌های کنترل افزایش یافت. بالاترین سطح رونویسی زن DXR در غلظت 50 میکرومولار میل جامشونات و 100 میلی‌گرم در لیتر پوترسین و وجود داشت که بهترین بیشتر از شرایط کنترل بود و همچنین کمترین میزان در تیمار پوترسین در غلظت های 50 و 100 میلی‌گرم در لیتر مشاهده شد.