

Effect of CO₂ Enrichment on Gas Exchanges, Biochemical Traits, and Minituber Yield in Potato (*Solanum tuberosum* L.) Cultivars

M. J. Ahmadi Lahijani^{1*}, M. Kafi¹, A. Nezami¹, J. Nabati², and J. E. Erwin³

ABSTRACT

Micro-propagated potato plantlets (*cvs.* Agria and Fontane) were grown in growth chambers under controlled conditions to determine the effect of different levels of CO₂ concentrations (400 vs. 800 μmol mol⁻¹) on physiological and biochemical traits and yield of minitubers, at the Ferdowsi University of Mashhad, in 2015. Irrespective of the cultivar, the elevated CO₂ significantly increased Net photosynthesis (N_p) and leaf Dark Respiration (R_d) compared to the control at 34 and 57 days after transplanting. Higher N_p under the elevated CO₂ resulted in a higher accumulation of leaflet starch and soluble sugar content. The elevated CO₂, compared to the ambient, induced allocation of more dry matter to the underground parts, especially tubers. CO₂ Enrichment did not significantly affect the number of tubers and mean tuber weight, however, the elevated CO₂ increased yield of Agria and Fontane by 17 and 39%, respectively. The yield of Fontane was increased more than Agria when exposed to elevated CO₂, mainly due to greater mean tuber weight than tuber number. The number of large size tubers increased under elevated CO₂. The results showed that the greater mean tuber weight might affect tuber yield more than the number of tubers. Our findings suggest that rising levels of CO₂ in minituber production systems could be beneficial to improve productivity and tuber yield.

Keywords: Carbohydrate, Dark respiration, Dry matter partitioning, Photosynthetic rate, Starch.

INTRODUCTION

Potato (*Solanum tuberosum* L.), with annual production of 382 million tons and 20 t ha⁻¹ yield, is ranked as the fourth important crop in the world (FAOSTAT, 2014). Potato is mostly propagated vegetatively through tubers, which decrease the seed quality by continued propagation mainly due to degradation in health status. Viral diseases are a major limiting factor in potato production. Virus free micro-propagated plantlets can produce disease-free seed

potato, preventing yield losses (Struik and Wiersema, 2012). Minitubers are healthier potatoes with a faster multiplication rate for a greater tuber yield (Sharma and Pandey, 2013).

For many decades, extensive researches have been carried out on improvement of agronomic and physiological performance and, consequently, increasing crop yield (Ahmadi Lahijani *et al.*, 2018; Shahkoomahally and Shahkoomahally, 2017; Skraly *et al.*, 2018; Struik and Wiersema, 2012). Applying supplemental CO₂ to increase yields and quality of agricultural

¹Department of Agrotechnology, Faculty of Agriculture, Ferdowsi University of Mashhad, Islamic Republic of Iran.

²Research Center of Plant Sciences, Ferdowsi University of Mashhad, Islamic Republic of Iran.

³Department of Horticultural Science, University of Minnesota, 305 Alderman Hall, St. Paul, MN 55108, USA.

* Corresponding author; e-mail: mj.ahmadi@mail.um.ac.ir



crops is becoming common among commercial growers (Li *et al.*, 2013). Due to the limitation of photosynthesis of C₃ plants under the present concentration of atmospheric CO₂, higher levels of CO₂ will increase both carbon fixation and plant biomass production (Aranjuelo *et al.*, 2005).

Increasing atmospheric CO₂ directly enhances photosynthesis in C₃ plants leading to increase in leaf sugar and starch content (Ainsworth and Lemonnier, 2018). Rising CO₂ concentration affects plants at the levels of molecular, developmental, and physiological functions. Changes in the plant functions may affect their ability to capture resources (e.g. leaf size or leaf number), and also alter the resource allocation among different parts, which may significantly influence yield and production (Gray and Brady, 2016).

Increased potato tuber yield due to elevated CO₂ has been reported in diverse studies (Finnan *et al.*, 2005; Högy and Fangmeier, 2009). Lawlor and Mitchell (1991) found that indeterminate crops like potato are supposed to respond more strongly to higher levels of CO₂ compared to determinate crops such as cereals. Potato and other crops with large belowground sinks for carbon and apoplastic mechanisms of phloem loading were suggested to highly respond to elevated CO₂ (Komor *et al.*, 1996). Finnan *et al.* (2005) attributed the increase in the tuber yield at elevated CO₂ to not only the higher tuber number and individual tuber weight but also a combination of both.

Elevated CO₂ stimulates the production of above- and belowground biomass, but their magnitude are varied with cultivar, plant species, growing condition, and agronomic practice (Finnan *et al.*, 2005; Reddy *et al.*, 2010). Several studies in potato reported that elevated CO₂ concentration enhanced greater dry matter partitioning in the tubers to aboveground parts. (Chen and Setter, 2012; Fleisher *et al.*, 2008a; Fleisher *et al.*, 2008b). Cell division and expansion of sink organs are highly responsive to the higher levels of CO₂ concentrations, hence, stronger sinks attract more assimilates and, consequently, enhance tuber size and tuber number (Chen and Setter, 2012).

Leaf dark respiration is a determinant factor controlling the growth and maintenance of plant tissues (Li *et al.*, 2013). However, the effect and mechanism of elevated CO₂ on the leaf dark respiration is not fully elucidated. Previous findings reported the effects of CO₂ elevation on respiration rate (Gomez-Casanovas *et al.*, 2007; Leakey *et al.*, 2009; Long *et al.*, 2004). Most of up- or down-regulation of respiration under elevated CO₂ conditions depends on developmental stage, plant species, growth conditions, and gas exchange measurement errors (Long *et al.*, 2004). Li *et al.* (2013) reported that higher dark respiration of tomato leaves under a higher concentration of CO₂ was due to greater content of proteins, carbohydrates, and transcripts involved in the glycolysis pathway, tricarboxylic acid cycle, and mitochondrial electron transport energy metabolism.

The number of tubers and mean tuber weight are of particular importance in minituber production systems. Although previous researches have reported the positive effects of elevated levels of CO₂ on photosynthetic rate, growth, and yield of potato plants, no reports documented the effect of elevated CO₂ concentrations on the respiratory pathway and its relationship with productivity and tuber yield of micro-propagated potato plantlets. The present work aimed to assay the potential positive effects of elevated level of CO₂ on physiological and biochemical traits, minituber production, and dry matter partitioning in two cultivars of potato plantlets (*cvs.* Agria and Fontane).

MATERIALS AND METHODS

Plant Materials and Growth Conditions

Uniform potato plantlets (*cvs.* Agria and Fontane) were transplanted in a sterile medium containing a mixture of perlite-coco peat (1:1, v:v) in plastic pots (diameter, 15 cm; depth, 30 cm; one per pot) in the controlled environment chambers (Conviroon, Winnipeg, Canada) at Ferdowsi University of Mashhad, in 2015. Plantlets

were derived from *in vitro* micro-propagated shoot tips grown in agar culture plates containing the Murashige and Skoog medium supplemented with 3% sucrose at Yekta Seed Technology Company, Mashhad, Iran. The plantlets were grown inside the chambers at Photosynthetic Photon Flux Density (PPFD) of 400 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ on the leaf surface supplied by fluorescent/incandescent lamps, photoperiod 12/12 (h) (day/night), day/night air temperature 24/16°C, and relative humidity of 50/60% (day/night). The plantlets were separated into two groups. Each group (including 9 pot per cultivar) were placed into a separate growth chamber deferring in CO₂ concentrations (400±10 and 800±10 $\mu\text{mol mol}^{-1}$) as “ambient” and “elevated” CO₂, respectively. The pots were subjected to CO₂ concentrations from the first day of transplanting. The CO₂-enriched air (a mixture of ambient air with commercial CO₂) from a compressed gas cylinder was injected into each chamber at a flow rate of one L min^{-1} , which was continuously monitored by a calibrated infrared gas analyzer (High-performance CO₂ meter, 77535, China). All plants were watered daily to prevent water stress and were fertilized with the standard Hoagland’s solution every five days (Hoagland and Arnon, 1950).

Sampling

Gas exchange variables were measured from the third fully expanded upper leaves at 34 (stolonization stage) and 57 (tuberization stage) Days After Transplanting (DAT). The same leaves were collected for the metabolic assays at the same stages.

Measurements of Gas Exchange Variables

Fully expanded leaves were enclosed in a gas exchange leaf chamber using a portable photosynthesis system (HCM-1000, Waltz,

Germany). Net photosynthesis was estimated at a PPFD of 1,500 $\mu\text{mol m}^{-2} \text{s}^{-1}$ (Wang *et al.*, 2004). Leaves were equilibrated for 5 minutes at 1,500 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD before each measurement. The leaf temperature, relative humidity inside the cuvette, and leaf-to-air vapor pressure deficit were set at 25°C, 50%, and 50 Pa kPa^{-1} , respectively. The leaf dark respiration was obtained through averaging three CO₂ efflux rates at zero PPFD for each plant at the end of the daily dark period from 05:00 to 07:00 hours. Three leaflets per replication were analyzed (n=9).

Leaf Soluble Carbohydrates and Starch Content

To assay the total Soluble Carbohydrates of leaves (SC), the method of Dubois *et al.* (1956) was used. A sample of 100 mg leaf fresh weight was homogenized in 70% methanol using a mortar and pestle. Total soluble carbohydrates content of leaflets were measured by a glucose standard curve. The method of Schlegel (1956) was applied to assay the Starch content of leaves (ST). Residuals of the soluble carbohydrates experiment were rinsed three times using perchloric acid. Absorbance was spectrophotometrically recorded at 485 nm. The starch content of leaves was determined through a glucose standard curve.

Crop Harvest and Growth Parameters

Plants were harvested 90 DAT, rinsed and separated into stems, leaves, roots, and tubers. Total green Leaf Number (LN), and number of tubers per plant (Katny *et al.*, 2005) were counted. Mean Tuber Weight (MTN) and tuber Yield per plant (Y) were also measured. The harvested tubers were classified based on smaller diameter in three size categories (< 15, 15-25, and > 25 mm). The above- and below-ground parts of the plants were weighted and then dried out at 75 °C until constant mass and weighed. The total plant DM was calculated



by adding the values obtained for roots, shoots, and tubers. Dry matter partitioning among the plant parts was calculated as a percentage of DM accumulated in the leaves, stems, roots, and tubers to the total plant DM. Leaf Area (LA) was measured using an automatic leaf area meter (Li-3100, LI-COR, Lincoln, NE).

Statistical Analysis

The experimental design was a completely randomized design in a factorial (two cultivars and two CO₂ concentrations) scheme with three replications. Data were presented as the mean ± standard deviation. Statistical analyses were performed by Analysis Of Variance (ANOVA) in SAS software. Differences between treatment means were determined by Fisher's Least Significant Differences (LSD) test at 0.05 probability level.

RESULTS

Gas Exchange Variables

Elevated CO₂ concentration increased *N_p* and *R_D* of cultivars at stolonization and tuberization stages (Table 1). A greater *N_p* at 34 DAT compared with 57 DAT was recorded, but *R_D* was greater at 57 DAT in both cultivars (Table 2). Net photosynthesis

rate of Agria increased more than Fontane at both measurement dates when exposed to the elevated CO₂. Elevated CO₂ increased *N_p* of Agria by 104 and 84% at 34 and 57 DAT, respectively, while the increases in Fontane were 51 and 12% at similar stages, respectively, compared with the ambient (Table 2).

There was no significant difference in *R_D* between cultivars at 34 or 57 DAT, but elevated CO₂ significantly influenced *R_D* at both stages (Table 1). Elevated CO₂ increased *R_D* of Agria at 34 and 57 DAT by 177 and 5%, respectively, while, these were 192 and 8% in Fontane at the same stages, respectively. Both at ambient and elevated CO₂, Agria showed a higher *R_D* at 34 DAT than 57 DAT. This superiority reversed in Fontane, with higher *R_D* under both CO₂ conditions at 57 DAT. (Table 2).

Leaf Soluble Carbohydrates and Starch Content

There were no significant differences between the cultivars in SC either at 34 or 57 DAT (Table 1). Despite producing higher values of SC at elevated CO₂, no statistical difference was observed between the CO₂ treatments (Table 3). Exposure to elevated CO₂ increased ST of Agria at both measurement times more than Fontane. Elevated CO₂ increased ST of Agria by 37 and 78% at 34 and 57 DAT, respectively,

Table 1. ANOVA results of gas exchange and chemical parameters at 34 and 57 days after transplanting.

Variables	Cultivar (V)	CO ₂ concentration (C)	V×C	r ²	CV
34 DAT^a					
Leaf Soluble sugars (SC)	NS	NS	NS	0.48	8.1
Leaf Starch content (ST)	NS	*	NS	0.68	13.2
Net photosynthetic rate (N _p)	**	**	**	0.96	11.2
Dark Respiration (R _D)	NS	**	NS	0.61	16.4
57 DAT^a					
Leaf Soluble sugars (SC)	NS	NS	NS	0.29	10.6
Leaf Starch content (ST)	**	**	*	0.96	7.7
Net photosynthetic rate (N _p)	**	**	NS	0.94	9.8
Dark respiration (R _D)	NS	**	NS	0.81	16.4

* Significant at $P < 0.05$, ** Significant at $P < 0.01$, NS: Not Significant, LSD test. ^a DAT: Day After Transplanting, 34 DAT (Stolonization stage), 57 DAT (Tuberization stage).

Table 2. Effects of CO₂ levels on photosynthetic rate and dark respiration of potato cultivars at 34 and 57 days after transplanting.^a

Treatment	N_p ($\mu\text{mol m}^{-2} \text{s}^{-1}$)		R_D ($\mu\text{mol m}^{-2} \text{s}^{-1}$)	
	34 DAT	57 DAT	34 DAT	57 DAT
C1 AM	17.95±2.53	13.85±3.79	0.63±0.79	3.19±0.06
C1 EL	36.68±1.50	25.5±1.26	1.75±0.27	3.34±1.33
C2 AM	13.89±1.38	14.1±3.14	0.53±0.33	4.11±0.12
C2 EL	21±1.35	15.85±1.75	1.55±0.67	4.46±0.88

^a Values are mean±SD of three replicates (n= 9). C1; Agria, C2; Fontane, AM; Ambient CO₂, EL; Elevated CO₂, N_p ; Net photosynthesis rate, R_D ; Dark Respiration, DAT; Day After Transplanting, 34 DAT (Stolonization stage), 57 DAT (Tuberization stage).

Table 3. Effects of CO₂ treatments on leaf soluble sugar and starch content of potato cultivars at 34 and 57 days after transplanting.^a

Treatment	Soluble carbohydrates (mg g^{-1} FW)		Starch (mg g^{-1} FW)	
	34 DAT	57 DAT	34 DAT	57 DAT
C1 AM	24.1±1.67	35.2±0.79	109.5±5.73	135.8±6.61
C1 EL	25.2±2.11	32.4±1.16	151.1±9.33	242.8±18.5
C2 AM	23.0±2.07	33.4±1.49	110.8±7.05	109.4±10.07
C2 EL	25.7±2.42	34.5±3.47	137.1±4.88	171.8±19.1

^a Values are mean±SD of three replicates (n= 9). Symbols as defined under Table 2.

while the increases in Fontane were 24 and 57% at the similar stages, respectively, compared with the ambient.

Growth Parameters and Dry Matter Partitioning

Leaf, stem, and total dry matter of the potato plants were influenced by either cultivar or CO₂ concentration (Table 4). Plants grown at elevated CO₂ produced 33, 42, and 35% greater leaf, stem, and total shoot DM, respectively, compared with ambient CO₂ conditions (Table 5). Dry matter partitioning to different parts of the plants was influenced by cultivar and CO₂ (Table 4). Carbon dioxide enrichment enhanced allocation of DM to the underground parts, especially tubers, and decreased it to the aerial parts of the plants (Table 5 and Figure 1).

The increase in shoot DM of Fontane was greater than Agria at elevated CO₂, and Agria also gained more tuber DM (Table 5). While Fontane allocated more biomass to the roots at elevated CO₂, Agria greatly allocated biomass to the tubers (Table 5). Tuber DM was greatly increased in Agria and Fontane under elevated CO₂ compared with the control. However, Fontane produced greater tuber dry matter than Agria either at ambient or elevated CO₂ (Table 5). Exposure to elevated CO₂ decreased shoot DM allocation in Agria and Fontane by 17 and 12%, respectively, compared to the ambient. In contrast, DM of the underground parts of Agria and Fontane increased by 13 and 7%, respectively (Figure 1).

The analysis of leaf area per plants revealed that there was a significant increase in the leaf area associated with higher level of CO₂ (Table 4). Leaf areas of Fontane and Agria increased by 31% and 12%, respectively, at elevated CO₂ compared to



Table 4. ANOVA results of growth parameters of potato cultivars.

Variables	Cultivar (V)	CO ₂ concentration (C)	V×C	r ²	CV
Leaf Dry Matter (LDM)	**	**	NS	0.85	10.4
Stem Dry Matter (SDM)	NS	**	NS	0.74	14.4
Total Shoot Dry Matter (SHDM)	**	**	NS	0.89	8.5
Root Dry Matter (RDM)	**	*	NS	0.80	25.5
Tuber Dry Matter (TDM)	NS	**	NS	0.75	22.5
Total Dry Matter (TDM)	**	**	NS	0.85	15.2
Leaf Area (LA)	**	**	NS	0.88	9.1
Leaf Number (LN)	**	**	NS	0.95	5.1
Tuber Yield (Y)	NS	**	NS	0.80	18.5
Tuber number (TN)	NS	NS	NS	0.74	15.1
Mean Tuber Weight (MTW)	NS	NS	NS	0.71	24.5
Small tuber (< 15 mm)	NS	NS	NS	0.40	26.7
Average tuber (15-25 mm)	NS	NS	NS	0.52	19.4
Large tuber (> 25 mm)	NS	**	NS	0.68	14.1

*Significant at $P < 0.05$, ** Significant at $P < 0.01$, NS: Not Significant, LSD test.

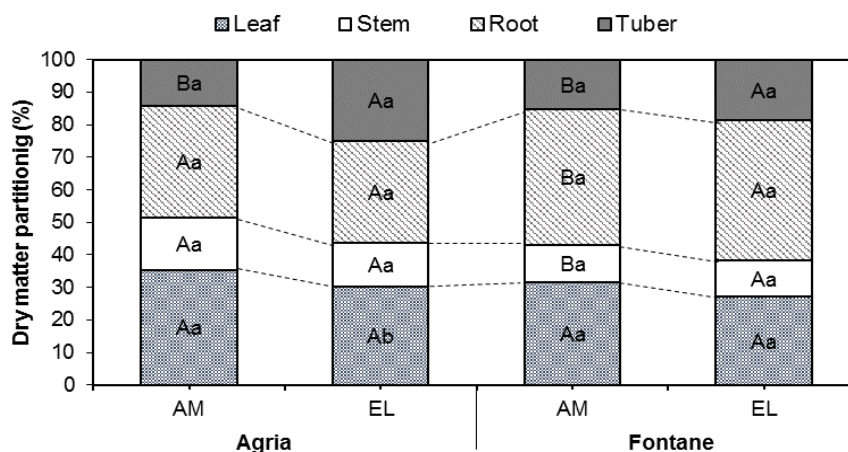


Figure 1. Effect of elevated CO₂ on dry matter partitioning of potato cultivars. AM: 400 ppm, EL: 800 ppm. Different lowercase letters indicate a significant difference between CO₂ levels for the same cultivar, whereas different uppercase letters indicate a significant difference between cultivars at the same CO₂ level, at $P < 0.05$, according to LSD test.

Table 5. Effects of CO₂ treatments on growth parameters of potato cultivars.^a

Treatment	LDM (g plant ⁻¹)	SDM (g plant ⁻¹)	SHDM (g plant ⁻¹)	RDM (g plant ⁻¹)	TUDM (g plant ⁻¹)	TDM (g plant ⁻¹)
C1 AM	1.59±0.27	0.71±0.17	2.31±0.34	1.55±0.52	0.64±0.16	4.51±0.74
C1 EL	2.05±0.06	0.91±0.13	2.96±0.18	2.12±0.65	1.70±0.47	6.78±1.17
C2 AM	2.08±0.33	0.74±0.11	2.82±0.34	2.73±0.63	1.10±0.09	6.56±0.81
C2 EL	2.84±0.06	1.15±0.06	4.00±0.01	4.5±0.91	1.95±0.51	10.46±1.42

^a The measurements were carried out when the plants were 90 days old. Values are mean±SD of three replicates (n=9). C1; Agria, C2; Fontane, AM; Ambient CO₂, EL; Elevated CO₂, LDM; Leaf Dry Matter, SDM; Stem Dry Matter, SHDM; Shoot Dry Matter, RDM; Root Dry Matter, TUDM; Tuber Dry Matter, TDM; Total Dry Matter.

the ambient (Table 6). The plants grown under elevated CO₂ conditions produced a higher number of leaves (Table 4). Fontane had a greater LN under both conditions, but elevated CO₂ increased LN of Agria greatly (Table 6).

Tuber Yield

Elevated CO₂ significantly stimulated the tuber yield per plant (Table 4). Fontane produced a greater tuber yield than Agria when exposed to elevated CO₂ (Table 6). Elevated CO₂ increased the yield of Agria and Fontane by 21 and 65%, respectively. There was no significant effect of higher CO₂ concentration on the TN and MTW of the plants (Table 4). Although insignificantly, a greater MTW was recorded at elevated CO₂ in Fontane compared with the ambient conditions (Table 6). Tuber number of Agria was increased (26%) more than Fontane (4%) under elevated CO₂ conditions, however, the change was not significant (Table 4). CO₂ enrichment significantly increased the proportion of large size tubers (> 25 mm) of the plants compared with the ambient conditions (Table 4). The percent distribution of medium size tubers (15-25 mm) was also higher in Agria at elevated CO₂ compared with the ambient conditions, although, the effect was not significant (Figure 2).

DISCUSSION

Photosynthesis of C₃ plants is not fully saturated at the current atmospheric levels of CO₂. Rising atmospheric CO₂ concentration has been elucidated to enhance the rate of photosynthesis and influences the growth and yield of C₃ plants (Reddy *et al.*, 2010). Our results revealed that *Np* was significantly stimulated in both cultivars of potato. Chen and Setter (2012) suggested that the availability of sufficient sink is important to increase the photosynthetic response of C₃ plants at elevated CO₂. Although there was no significant effect of elevated CO₂ on the number of tubers and mean tuber weight, the plants grown at elevated CO₂ produced slightly more tuber number and mean tuber weight, which could serve as a greater sink strength. The greater number of tubers increase sink strength, thereby, reducing the feedback inhibition of starch build-up on the photosynthetic rate. Finnan *et al.* (2005) stated that the tuberization process would be stimulated under elevated CO₂ conditions, and a greater number of tubers would increase the sink strength to attract more assimilates. Aien *et al.* (2014) also reported that the photosynthetic rate of two potato cultivars increased at 30 and 60 DAT under elevated levels of CO₂. They concluded that increase in the rate of photosynthesis under elevated CO₂ might be attributed to the higher number of tubers under such conditions.

Table 6. Effects of CO₂ treatments on yield, yield component, leaf area, and leaf number per plant of potato cultivars.^a

Treatment	Yield (g plant ⁻¹)	Mean tuber weight (g)	Tuber number (plant ⁻¹)	Leaf area (cm ² plant ⁻¹)	Leaf number
C1 AM	10.0±1.87	3.8±1.00	2.6±0.57	399±49.4	15.7±0.75
C1 EL	12.1±3.12	3.7±1.28	3.3±0.32	449±15.2	17.7±0.39
C2 AM	9.3±1.33	3.7±0.24	2.5±0.40	510±8.66	22.4±0.80
C2 EL	15.4±2.01	5.9±1.32	2.6±0.35	671±10.6	26.1±0.23

^a The measurements were carried out when the plants were 90 days old. Each value represents the mean±SD. C1; Agria, C2; Fontane, AM; Ambient CO₂, EL; Elevated CO₂.

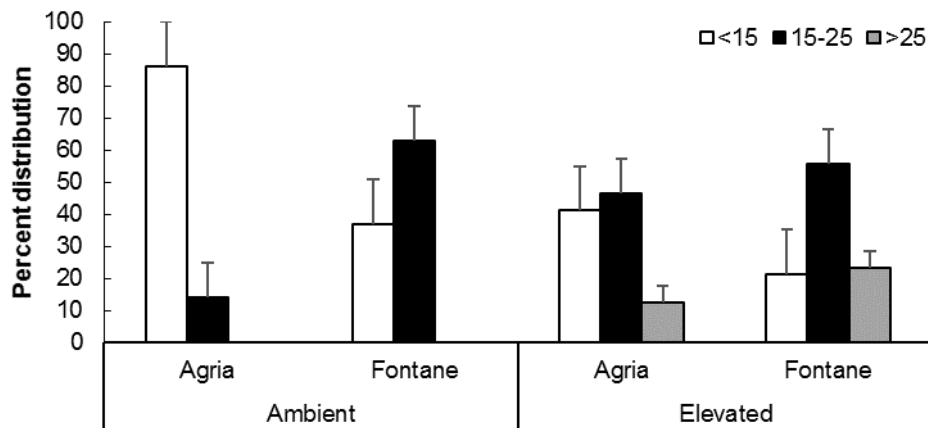


Figure 2. Percentage of the number of tubers produced per size category for different cultivars under ambient and elevated CO₂ conditions. Ambient; 400 $\mu\text{mol mol}^{-1}$, Elevated; 800 $\mu\text{mol mol}^{-1}$. Size category (mm). Vertical bar shows Standard Error of the difference (SE).

Measurement of the leaf dark respiration revealed that the respiratory metabolism was up-regulated under elevated CO₂. The metabolic analysis showed that starch and soluble carbohydrates content increased under elevated CO₂. Because of the enhanced photosynthetic rate at elevated CO₂, higher accumulation of carbohydrates might be a consequence of higher substrate availability to stimulate the respiratory pathway (Aranjuelo *et al.*, 2011; Aranjuelo *et al.*, 2006). Energy demand and substrate supply are able to control respiration rates, which indicates that plants regulate carbon and nutrients allocation to increase photosynthesis and growth (González-Meler *et al.*, 2009). A common consequence of growing plants at elevated CO₂ is a higher production of photoassimilates and a higher concentration of carbohydrates in the plant tissues (Kimball *et al.*, 2002). Li *et al.* (2013) also reported a significant increase in the leaf carbohydrate content and dark respiration in tomato plants when exposed to elevated CO₂. Higher leaf dark respiration because of greater leaf carbohydrate and starch content helps plants to meet energy demand for growth and maintenance. Carbohydrates play a vital role in starch

biosynthesis and tuber growth and are known as the stimulators of tuber initiation (Ramawat and Merillon, 2013). An increase in carbohydrates content activate a number of genes involved in starch biosynthesis and increase the tubers capacity for storing more photoassimilates (Kloosterman *et al.*, 2005).

This study showed greater tuber yield in the plants grown under elevated CO₂ conditions. This is consistent with Fleisher *et al.* (2013) who found substantial stimulation in the tuber yield of potato exposed to elevated CO₂. Lawson *et al.* (2001), however, reported no significant effect of such conditions on the tuber yield. Numerous studies on the effect of elevated CO₂ on potato yield have reported an increase in the tuber number associated with a greater tuber yield (Aien *et al.*, 2014). However, in spite of increase in tuber yield, a decrease in the tuber number was also reported under higher CO₂ concentration (Finnan *et al.*, 2005; Finnan *et al.*, 2002). The results of the CHIP project showed that the increase in potato tuber number under elevated CO₂ was insufficient to quantify the tuber yield, therefore, mean tuber weight also contributes to increasing yield (Craigon *et al.*, 2002). In agreement with this, it was

observed that the mean tuber weight was more stimulated than the tuber number when plants were exposed to a higher level of CO₂.

The results of our study showed that elevated CO₂ concentration increased the percent distribution of medium and large size tubers compared with the ambient conditions (Figure 2). The results are in agreement with Högy and Fangmeier (2009) who reported an increase in the size of individual tubers under elevated CO₂. They concluded that the stimulation of potato tuber yield in response to CO₂ enrichment was mainly due to increased growth of the formerly initiated tubers than an increase in the number of tubers. During the tuber formation stage, numerous potential tubers initiate on potato plants, but some of them do not develop due to limited assimilate flow toward the tubers (Struik, 2007). We observed that elevated CO₂ increased leaf area and leaf number of the plants. Therefore, it is possible that competition between the newly developing leaves with the tubers resulted in a lower positive effect of greater assimilate production on the mean tuber weight and the tuber number of plants under elevated CO₂ concentrations.

Despite higher *Np* in Agria under both CO₂ levels, the tuber yield of Fontane increased more than Agria under elevated CO₂. The higher tuber yield of Fontane was closely associated with the greater photosynthetic units (LA and LN) than *Np* on the leaf area basis. Additionally, the lower leaf starch content and the higher tuber dry matter of Fontane compared with Agria likely indicated the better translocation of photoassimilates to underground parts in this cultivar. Fleisher *et al.* (2008a) observed no significant changes in leaf area of potato plants at higher CO₂ concentration. They concluded that higher productivity of the potato plants was due to higher assimilation rate under such conditions. However, Finnan *et al.* (2005) concluded that the increase in potato tuber yield under elevated CO₂ could be a result of either a higher photosynthetic rate per leaf

area unit or an increase in leaf area, and likely a combination of both.

The results of this study showed that exposure to elevated CO₂ altered dry matter allocation to the tubers. This is in agreement with the results of previous studies that reported a greater increase in mean tuber weight and tuber number of potato plants (Fleisher *et al.*, 2008b; Högy and Fangmeier, 2009). Studies on the effect of higher CO₂ concentration on the growth and productivity of tomato (Cong and Tanksley, 2006) and Arabidopsis (Horiguchi *et al.*, 2006) indicated that cell proliferation and expansion in the sink organs are important factors for developing sink capacity. Higher levels of CO₂ concentration shift partitioning of photoassimilates toward the tubers. Since cell division in the sinks is highly responsive to elevated CO₂, it results in producing larger sinks to store more assimilates (Chen and Setter, 2012). However, Chen and Setter (2012) reported that a greater potato tuber growth under elevated CO₂ is more related to stimulated tuber cell proliferation than cell expansion.

CONCLUSIONS

Elevated CO₂ concentration increased dry matter partitioning toward the underground parts of the plants, especially tubers. There was a significant increase in the tuber yield of both cultivars. However, no significant increase in the number of tubers or mean tuber weight of cultivars was observed at elevated CO₂. It seems that a slight increase in either tuber number or individual tuber weight potentially contributed to increase in tuber yield under elevated CO₂. The increase in tuber yield of Fontane seemed to be more associated with mean tuber weight, while, in Agria, it was more associated with the number of tubers under elevated CO₂. Additionally, Elevated level of CO₂ significantly increased the number of large size tubers as a portion of total tuber numbers, which would be beneficial for a better establishment of plants in the field.



Our results demonstrated that the higher level of CO₂ concentration stimulated leaf soluble carbohydrates and starch content of both cultivars, which could serve as a greater substrate for increasing the leaf dark respiration. Overall, these results suggest that raising the levels of CO₂ in minituber production systems could be beneficial to stimulate the productivity and tuber yield.

ACKNOWLEDGEMENTS

The authors sincerely thank Dr. Mohammad Zare Mehrjerdi for technical support of this work and Dr. Sama Dangla for manuscript revision.

REFERENCES

1. Ahmadi Lahijani, M. J., Kafi, M., Nezami, A., Nabati, J. and Erwin, J. 2018. Effect of 6-Benzylaminopurine and Abscisic Acid on Gas Exchange, Biochemical Traits, and Minituber Production of Two Potato Cultivars (*Solanum tuberosum* L.). *J. Agri. Sci. Tech.*, **20(1)**: 129-139.
2. Aien, A., Pal, M., Khetarpal, S. and Kumar Pandey, S. 2014. Impact of Elevated Atmospheric CO₂ Concentration on the Growth, and Yield in Two Potato Cultivars. *J. Agr. Sci. Tech.*, **16**: 1661-1670.
3. Ainsworth, E. A. and Lemonnier, P. 2018. Phloem Function: A Key to Understanding and Manipulating Plant Responses to Rising Atmospheric [CO₂]? *Curr. Opin. Plant Biol.*, **43**: 50-56.
4. Aranjuelo, I., Cabrera-Bosquet, L., Morcuende, R., Avicé, J. C., Nogués, S., Araus, J. L., Martínez-Carrasco, R. and Pérez, P. 2011. Does Ear C Sink Strength Contribute to Overcoming Photosynthetic Acclimation of Wheat Plants Exposed to Elevated CO₂? *J. Exp. Bot.*, **62(11)**: 3957-3969. doi:10.1093/jxb/err095
5. Aranjuelo, I., Irigoyen, J. J., Perez, P., Martinez-Carrasco, R. and Sanchez-Diaz, M. 2006. Response of Nodulated Alfalfa to Water Supply, Temperature and Elevated CO₂: Productivity and Water Relations. *Environ. Exp. Bot.*, **55(1)**: 130-141.
6. Aranjuelo, I., Pérez, P., Hernández, L., Irigoyen, J. J., Zita, G., Martínez-Carrasco, R. and Sánchez-Díaz, M. 2005. The Response of Nodulated Alfalfa to Water Supply, Temperature and Elevated CO₂: Photosynthetic Downregulation. *Physiol. Plant.*, **123(3)**: 348-358.
7. Chen, C. -T. and Setter, T. L. 2012. Response of Potato Dry Matter Assimilation and Partitioning to Elevated CO₂ at Various Stages of Tuber Initiation and Growth. *Environ. Exp. Bot.*, **80**: 27-34.
8. Cong, B. and Tanksley, S. D. 2006. FW2.2 and Cell Cycle Control in Developing Tomato Fruit: A Possible Example of Gene Co-Option in the Evolution of a Novel Organ. *Plant Mol. Biol.*, **62(6)**: 867-880.
9. Craigan, J., Fangmeier, A., Jones, M., Donnelly, A., Bindi, M., De Temmerman, L., Persson, K. and Ojanpera, K. 2002. Growth and Marketable-Yield Responses of Potato to Increased CO₂ and Ozone. *Eur. J. Agron.*, **17(4)**: 273-289.
10. Dubois, M., Gilles, K. A., Hamilton, J. K., Rebers, P. and Smith, F. 1956. Colorimetric Method for Determination of Sugars and Related Substances. *Anal. chem.*, **28(3)**: 350-356.
11. FAOSTAT. 2014. *Agriculture Organization of the United Nations. Rome*, (17.05.14).
12. Finnan, J., Donnelly, A., Jones, M. and Burke, J. 2005. The Effect of Elevated Levels of Carbon Dioxide on Potato Crops: A Review. *J. Crop Improv.*, **13(1-2)**: 91-111.
13. Finnan, J. M., Donnelly, A., Burke, J. I. and Jones, M. B. 2002. The Effects of Elevated Concentrations of Carbon Dioxide and Ozone on Potato (*Solanum tuberosum* L.) Yield. *Agr. Ecosyst. Environ.*, **88(1)**: 11-22.
14. Fleisher, D., Wang, Q., Timlin, D., Chun, J. -A. and Reddy, V. 2013. Effects of Carbon Dioxide and Phosphorus Supply on Potato Dry Matter Allocation and Canopy Morphology. *J. Plant Nut.*, **36(4)**: 566-586.
15. Fleisher, D. H., Timlin, D. J. and Reddy, V. 2008a. Elevated Carbon Dioxide and Water Stress Effects on Potato Canopy Gas Exchange, Water Use, and Productivity. *Agr. Forest Meteorol.*, **148(6)**: 1109-1122.
16. Fleisher, D. H., Timlin, D. J. and Reddy, V. R. 2008b. Interactive Effects of Carbon Dioxide and Water Stress on Potato Canopy Growth and Development. *Agron. J.*, **100(3)**: 711-719.

17. Gomez-Casanovas, N., Blanc-Betes, E., Gonzalez-Meler, M. A. and Azcon-Bieto, J. 2007. Changes in Respiratory Mitochondrial Machinery and Cytochrome and Alternative Pathway Activities in Response to Energy Demand Underlie the Acclimation of Respiration to Elevated CO₂ in the Invasive *Opuntia Ficus-Indica*. *Plant Physiol.*, **145(1)**: 49-61.
18. González-Meler, M. A., Blanc-Betes, E., Flower, C. E., Ward, J. K. and Gomez-Casanovas, N. 2009. Plastic and Adaptive Responses of Plant Respiration to Changes in Atmospheric CO₂ Concentration. *Physiol. Plant.*, **137(4)**: 473-484.
19. Gray, S. B. and Brady, S. M. 2016. Plant Developmental Responses to Climate Change. *Dev. Biol.*, **419(1)**: 64-77.
20. Hoagland, D. R. and Arnon, D. I. 1950. The Water-Culture Method for Growing Plants without Soil. *Circular. California Agric. Exp. Station*, **347(2nd Edit)**.
21. Högy, P. and Fangmeier, A. 2009. Atmospheric CO₂ Enrichment Affects Potatoes: 1. Aboveground Biomass Production and Tuber Yield. *Eur. J. Agron.*, **30(2)**: 78-84.
22. Horiguchi, G., Ferjani, A., Fujikura, U. and Tsukaya, H. 2006. Coordination of Cell Proliferation and Cell Expansion in the Control of Leaf Size in *Arabidopsis thaliana*. *J. Plant Res.*, **119(1)**: 37-42.
23. Katny, M. A. C., Hoffmann-Thoma, G., Schrier, A. A., Fangmeier, A., Jäger, H. -J. and van Bel, A. J. 2005. Increase of Photosynthesis and Starch in Potato under Elevated CO₂ Is Dependent on Leaf Age. *J. Plant Physiol.*, **162(4)**: 429-438.
24. Kimball, B., Kobayashi, K. and Bindi, M. 2002. Responses of Agricultural Crops to Free-Air CO₂ Enrichment. *Adv. Agron.*, **77**: 293-368.
25. Kloosterman, B., Vorst, O., Hall, R. D., Visser, R. G. and Bachem, C. W. 2005. Tuber on a Chip: Differential Gene Expression during Potato Tuber Development. *Plant Biotech. J.*, **3(5)**: 505-519.
26. Komor, E., Orlich, G., Weig, A. and Köckenberger, W. 1996. Phloem Loading—Not Metaphysical, Only Complex: Towards a Unified Model of Phloem Loading. *J. Exp. Bot.*, **47(Special Issue)**: 1155-1164.
27. Lawlor, D. and Mitchell, R. 1991. The Effects of Increasing CO₂ on Crop Photosynthesis and Productivity: A Review of Field Studies. *Plant Cell Environ.*, **14(8)**: 807-818.
28. Lawson, T., Craigon, J., Black, C., Colls, J., Tulloch, A. -M. and Landon, G. 2001. Effects of Elevated Carbon Dioxide and Ozone on the Growth and Yield of Potatoes (*Solanum tuberosum*) Grown in Open-Top Chambers. *Environ. Pollut.*, **111(3)**: 479-491.
29. Leakey, A. D., Xu, F., Gillespie, K. M., McGrath, J. M., Ainsworth, E. A. and Ort, D. R. 2009. Genomic Basis for Stimulated Respiration by Plants Growing under Elevated Carbon Dioxide. *Proc. Nat. Acad. Sci.*, **106(9)**: 3597-3602.
30. Li, X., Zhang, G., Sun, B., Zhang, S., Zhang, Y., Liao, Y., Zhou, Y., Xia, X., Shi, K. and Yu, J. 2013. Stimulated Leaf Dark Respiration in Tomato in an Elevated Carbon Dioxide Atmosphere. *Sci. Rep.*, **3:3433**: 1-8. doi:10.1038/srep03433
31. Long, S. P., Ainsworth, E. A., Rogers, A. and Ort, D. R. 2004. Rising Atmospheric Carbon Dioxide: Plants FACE the Future. *Annu. Rev. Plant Biol.*, **55**: 591-628.
32. Ramawat, K. G. and Merillon, J. -M. 2013. *Bulbous Plants: Biotechnology*. CRC Press, USA. 450PP.
33. Reddy, A. R., Rasineni, G. K. and Raghavendra, A. S. 2010. The Impact of Global Elevated CO₂ Concentration on Photosynthesis and Plant Productivity. *Curr. Sci.*, **99(1)**: 46-57.
34. Schlegel, H. G. 1956. Die Verwertung Orgngischer Souren Durch Chlorella Lincht. *Planta.*, **47**: 510-526.
35. Shahkoomahally, E. and Shahkoomahally, S. 2017. Investigating of N and K Fertilizers on Yield and Components of Soybean [*Glycine max* (L.) Merr.]. *J. Agri. Sci.*, **9(10)**: 85.
36. Sharma, A. K. and Pandey, K. 2013. Potato Mini-Tuber Production through Direct Transplanting of *In Vitro* Plantlets in Green or Screen Houses: A Review. *Potato J.*, **40(2)**: 95-103.
37. Skraly, F. A., Ambavaram, M. M., Peoples, O. and Snell, K. D. 2018. Metabolic Engineering to Increase Crop Yield: From Concept to Execution. *Plant Sci.*, **273**:23-33. doi:10.1016/j.plantsci.2018.03.011
38. Struik, P. C. 2007. Above-Ground and Below-Ground Plant Development. In: "*Potato Biology and Biotechnology*:"



- Advances and Perspectives*", (Ed.): Verugdenhil, D. Elsevier, Amsterdam, PP. 219-236.
39. Struik, P. C. and Wiersema, S. G. 2012. *Seed Potato Technology*. Wageningen Press, Wageningen, 383 PP.
40. Wang, X., Roger Anderson, O. and Griffin, K. L. 2004. Chloroplast Numbers, Mitochondrion Numbers and Carbon Assimilation Physiology of *Nicotiana sylvestris* as Affected by CO₂ Concentration. *Environ. Exp. Bot.*, **51**: 21-31.

تأثیر افزایش دی اکسید کربن بر تبادل گازی، ویژگی های بیوشیمیایی و عملکرد ریزغده های سیب زمینی (*Solanum tuberosum* L.)

م. ج. احمدی لاهیجانی، م. کافی، ا. نظامی، ج. نباتی، ج. ا. اروین

چکیده

به منظور بررسی اثر سطوح مختلف دی اکسید کربن (۴۰۰ و ۸۰۰ میکرومول بر مول) بر ویژگی های بیوشیمیایی و فیزیولوژیک و عملکرد ریزغده، گیاهچه های ریزازدیاد شده سیب زمینی (رقم آگریا و فونتانه) در شرایط کنترل شده در دانشگاه فردوسی مشهد در سال ۱۳۹۴ کشت شدند. بدون توجه به رقم، افزایش دی اکسید کربن موجب افزایش معنی دار فتوسنتز و تنفس تاریکی در ۳۴ و ۵۷ روز پس از کاشت در مقایسه با شاهد شد. فتوسنتز بیشتر تحت تاثیر افزایش دی اکسید کربن منجر به تجمع بیشتر نشاسته و کربوهیدرات محلول در برگ شد. افزایش دی اکسید کربن منجر به تسهیم ماده خشک بیشتر به سمت اندام های زیرزمینی به ویژه ریزغده ها شد. افزایش دی اکسید کربن تاثیر معنی داری بر تعداد و میانگین وزن ریزغده نداشت، اما بهرحال، افزایش دی اکسید کربن باعث افزایش عملکرد غده آگریا و فونتانه به ترتیب به میزان ۱۷ و ۳۹ درصد شد. عملکرد فونتانه بیش از آگریا در اثر افزایش دی اکسید کربن افزایش یافت که عمدتاً به دلیل میانگین وزن غده بیشتر بود. نتایج نشان داد که میانگین وزن غده بیش از تعداد غده عملکرد را تحت تاثیر قرار می دهد. با توجه به نتایج، به نظر می رسد که افزایش غلظت دی اکسید کربن در سیستم های تولید ریزغده سیب زمینی می تواند برای بهبود تولید کنندگی و عملکرد ریزغده سودمند باشد.