Importance of Hormonal Elicitors in Inducing Morphine Biosynthesis in the Cell Culture of (*Papaver bracteatum* Lindl.)

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ABSTRACT

Plant cells have enough capacity to produce many of secondary metabolites, similar to the whole plants. Elicitation is one of the most significant methods to increase the synthesis of secondary metabolites in the medicinal plants. The purpose of this study was to investigate the effect of three different hormones on alkaloids production in suspension culture of *Papaver bracteatum* Lindl., in order to identify the relationship between the alkaloid biosynthesis and gene expression. Inducible factors initiate Benzylisoquinoline Alkaloids (BIAs) biosynthesis in Opium poppy. The current study investigated the accumulation of alkaloids content and Tyrosine/Dopamine Carboxylase (TYDC), Berberine Bridge Enzyme (BBE), Salutaridinol Acetyl Transferase (SAT), and Codeinone Reductase (COR) gene transcripts in suspension culture of *P. bracteatum*. Indole-3-Acetic Acid (IAA), Indole Butyric Acid (IBA) and Gibberellic Acid (GA) were used as hormonal elicitors in the suspension cultures with three different doses and two timings along with the control. This research showed the induction of morphine alkaloid in the suspension culture of *P. bracteatum*. Elicitation by 20 mg L⁻¹ concentration of IAA after 48 h indicated significant increase in morphine amount. Comparison among genes revealed that the expression levels of COR dramatically increased while TYDC, BBE and SAT had no significant difference compared to the control. After elicitation by IAA, IBA, and GA, the highest levels of morphine were measured as 243.2, 207.2, and 178.1 mg g⁻¹, respectively. The results demonstrated that timing had a significant effect on the hormonal elicitation: 48 h treatment could induce more morphine alkaloids compared to 24 hours treatments.

Keywords: Elicitation, Gene expression, Gibberellic acid, Indole butyric acid, Indole-3-acetic acid.

INTRODUCTION

Iranian Poppy (*Papaver bracteatum*) is an important medicinal plant that is the main source of the opium alkaloids. In addition, opium and the isoquinoline alkaloids morphine, codeine, noscapine, papaverine, and thebaine are isolated from the dried material. Some accessions of *P. bracteatum* contain 98 percent thebaine of whole alkaloids. Therefore, this species is an outstanding source of basic materials to produce narcotics, 0.7 to 1.3 percent in the roots, 26 percent in dry latex or 3.5 percent in the dry, ripe capsules (Day et al., 1986). The original habitat of *P. bracteatum* is the mountainous areas of Iran (the Alborz Mountains and Mahabad regions), Turkey and Transcaucasian Russia. The flowers of *P. bracteatum* are red with long petals along with big blackish spots in the middle. According to the cytogenetic researches, this species is diploid with the chromosomal base number 2n= 14 (2X) (Balažová et al., 2008; Cline and Coscia, 1988).

Alkaloid biosynthesis in plants involves many catalytic steps, catalyzed by enzymes that belong to a wide range of protein families (Bohm, 1967; Rostampour et al., 2009; Sharafi et al., 2013 Nasiri et al., 2016). Thebaine, as one of the alkaloid components,
Figure 1. Biosynthesis pathway of thebaine, codeine, morphine, papaverine in *P. somniferum* TYDC, BBE, SAT and COR.
significant increase in comparison to the control group. The results indicated a 10-fold increase in morphinan alkaloids content in comparison with the control plants (Larkin et al., 2007). We propose research design to increase the amount of alkaloid in cell culture by using three different hormones, namely, Indole-3-Acetic Acid (IAA), Indole Butyric Acid (IBA) and Gibberellic Acid (GA) as elicitors. In this project, we aimed to establish the cell cultures and induce gene activation using elicitors. Finding effective elicitors to induce more alkaloids and their influence on the gene expression of alkaloid biosynthesis was also evaluated. Additionally, the current study investigated the accumulation of alkaloids content and TYDC, BBE, SAT and COR gene transcripts in suspension culture of P. bracteatum.

MATERIALS AND METHODS

Plant Material

Seeds used as experimental material were obtained from the Research Institute of Forest and Rangelands, Tehran, Iran (Polur, 14305 accession number). The ethanol 70% (v/v) was utilized to sterilize the seeds for 1 minute (3 times) and sodium hypochlorite solution 50% for 8 minutes, then, washing with double sterile distilled water. Modified MS medium was used for germination of seeds (Rostampour et al., 2009; Zare et al., 2014). The cell cultures of P. bracteatum were grown at 16 hours photoperiod cool white fluorescent light. Seeds germinated after adding 7.5 g L\(^{-1}\) agar in the dark room at 25\(^\circ\)C (Kamo et al., 1987). The explants were separated from roots after germination during four weeks of the planting.

Cell Suspension Culture

The explants of P. bracteatum were transferred to the modified MS medium including 0.5 mg L\(^{-1}\) BAP, 0.5 mg L\(^{-1}\) 2,4-D and 7.5 g L\(^{-1}\) agar were added in the solid medium for callus induction. Callus culture was the subculture of the fresh medium every 21 days. In the next step, to initiate the suspension cultures, friable calli (50 mg) was transferred to 25 mL fresh medium with the same concentration of hormones in 100 mL flasks were incubated on the shaker at 120 rpm, in a dark room at 25±2\(^\circ\)C. The suspension cultures were established after three different subcultures. The percentage of volume and weight cells was measured. The cells entered the growth phase and, gradually, the rate of cell growth diminished (Farjaminezhad et al., 2013). Elicitations on samples were performed after 23 days of inoculation when the cell cultures were in the exponential phase of growth. To investigate the effects of IAA, IBA and GA, hormones elicitors were added in three different concentrations (5, 10, and 20 mg L\(^{-1}\)) to the 25 mL suspension culture of P. bracteatum. Additionally, an equal amount of medium with no elicitation was used as the control. Then, all cells were collected by filter papers after 24 and 48 hours treatments to evaluate the optimal time and elicitors for the highest level of alkaloid production.

Alkaloid Extraction and HPLC Analysis

Freeze-drier (Lyotrap, Fisher Scientific, USA) was used to dry the cells. Alkaloids were extracted by the mixture of methanol and HCL. Alkaloids were measured by HPLC system along with standard compounds. HPLC analysis was performed on a Knauer Smartline HPLC system (UV detector). A volume of 25 mL of samples was injected in a C-18 reverse-phase Knauer Smartline column (201Tp54 VDAC). The mobile phase for alkaloid elution was 50% methanol and 50% water. (Tisserat and Berhow, 2009). In addition, thebaine, morphine, codeine, papaverine, noscapine, and sanguinarine as standards were obtained from Temad Chemical Co (Tehran, Iran).

Gene Expression

Approximately 100 mg of cells treated with elicitors were applied for the expression on the analysis of alkaloid biosynthetic genes. Then, they were grounded to a powder in
liquid nitrogen, and the total RNAs were isolated with the RNeasy Plant Mini Kit (Qiagen). The quality and quantity of the RNAs were estimated by spectrophotometry and Nano drop. Reverse transcription was performed using a First Strand cDNA Synthesis Kit (Fermentas), and Elongation Factor 1 alpha (EFa1) was the reference gene with primers generated for it according to the method used in Zhang’s study (2008). RT-PCR was performed by TYDC, BBE, SAT, and COR in genes to analyze the expression levels of the gene. These primers were designed using Primer3 and Perlprimer software programs. The primers are listed in Table 1. The Real-Time PCR on SYBR Green was conducted using an iQ5 (BIORAD) machine under the following conditions: the process was performed at 95°C for 90 seconds followed by 40 cycles of 95°C for 15 seconds, 60°C for 20 seconds, and 72°C for 20 seconds. In addition, data were normalized using the levels of EFa1 transcripts (Ranasinghe et al., 2008).

Statistical Analysis

The statistical significance of data was evaluated by one-way Analysis Of Variance (ANOVA). Each treatment contained three independent replicates and SAS Statistic performed the experiments three times. Furthermore, mean comparisons were carried out using Duncan’s multiple range tests at a probability level of 0.05.

Table 1. Primer sequences designed for real-time amplification of alkaloid biosynthetic genes of *P. bracteatum*.

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequences</th>
</tr>
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<tbody>
<tr>
<td>COR-F</td>
<td>CCACCATTTGACCAGCATTAG</td>
</tr>
<tr>
<td>COR-R</td>
<td>GGGGCTCATCTCCACCTTAATC</td>
</tr>
<tr>
<td>SAT-F</td>
<td>GGGGCTCATCTCCACCTTAATC</td>
</tr>
<tr>
<td>SAT-R</td>
<td>GCTGGTAAAGAAGCCGAAC</td>
</tr>
<tr>
<td>TYDC-F</td>
<td>AACCACAATGACCCCTATGATGA</td>
</tr>
<tr>
<td>TYDC-R</td>
<td>GACCTGGCTTCTAACTGGATAAC</td>
</tr>
<tr>
<td>BBE-F</td>
<td>GGGTATCTGCTGGTGTG</td>
</tr>
<tr>
<td>BBE-R</td>
<td>AATGCATTCCAGACGTGTTATC</td>
</tr>
<tr>
<td>Elf1a-F</td>
<td>AGATGATTCCACAACAGCCA</td>
</tr>
<tr>
<td>Elf1a-R</td>
<td>CCTTGATGACACCAACAGCA</td>
</tr>
</tbody>
</table>
a). Using indole butyric acid as elicitor on cell culture of Iranian poppy indicated that morphine enhanced after 48 hours treatment. The elicitation of cells with 10 mg L⁻¹ Indole butyric acid after 48 hours treatment had significant effect on morphine production. At this level, morphine reached the 2.03-fold level of the control. The highest amount of morphine alkaloid was 9.09-fold more than the lowest level treatment with Indole butyric acid (Figure 2-b).

Gibberellic acid elicitation with 20 mg L⁻¹ after 24 hours treatment produced more morphine compared to other concentrations. The application of cells with Gibberellic acid at a 20 mg L⁻¹ dose after 48 hours treatment had a significant effect on morphine production. The highest morphine yield obtained at the 20 mg L⁻¹ concentration enhanced this alkaloid 1.75-fold in comparison with the control. The application of Gibberellic acid at the 20 mg L⁻¹ dose after 48 hours increased the quantity of morphine 2.81-fold more than the treatment by 10 mg L⁻¹ in 24 hours. The lowest morphine yield was obtained at the 10 mg L⁻¹ concentration in 24 hours after treatment with all the three elicitors; indole-3-acetic acid, indole butyric acid and gibberellic acid hormones (Figure 2-c).

We used Real-time PCR to recognize the responses of the genes TYDC, BBE, SAT, and COR to test whether IAA, IBA and GA hormones could influence transcription of the prominent genes in alkaloid biosynthesis. The current study revealed that indole-3-acetic acid increased the accumulation of morphine through the mechanism shown in Figure 1, in which the morphine content continuously increases until 48 hours after the application of 20 mg L⁻¹. Additionally, the main difference between the highest and the lowest morphine concentration for those plants treated with indole-3-acetic acid is related to the COR gene. At the 48 h mark, the expression levels of COR up-regulated in comparison to the control, but TYDC, BBE and SAT did not show...
any significant difference with the control. On the other hand, in the lowest amounts of morphine after 24 h elicitation with 10 mg/l of aforementioned hormone, no difference between the transcript levels of TYDC and SAT in comparison with the control group was observed. There was no difference between the transcript levels of TYDC and SAT in comparison with the control group. Also, the gene expression of BBE and COR down-regulated (Figure 3-a). As a result, Plants treated with Indole butyric acid with 10 mg L\(^{-1}\) after 48 hours treatment showed only significant changes in the transcription levels of BBE, SAT and COR compared to the control. Additionally, in the lowest amounts of morphine after elicitation by Indole butyric acid, transcriptional levels of TYDC and BBE showed down-regulation. However, the expression level of COR did not change significantly in comparison with the control, and SAT transcript levels were up and down regulated at this concentration (Figure 3-b).

The transcript levels of genes in those plants exposed to Gibberellic acid with the highest level of morphine and lowest level of this alkaloid were more variable. At 48 hours treatment with 20 mg L\(^{-1}\) hormone, the expression levels of BBE and SAT up-regulated, but TYDC and COR had no significant difference compared to the control. In contrast, gene expression levels for TYDC, BBE, SAT and COR in the 10 mg L\(^{-1}\) at 24 hours, considered as the lowest production of morphine alkaloid, did not significantly alter in comparison with the control (Figure 3-c).

In some other researches, different
accessions contained various alkaloids as the major secondary metabolites. Comparing the distinct accessions of *P. bracteatum* indicated that some strains, such as West Germany, only had thebaine as the main alkaloids, while the other seven strains in this report contained isothebaine as the major alkaloid (Kamimura *et al*., 1967). Research on the natural abundance of morphinan alkaloids in *P. bracteatum* along with the results from feeding codeine-one-[16-3H] and codeine-[16-3H] indicated that this species could reduce codeine to the codeine (Hodges *et al*., 1977). In another study on the tissue culture of *P. bracteatum*, accumulation of large amounts of dopamine and a small quantity of thebaine and sanguinarine were reported (Larkin *et al*., 2007). Even though in most studies thebaine has been found as a major alkaloid in the *P. bracteatum* (Farjaminezhad *et al*., 2013; Zare *et al*., 2014, Rostampour *et al*., 2009), there are some reports that show the other alkaloids can be detected from the cell culture of *P. bracteatum*. For instances, sanguinarin accumulated in the cell suspension of *P. bracteatum* after using fungal elicitors and hormones and thebaine were produced in the media without hormones (Cline and Coscia, 1988). Brochmann-Hanssen and Wunderly (1978) confirmed that codeine could be found in *P. bracteatum*, though morphine was not found. This result emphasized that the biosynthesis O-6-demethylation of thebaine to neopinone can be considered restrictive step. The existence of neopinone and codeinone can be caused by acidic condition in the plant, which hydrolyzes nonenzymatically the enol ether group of thebaine. Therefore, these two compounds can be reduced to the related alcohols in the plant quickly (Brochmann-Hanssen and Wunderly, 1978). Moreover, codeine oxidation could produce 14-hydroxycodeine and norcodeine in *Streptomyces griseus* (Kunz *et al*., 1985). It can be concluded that, morphine can be produced by two substantial genes, 6-O-demethylase and codeine O-demethylase and/or codeinone reductase genes, which can proceed the genetic pathway of morphinan alkaloid thebaine to the codeine and morphine. According to the special patent, genetically modified *P. bracteatum* with nucleic acid and amino acid sequence variants of the aforementioned genes are capable of producing more quantity of an alkaloid selected from codeine, oripavine and/or morphine in comparison with a wild type of this plant. Also, salutaridinol 7-O-acetyltransferase gene converts the phenanthrene alkaloid salutaridinol to salutaridinol-7-O-acetate, which is an important enzyme to biosynthesize morphine in *P. somniferum*. The transcripts of this gene have been identified in *P. bracteatum* in addition to *P. somniferum* (Grothe *et al*., 2001). Over expression of codeinone reductase in Opium Poppy, which is essential to produce morphine, codeine and various semi-synthetic analgesics, could increase morphinan alkaloids (Larkin *et al*., 2007).

Unlike primary data, which supported the hypothesis that *P. bracteatum*, has outstanding potential to produce thebaine alkaloid, our results demonstrated that morphine alkaloid can be detected in the cell culture of this plant. In addition, the investigations of gene expression elucidate genetic pathway and molecular basis of alkaloids. The current study revealed that using Indole-3-Acetic Acid (IAA), Indole butyric acid, and Gibberellic acid hormones treatments as elicitors can enhance alkaloid production in the cell culture of *P. bracteatum* with significant changes in the expression of the studied genes.

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REFERENCES


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