

## Does *Wolbachia* Change Diapause and Energy Reserves of *Trichogramma brassicae* in Response to Light Wavelengths?

S. Rahimi-Kaldehy<sup>1</sup>, A. Bandani<sup>1</sup>, and A. Ashouri<sup>1\*</sup>

### ABSTRACT

The present study examined the light wavelengths effect on the diapause percentage of progeny and energy reserves of maternal generation in sexual and asexual *Trichogramma brassicae* that had been reared under different light wavelengths before oviposition. Photoperiod has a maternal effect on the diapause induction in *Trichogramma* wasps; however, the light wavelengths effect on their diapause has not been studied. In this study, we reared the maternal generation of both strains under five light wavelengths including blue (455–475 nm), green (515–535 nm), orange (585–595 nm), red (620–630 nm), and white (5,000–10,000 K), and allowed 24 hours old females to oviposit in *Ephestia kuehniella* eggs. The diapausing generation was placed at 10°C and absolute darkness for two months. The results showed that *Wolbachia* infection and light wavelengths had significant effects on the diapause percentage and energy reserves of *T. brassicae*, excepting glycogen contents. The maximum and minimum diapause percentage was observed under green and white light in asexual, and under white and green light in sexual strain. The data showed that the sexual strain had lower lipid and protein levels than the asexual strain, except when exposed under white light. The diapause percentage in the sexual strain was higher than in the asexual strain under all light wavelengths, and the reaction of parasitoids toward light wavelengths was different in the two strains. Therefore, *Wolbachia* can cause a different reaction to light wavelengths in both diapause percentage and pattern of the parasitoid. These results should be considered to improve mass-rearing and long-term storage of this parasitoid.

**Keywords:** Egg parasitoid, Mass-rearing, Maternal effect.

### INTRODUCTION

Diapause is a critical component of unfavorable conditions for most insects in temperate regions (Denlinger, 1991). It represents a complex process characterized by physiological and behavioral features (Kostal, 2006). It is known that most of the *Trichogramma* species show facultative diapause during the prepupal stage when they encounter low temperatures in nature or laboratory conditions.

Low temperatures are one of the most important factors that induce diapause in *Trichogramma* species (Zaslavski and

Umarova, 1981; Keller, 1986; Sorokina and Malesnnikova, 1987; Garcia *et al.*, 2002). While the maternal influence of photoperiod on the diapause induction of many *Trichogramma* wasps has been studied (Zaslavski and Umarova, 1990; Garcia *et al.*, 2002; Ma and Chen, 2006; Ivanov and Reznik, 2008; Pizzol and Pintureau, 2008; Reznik *et al.*, 2011; Voinovich *et al.*, 2013), no studies have investigated the effect of light wavelengths on the diapause induction of *Trichogramma* wasps.

On the one hand, *Trichogramma* is positive light-dependent wasps (Costas, 1951) and, on the other hand, light wavelengths have an influence on insect

<sup>1</sup> Department Plant Protection, College of Agriculture and Natural Resources, University of Tehran, Karaj, Islamic Republic of Iran.

\*Corresponding author; e-mail: ashouri@ut.ac.ir



behavior (Ripfel and Becker, 1982). Experimental works have shown that color perception exists in insects and they are sensitive to light color ranging from ultraviolet to red, being especially responsive to the shorter wavelengths of the visible spectrum and UV light (Hamdorf *et al.*, 1971). When insects are exposed to light, they may show positive or negative phototaxis to the source of light, they may increase or decrease the rate of their general activity in response to the light wavelengths (Bertholf, 1940). In the case of diapause, insects showed a different response to the light wavelengths (Williams *et al.*, 1965; Harris *et al.*, 1967; Beck, 1980; Ismail *et al.*, 2011) thus we assumed that the percentage of diapause in *Trichogramma* may also be altered by light wavelengths.

*T. brassicae* is known as the most abundant species that parasitize a wide range of economically important agricultural and forest Lepidopteran pests in Iran (Poorjavad, 2011). Similar to most *Trichogramma* species, *T. brassicae* has two reproductive modes: arrhenotoky, which produces sexual wasps, and thelytoky, which produces asexual wasps. Most of the thelytokous *Trichogramma* wasps are infected by an endosymbiont bacterium, which is called *Wolbachia* (Stouthamer and Werren, 1993).

The aim of this study was to check two hypotheses. The first one was that white lights might increase the percentage of diapause in *T. brassicae*, which is followed by more energy reserves in females reared under this light. The second hypothesis was that there should be a difference in response of sexual and asexual strains to various light wavelengths, which is followed by different energy reserves in those females. Since there was no information regarding energy reserves of the maternal generation of *Trichogramma* wasps who were ovipositing progenies for entering diapause, this research was carried out to compare energy reserves of newly emerged females reared on different light wavelengths.

## MATERIALS AND METHODS

### Insects

Both sexual and asexual *T. brassicae* strains used in this study were identified by Dr. N. Pourjavad (Isfahan University of Technology, Isfahan, Iran). Both of these were collected from the North of Iran (south of the Caspian Sea) from eggs of *Ostrinia nubilalis* (Hubner) (Lep: Crambidae), and obtained from a culture maintained at the Ecology and Behavior Laboratory, University of Tehran. They were reared on eggs of *E. kuehniella* Zeller (Lep: Pyralidae) under constant Laboratory conditions ( $20\pm 1^\circ\text{C}$ , 16L:8D photoperiod and  $70\pm 5\%$  RH) for many generations (more than 100 generations).

### Experimental Design for Diapause Induction

Five cardboard paper strips ( $8\times 1$  cm) with about 200-300 *E. kuehniella* eggs, which were less than 24 hours old, were used for each strain and light wavelength. The eggs of Mediterranean flour moth were subjected for 4 hours to parasitization by 100 asexual and 200 sexual *T. brassicae* in plastic cylinders (approximately 18 cm tall $\times$ 8 cm in diameter) with an opening that was covered with a mesh in order to allow for ventilation. Then, these cards with parasitized host eggs were individually placed in glass tubes and incubated at  $20\pm 1^\circ\text{C}$ ,  $70\pm 5\%$  RH and 10L:14D photoperiod under five different light wavelengths, where white light (5,000~10,000 K) was the control and the other four treatments were as follows: blue (455~475 nm), green (515~535 nm), orange (585~595 nm), and red (620~630 nm). Introduction of the best light wavelengths for each strain could improve diapause percentage of *T. brassicae*. The glass tubes were positioned in boxes with white background that had light barriers on both sides. All white boxes were placed in a bigger black box. At the day of mass emergence, 40 cardboard paper strips ( $5\times 1$

cm) with about 50 host eggs per card were offered to emerged females (24 hours old) of each strain and exposed to light wavelength for 2 hours in plastic cylinders, individually. In all treatments, oviposited females were removed after 2 hours, and the parasitized eggs were stored at  $70\pm 5\%$  RH, in absolute darkness, and  $10\pm 1^\circ\text{C}$ . Parasitized eggs were transferred to control temperature ( $20^\circ\text{C}$ ) after two months to speed up the development and to facilitate the emergence of living individuals. In the end, the emergence rate of both *T. brassicae* strains was measured by dividing the number of parasitized eggs with an emergence hole with the total number of parasitized eggs. We did not include cardboard paper strips with less than 10 parasitized *E. kuehniella* eggs in our experiments.

#### Sample Preparation to Determine Energy Reserves

About 100 newly emerged females ( $0.001\pm 0.0001$  g) were used to quantify the content of proteins, glycogen, carbohydrate and total lipids of maternal *T. brassicae* generation, which were reared at  $20\pm 1^\circ\text{C}$ ,  $70\pm 5\%$  RH and 10L:14D under different light wavelengths, with three replications. Protein, glycogen, carbohydrate and lipid levels were determined using common biochemical analysis protocols (Foray *et al.*, 2012). First, *T. brassicae* were weighed and homogenized in 90  $\mu\text{L}$  of aqueous lysis buffer solution [100 mm  $\text{KH}_2\text{PO}_4$ , 1 mm DiThioThreitol (DTT) and 1 mm EthyleneDiamineTetraacetic Acid (EDTA), pH 7.4]. Then, the samples were centrifuged at 180 g for 10 min at  $4^\circ\text{C}$ .

#### Protein Determination

The protein concentration was determined as described by Bradford (1976). At the beginning, 10  $\mu\text{L}$  of supernatant was mixed with 500  $\mu\text{L}$  Coomassie Brilliant Blue G-250. Then, protein concentration was measured at 595 nm after incubation at room

temperature for 15–20 minutes, using a dilution-series of Bovine Serum Albumin (BSA) as a standard. Each measurement was performed in triplicates.

#### Glycogen Determination

To determine the glycogen, carbohydrate, and lipid contents, 10  $\mu\text{L}$  of lysis buffer solutions, 10  $\mu\text{L}$  of 2%  $\text{Na}_2\text{SO}_4$  and 750  $\mu\text{L}$  of chloroform: methanol (1:2) were added to the first supernatant. Then, the samples were centrifuged for 15 minutes at  $180\times g$  and  $4^\circ\text{C}$  to remove glycogen from the supernatant, which was transferred into a new tube to determine the carbohydrate and lipid contents. The pellet was used to determine the glycogen content. After vigorous vortexing and centrifuging for 5 min at  $16,000\times g$  and  $4^\circ\text{C}$ , 500  $\mu\text{L}$  of anthrone reagent was added to the pellet, followed by 15 min of incubation at  $90^\circ\text{C}$  in a water bath. The glycogen content was then determined by measuring the absorbance of the sample at 625 nm with glycogen as the standard. Each measurement was performed in triplicates.

#### Carbohydrate Determination

To determine the carbohydrate contents, 150  $\mu\text{L}$  of supernatant was transferred to a microtube and was evaporated for approximately 30 minutes at room temperature. Then, it was mixed with 240  $\mu\text{L}$  of anthrone reagent and was heated for 15 minutes at  $90^\circ\text{C}$  in a water bath. The carbohydrate contents were then determined by measuring the absorbance of the sample at 625 nm with glucose as the standard (van Handel, 1965; 1985a). Each measurement was performed in triplicates.

#### Lipid Determination

The cholesterol was used as the standard to determine the total amount of lipids of each

treatment according to the vanillin assay procedure (van Handel, 1985b). For the assay, 100 µL of the supernatant was transferred into a borosilicate microplate well and heated at 90°C until complete solvent evaporation. Then, 10 µL of 98% sulfuric acid was added and the microplate was incubated at 90°C for 2 minutes in a water bath. After cooling the microplate on ice, 190 µL of vanillin reagent was added. The plate was homogenized, incubated at room temperature for 15 minutes, and its absorbance was measured spectrophotometrically at 525 nm. Each measurement was performed in triplicates.

### Statistical Analysis

The data was transformed by Johnson transformation  $[(-2.06231+1.33263) \times \ln(X+39.2659)/(100.991-X)]$  and the effects of strain (*Wolbachia* infection) and light wavelengths on the percentage of diapause of *T. brassicae* and energy compartments were compared using two-way ANOVA. Data was subjected to two-way Analyses Of Variance (ANOVA), followed by Tukey test to separate means when differences were significant at  $P < 0.05$ . All analyses were

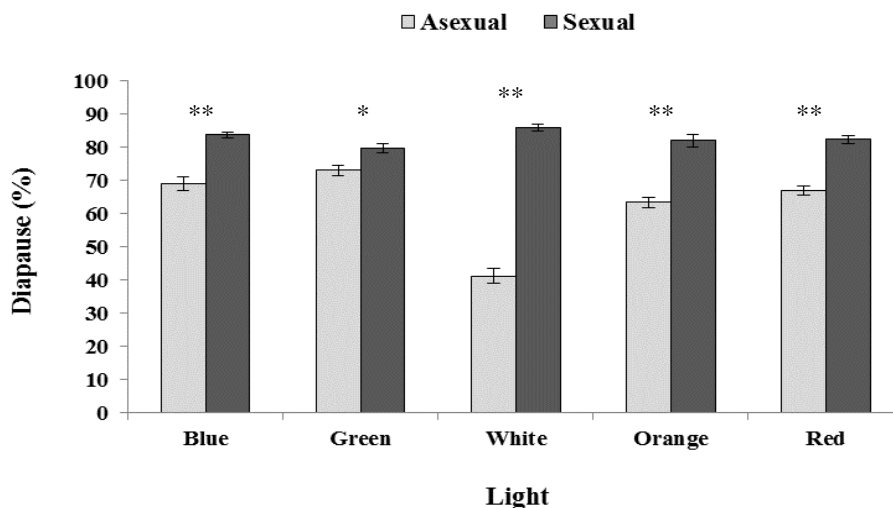
performed with SAS 9.1.3. Version.

## RESULTS AND DISCUSSION

The results revealed that strain ( $F = 339.42$ ;  $df = 1, 390$ ;  $P < 0.0001$ ), light wavelengths ( $F = 7.20$ ;  $df = 4, 390$ ;  $P < 0.0001$ ) and interaction between strain and light wavelengths ( $F = 24.06$ ;  $df = 4, 390$ ;  $P < 0.0001$ ) had statistically significant influence on the diapause percentage of *T. brassicae*.

White light caused the highest percentage of diapause ( $85.88 \pm 1.07\%$ ), followed by blue ( $83.74 \pm 0.95\%$ ), red ( $82.35 \pm 1.13\%$ ), orange ( $82.00 \pm 1.79\%$ ), and green ( $79.66 \pm 1.46\%$ ) lights in the sexual strain, whereas the highest percentage of diapause occurred under green light ( $73.02 \pm 1.46\%$ ), followed by blue ( $69.01 \pm 2.16\%$ ), red ( $66.99 \pm 1.44\%$ ), orange ( $63.44 \pm 1.51\%$ ), and white ( $41.19 \pm 2.16\%$ ) lights in the asexual strain (Figure 1). According to the results, sexual and asexual *T. brassicae* strains responded differently when they were exposed to different light wavelengths.

Thus, we concluded that diapause was greatly influenced by light wavelengths in both strains, although more diapause occurred in sexual *T. brassicae* compared to



**Figure 1.** Effects of strain (*Wolbachia* infection) and different light wavelengths on the diapause percentage of *T. brassicae*. The different light wavelengths include blue (455~475 nm), green (515~535 nm), cool white (5,000~10,000 K), orange (585~595 nm) and red (620~630 nm). There is a statistically significant difference ( $P < 0.0001$ ) according to the two-way ANOVA.

the asexual strain. Similar results were reported in *Nasonia vitripennis* (Walker) (Hym: Pteromalidae) (Bordenstein and Werren, 2000) and *T. oleae* Voegelé and Pointel (Pintureau *et al.*, 2002).

Similar to the sexual strain observed in this research, Ismail *et al.* (2011) showed that the highest diapause percentage occurred under white colors, which was followed by green in *Tetranychus urticae* Koch (Acari: Tetranychidae), but none of them entered diapause under red color. Bunning and Joerrens (1960) indicated that blue light induced diapause in *Pieris brassicae* L. (Lep: Pieridae) during early photophase, while diapause occurred later under red light. Beck (1980) indicated that the most effective wavelengths for diapause induction are between 400-550 nm for most species. According to his experiment, red light works very much like absolute darkness in diapause induction, whereas when the pupae of *Antheraea pernyi* Guer (Lep: Attacidae) were exposed to wavelengths shorter than 560 nm, it caused complete breaking of diapause (Williams *et al.*, 1965). Harris *et al.* (1967) indicated the same results: *Anthonomus grandis* Boh (Col: Curculionidae) could not enter diapause when it was exposed to wavelengths between 485 and 560 nm.

### Lipid, Protein, Carbohydrate, and Glycogen Concentration

Most energy reserves in the maternal generation of *T. brassicae* were in the form of lipids (Figure 2). The light wavelengths, strain and their interaction had a significant effect on the lipid concentration of *T. brassicae* (Table 1). Lipids are usually the main energy reserve for overwintering of insects (Yaginuma and Yamashita, 1978; Beenackers *et al.*, 1981; Storey and Storey, 1983). It seems that accumulation of lipids in insects is to support their energy demands during harsh conditions (Bashan *et al.* 2002). Amiri and Bandani (2013) reported that lipids were accumulated by prediapause

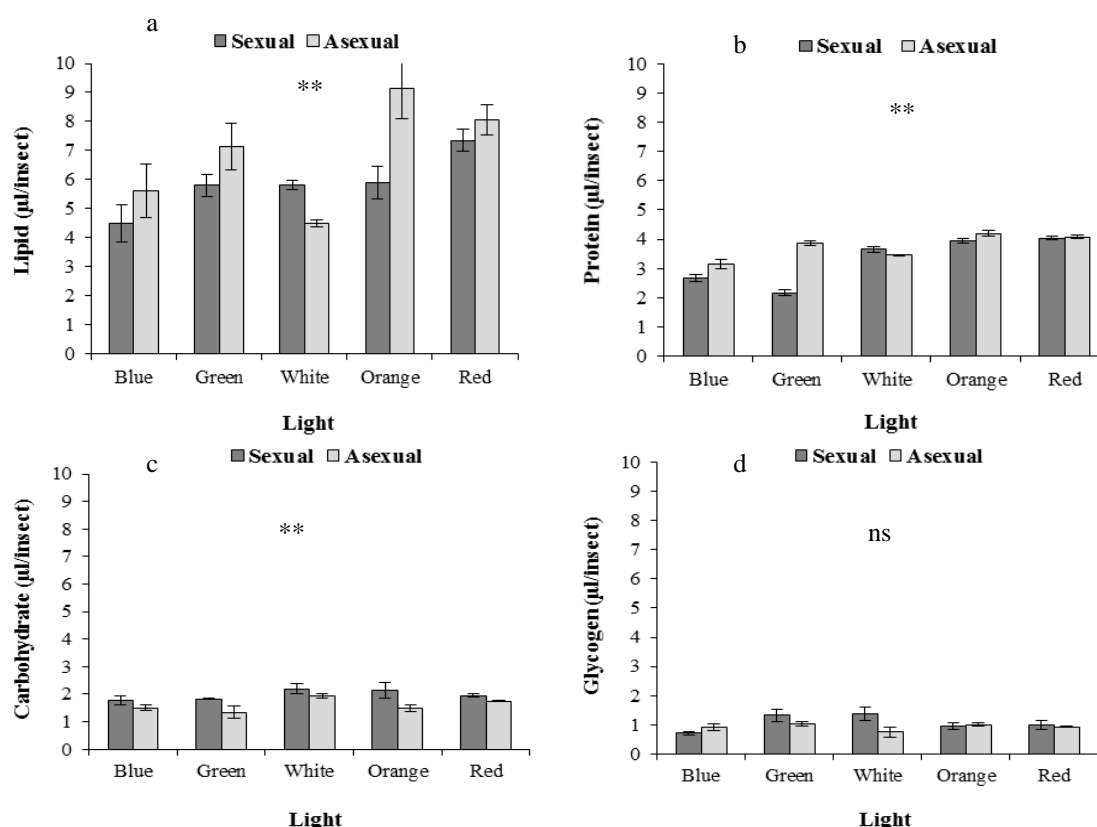
Sunn pest bugs for their metabolic needs during diapause. Data showed that the sexual strain had lower lipids than the asexual strain, with the exception of white light (Figure 2-a).

Similarly, the light wavelengths, strain, and their interaction had a significant effect on the protein concentration of *T. brassicae* (Table 1). According to the results, asexual *T. brassicae* had higher protein levels than sexual *T. brassicae*, with the exception of white light (Figure 2-b).

Sexual *T. brassicae* had higher carbohydrate than asexual *T. brassicae* in all the light wavelengths (Figure 2-c). Light wavelengths and strain had significant effects on the carbohydrate concentration (Table 1), whereas the interaction of light wavelengths and strain did not have a significant effect on the carbohydrate concentration of the parasitoid.

Glycogen content was the lowest energy of reserves in *T. brassicae*. According to the data analysis, sexual *T. brassicae* had higher glycogen than asexual *T. brassicae*, with the exception of blue and orange lights (Figure 2-d). Light wavelengths, strain, and their interaction did not show significant effect on the glycogen concentration of *T. brassicae* (Table 1), which indicates that this parasitoid may not use it as an energy reserve for diapause. However, previous studies have recognized the influence of accumulation of low molecular weight sugars and polyols at the beginning of diapause in many species of insects (Han *et al.*, 2005; Khani *et al.*, 2007; Han *et al.*, 2008).

The results demonstrated that carbohydrates and glycogen may convert to lipids in the maternal generation of both sexual and asexual *T. brassicae*. Numerous studies have stated that carbohydrates were converted into lipids within the insects' fat body (Hines and Smith, 1963; Bailey, 1975; Venkatesh and Morrison, 1980; Inagaki and Yamashita, 1986; Briegel, 1990). Zhou *et al.* (2004) have reported that the fat body of *Aedes aegypti* (L.) (Dip: Culicidae) females have a high capacity for the synthesis of



**Figure 2.** Effects of strain (*Wolbachia* infection) and different light wavelengths on the contents of (a) Lipid, (b) Protein, (c) Carbohydrate and (d) Glycogen ( $\mu\text{g insect}^{-1}$ ) of *T. brassicae* females. The light wavelengths are defined under Figure 1 and text. There is a statistically significant difference ( $P < 0.0001$ ) according to the two-way ANOVA.

**Table 1.** Effects of the strain (*Wolbachia* infection), light wavelengths and the interaction of strain and light wavelengths on lipid, protein, carbohydrate and glycogen contents ( $\mu\text{g insect}^{-1}$ ) of *T. brassicae*.

Energy reserves	Source of variation	df	Mean square	F
Lipid	Strain	1	7.70	6.73*
	Wavelength	4	9.56	8.36**
	Strain×Wavelength	4	3.93	3.44*
	Error	20	1.14	
Protein	Strain	1	1.48	50.65**
	Wavelength	4	1.82	62.31**
	Strain×Wavelength	4	0.81	27.55**
	Error	20	0.03	
Carbohydrate	Strain	1	1.06	16.46**
	Wavelength	4	0.21	3.33*
	Strain×Wavelength	4	0.06	0.96 <sup>ns</sup>
	Error	20	0.06	
Glycogen	Strain	1	0.17	3.01 <sup>ns</sup>
	Wavelength	4	0.11	1.99 <sup>ns</sup>
	Strain×Wavelength	4	0.16	2.74 <sup>ns</sup>
	Error	20	0.06	

\*, \*\*: Significantly difference at 5 and 1% probability level, respectively.

lipids than glycogen, and this may explain the higher content of lipids compared to glycogen in the maternal generation of both sexual and asexual *T. brassicae*. As glycogen is rapidly catabolized (Rambabu and Rao, 1994; Sancho *et al.*, 1998), the reduction in glycogen contents may be a result of energy generation for flight. It seems that the high amount of proteins in comparison with carbohydrate and glycogen is due to using protein as an energy source, which has been reported in some insects such as *Glossina* spp. (Dip: Glossinidae) and *Leptinotarsa decemlineata* (Col: Chrysomelidae) (Niaqi *et al.*, 1992).

In conclusion, our results documented some behavioral and biochemical adaptations for maternal generation of sexual and asexual *T. brassicae*. A large amount of lipids and proteins accumulated prior to oviposition with overwintering abilities. For the first time a study showed how the form of energy reserves changes under the influence of different light wavelengths in the maternal generation of *T. brassicae*, which were ovipositing progenies for entering diapause. According to the results, lipids were the most abundant (about 50%) energy reserves in all the treatments. It seems that mothers accumulate lipids to protect their progenies from cold and adverse effects.

Light wavelengths had a significant effect on the diapause percentage of the parasitoids that were affected by *Wolbachia* infection. Furthermore, there was a relation between the diapause percentage of progenies of females developed under different light wavelengths and the amount of energy reserves of ovipositing females. On the one hand, the lowest lipid content under white light in comparison with other lights was followed by a reduction in the diapause percentage in the asexual strain. On the other hand, the lowest protein content under green light in sexual females was followed by the lowest diapause percentage in these wasps in comparison with other wavelengths. The maternal generation accumulated more lipid as they developed

under the short photoperiod and 20°C, which induced diapause in their progenies (the same condition as that of late summer or onset of autumn). We assume that they probably transfer the lipids to the eggs of their diapausing generation. Finally, we can introduce the white light as a suitable wavelength for rearing sexual *T. brassicae*, and green light for asexual *T. brassicae*.

### ACKNOWLEDGEMENTS

The authors appreciate Dr. Nafiseh Poorjavad (Isfahan University of Technology, Isfahan, Iran) for molecular and morphological identification of asexual and sexual *T. brassicae* strains, and Petra Pribicevic (Faculty of Philology, University of Belgrade, Serbia) for her proofreading.

### REFERENCES

1. Amiri, A. and Bandani, A. R. 2013. Comparison of Energy Reserves in Prediapause and Diapausing Adult Sunn Pest, *Eurygaster integriceps* Puton (Hemiptera: cutelleridae). *J. Agr. Sci. Tech.*, **15**: 435-444.
2. Bailey, E. 1975. Biochemistry of Insect Flight. Part 2. Fuel Supply. In: "*Insect Biochemistry and Function*", (Eds.): Candy, D. J. and Kilby, B. A. Chapman and Hall, London, UK, PP. 89-176.
3. Bashan, M., Akbas, H. and Yurdakoc, K. 2002. Phospholipid and Triacylglycerol Fatty Acid Composition of Major Life Stages of Sunn Pest, *Eurygaster integriceps* (Heteroptera: Scutelleridae). *Comp. Biochem. Physiol.*, **132**: 375-380.
4. Beck, S. D. 1980. *Insect Photoperiodism*. 2<sup>nd</sup> Edition, Academic Press, New York, London, 387 PP
5. Beenackers, A. M. T., Van der Horst, D. J. and Van Marrewijk, W. J. A. 1981. Role of Lipids in Energy Metabolism. In: "*Energy Metabolism in Insects*", (Ed.): Downer, R. J. H. Plenum Press, New York, USA.
6. Bertholf, L. M. 1940. Reactions to Light in Insects. *Bios.*, **11**: 39-43.



7. Bordenstein, S. R. and Werren, J. H. 2000. Do *Wolbachia* Influence Fecundity in *Nasonia vitripennis*? *Heredity*, **84**: 54-62.
8. Bradford, M. M. 1976. A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Proteins Utilizing the Principle of Protein Dye Binding. *Anal. Biochem.*, **72**: 248-254.
9. Briegel, H. 1990. Metabolic Relationship between Female Body Size, Reserves, and Fecundity of *Aedes aegypti*. *J. Insect Physiol.*, **36**: 165-172.
10. Bunning, E. and Joerrens, G. 1960. Tagesperiodische Antagonistische Schwankungen der Blau-Violett- und Gelbrot-Empfindlichkeit als Grundlage der Photoperiodischen Diapause-Induktion bei *Pieris brassicae*. *Zeitschrift für Naturforschung.*, **15**: 205-213.
11. Costas, L. A. 1951. The Effect of Varying Conditions on Oviposition by *Trichogramma* on Eggs of Angoumois Grain Moths. *J. Econ. Entomol.*, **34**: 57-58.
12. Denlinger, D. L. 1991. Relationship between Cold Hardiness and Diapause. In: "Insect at Low Temperature", (Eds.): Lee, R. E. and Denlinger D. L. Chapman and Hall, New York, USA, PP. 174-198.
13. Foray, V., Pelisson, P. F., Bel-Venner, M. C., Desouhant, E., Venner, S., Menu, F., Giron, D. and Rey, B. 2012. A Handbook for Uncovering the Complete Energetic Budget in Insects: The van Handel's Method (1985) Revisited. *Physiol. Entomol.*, **37**: 295-302.
14. Garcia, P. V., Wajnberg, E., Pizzol, J. and Olivejra, M. L. M. 2002. Diapause in the Egg Parasitoid *Trichogramma cordubensis*: Role of Temperature. *J. Insect Physiol.*, **48**: 349-355.
15. Hamdorf, K., Schwemer, J. and Gogala, M. 1971. Insect Visual Pigment Sensitive to Ultraviolet Light. *Nature.*, **231**: 458-459.
16. Han, R. D., Gan, Y. L., Kong, X. H. and Ge, F. 2008. Physiological and Endocrine Differences between Diapausing and Non-Diapausing Larvae of the Pine Caterpillar *Dengrolimus tabulaeformis* (Lepidoptera: Lasiocampidae). *Zool. Stud.*, **47**(1): 96-102.
17. Han, R. D., Ge, F., Yardim, N. E. and He, Z. H. 2005. The Effect of Low Temperatures on Diapause and Non-Diapause Larvae of Pine Caterpillar *Dengrolimus tabulaeformis* (Lepidoptera: Lasiocampidae). *Appl. Entomol. Zool.*, **40**: 429-435.
18. Harris, F. A., Lloyd, E. P., Lane, H. C. and Burt, E. C. 1967. Influence of Light on Diapause in the Boll Weevil I. Dependence of Diapause Response on the Spectral Composition of the Light Used to Extend the Photoperiod. *J. Econ. Entomol.*, **60**: 1565-1567.
19. Hines, W. J. W. and Smith, M. J. H. 1963. Some Aspects of the Intermediary Metabolism in the Desert Locust, *Schistocerca gregaria* Forskal. *J. Insect Physiol.*, **9**: 463-468.
20. Inagaki, S. and Yamashita, O. 1986. Metabolic Shift from Lipogenesis to Glycogenesis in the Last Instar Larval Fat Body of the Silkworm, *Bombyx mori*. *Insect Biochem.*, **16**: 327-331.
21. Ismail, M. S. M., AboGhalia, A. H., Soliman, M. F. M. and Ghallab, M. M. A. 2011. Certain Effects of Different Spectral Colors on Some Biological Parameters of the Two-Spotted Spider Mite, *Tetranychus urticae*. *Egypt. Acad. J. Biol. Sci.*, **3**(1): 27-39.
22. Ivanov, M. F. and Reznik, S. Ya. 2008. Photoperiodic Regulation of the Diapause of the Progeny in *Trichogramma embryophagum* Htg. (Hymenoptera, Trichogrammatidae): Dynamics of Sensitivity to Photoperiod at Immature Stages of Maternal Females. *Annu. Rev. Entomol.*, **88**(3): 261-268.
23. Keller, M. A. 1986. Overwintering by *Trichogramma exiguum* in North Carolina. *Environ. Entomol.*, **15**: 659-661.
24. Khani, A., Moharamipour, S. and Barzegar, M. 2007. Cold Tolerance and Trehalose Accumulation in Overwintering Larvae of the Codling Moth, *Cydia pomonella* (Lepidoptera: Tortricidae). *Eur. J. Entomol.*, **104**: 385-392.
25. Kostal, V. 2006. Eco-Physiological Phases of Insect Diapause. *J. Insect Physiol.*, **52**: 113-127.
26. Ma, C. S. and Chen, Y. W. 2006. Effects of Constant Temperature, Exposure Period, and Age on Diapause Induction in *Trichogramma dendrolimi*. *Biol. Control.*, **36**: 267-273.
27. Niaqi, E. N., Olembo, N. K. and Pearson, D. J. 1992. Proline Transport by Tsetse Fly *Glossina morsitans* Flight Muscle Mitochondria. *Comp. Biochem. Physiol. B Comp. Biochem.*, **102**(3): 579-584.



28. Pintureau, B., Lassabliere, F., Daumal, J. and Grenier, S. 2002. Does a Cyclic Natural Thermal Cure Occur in *Wolbachia*-Infected *Trichogramma* Species? *Ecol. Entomol.*, **27**: 366-372.
29. Pizzol, J. and Pintureau, B. 2008. Effect of Photoperiod Experienced by Parents on Diapause Induction in *Trichogramma cacoeciae*. *Entomol. Exp. Appl.*, **127**: 72-77.
30. Poorjavand, N. 2011. Morphological, Molecular and Reproductive Compatibility Studies on the Systematic of the Genus *Trichogramma* Westwood (Hymenoptera: Trichogrammatidae) in Tehran and Mazandran Provinces (Iran). PhD. Dissertation, University of Tehran, Iran.
31. Rambabu, J. P. and Rao, M. B. 1994. Effect of Organochlorine and Three Organophosphate Pesticides on Glucose, Glycogen, Lipid and Protein Contents in Tissues of the Freshwater Snail *Bellamya dissimilis* (Muller). *Bull. Environ. Contam. Toxicol.*, **53**: 142-148.
32. Reznik, S. Ya., Voinovich, N. D. and Vaghina, N. P. 2011. Maternal Influence on Diapause Induction in *Trichogramma* (Hymenoptera, Trichogrammatidae): the Dynamics of Pphotosensitivity. *J. Appl. Entomol.*, **135**(6): 438-445.
33. Ripfel, J. and Becker, J. 1982. Light-Dependent Mating of *Drosophila subobscura* and Species Discrimination. *Behav. Genet.*, **12**(3): 241-260.
34. Sancho, E., Ferrando, M. D., Fernandez, C. and Andreu, E. 1998. Liver Energy Metabolism of *Anguilla anguilla* after Exposure to Fenitrothion. *Ecotoxicol. Environ. Saf.*, **41**: 168-175.
35. Sorokina, A. P. and Maslennikova, V. A. 1987. Temperature Optimum for Diapause Induction in Species of the Genus *Trichogramma* Westw. (Hymenoptera, Trichogrammatidae). *Entomol. Obozr.*, **66**(4): 689-699.
36. Stouthamer, R. and Werren, J. R. 1993. Microbes Associated with Parthenogenesis in Wasps of the Genus *Trichogramma*. *J. Invertebr. Pathol.*, **61**: 6-9.
37. Storey, J. M. and Storey, K. B. 1983. Regulation of Cryoprotectant Metabolism in the Overwintering Gall Fly Larva, *Eurosta solidaginis*: Temperature Control of Glycerol and Sorbitol Levels. *J. Comp. Physiol.*, **149**: 495-502.
38. van Handel, E. 1965. Microseparation of Glycogen, Sugars, and Lipids. *Anal. Biochem.*, **11**: 266-271.
39. van Handel, E. 1985a. Rapid Determination of Glycogen and Sugars in Mosquitoes. *J. Am. Mosq. Control Assoc.*, **1**: 299-301.
40. van Handel, E. 1985b. Rapid Determination of Total Lipids in Mosquitoes. *J. Am. Mosq. Control Assoc.*, **1**: 302-304.
41. Venkatesh, K. and Morrison, P. E. 1980. Studies of Weight Changes and Amount of Food Ingested by the Stable Fly, *Stomoxys calcitrans* (Diptera: Muscidae). *Can. Entomol.*, **112**: 141-49.
42. Voinovich, N. D., Vaghina, N. P. and Reznik, S. Y. 2013. Comparative Analysis of Maternal and Grand-Maternal Photoperiodic Responses of *Trichogramma* Species (Hymenoptera: Trichogrammatidae). *Eur. J. Entomol.*, **110**(3): 451-460.
43. Williams, C. M., Adkisson, P. L. and Walcotts, C. 1965. Physiology of Insect Diapause. XV. The Transmission of Photoperiod Signals to the Brain of the Oak Silkworm, *Antheraea pernyi*. *Biol. Bull.*, **128**(3): 497-507.
44. Yaginuma, T. and Yamashita, O. 1978. Polyol Metabolism Related to Diapause in *bombyx* Eggs: Different Behavior of Sorbitol from Glycerol during Diapause and Post Diapause. *J. Insect Physiol.*, **24**: 347-354.
45. Zaslavski, V. A. and Umarova, T. Ya. 1981. Photoperiodic and Temperature Control of Diapause in *Trichogramma evanescens* Westw. (Hymenoptera, Trichogrammatidae). *Entomol. Obozr.*, **60**(4): 721-731.
46. Zaslavski, V. A. and Umarova, T. Ya. 1990. Environmental and Endogenous Control of Diapause in *Trichogramma* Species. *Entomophaga.*, **35**: 23-29.
47. Zhou, G., Pennington, J. E. and Wells, M. A. 2004. Utilization of Pre-Existing Energy Stores of Female *Aedes aegypti* Mosquitoes during the First Gonotrophic Cycle. *Insect Biochem. Mol. Biol.*, **34**: 919-925.



## آیا آلودگی به ولباکیا، درصد دیابوز و میزان ذخایر انرژی زنبور پارازیتوئید *Trichogramma brassicae* را در پاسخ به طیف نوری تغییر می دهد؟

س. رحیمی کلدی، ع. بندانی، و ا. عاشوری

### چکیده

در این پژوهش، اثر قرارگیری در طیف های مختلف نوری بر تغییرات ذخایر انرژی پارازیتوئیدهای ماده آلوده به ولباکیا و غیرآلوده *Trichogramma brassicae* و نیز درصد دیابوز نتاج آن ها مورد مطالعه قرار گرفت. طول دوره نوری دارای اثر مادری در القای دیابوز در زنبورهای پارازیتوئید جنس *Trichogramma* است اما تاکنون اثر طیف های نوری بر دیابوز آن ها مطالعه نشده است. نسل مادری تحت طیف های نوری: آبی (۴۷۵-۴۵۵ نانومتر)، سبز (۵۳۵-۵۱۵ نانومتر)، نارنجی (۵۹۵-۵۸۵ نانومتر)، قرمز (۶۳۰-۶۲۰ نانومتر) و سفید (۵۰۰ تا ۱۰۰۰۰ کلوین) پرورش یافت و به ماده های ۲۴ ساعته اجازه داده شد تا در تخم های *Ephestia kuehniella* تخم گذاری کنند. نسل دیابوز گذران در تاریکی مطلق و دمای ۱۰ درجه سلسیوس به مدت دو ماه نگهداری شد. بر اساس نتایج به دست آمده، آلودگی به ولباکیا و قرارگیری نسل مادری تحت طیف های مختلف نوری، بطور معنی داری در القای دیابوز و نیز میزان ذخایر انرژی *T. brassicae* به استثنا میزان گلیکوژن اثر گذار است. حداکثر و حداقل درصد دیابوز در جمعیت آلوده در طیف های سبز و سفید و در جمعیت غیرآلوده در طیف های سفید و سبز مشاهده شد. ماده های غیرآلوده در تمامی طیف های نوری به استثنا طیف نوری سفید، لیپید و پروتئین کمتری نسبت به ماده های آلوده داشتند. درصد دیابوز در جمعیت غیرآلوده بیش از آلوده در تمامی طیف های نوری بود. دو جمعیت واکنش کاملاً متفاوتی را در برابر طیف های نوری نشان دادند. ولباکیا سبب بروز پاسخ متفاوت دو جمعیت از لحاظ درصد و الگوی دیابوز شد. نتایج حاصل از این پژوهش می تواند نقش کلیدی در بهبود روند تولید انبوه و ذخیره بلندمدت این عامل کنترل بیولوژیک داشته باشد.