
J. Sharifi-Rad$^1$, M. Sharifi-Rad$^2$, and J. A. Teixeira da Silva$^3$

**ABSTRACT**

In this research, two field crops (*Zea mays* L. and *Phaseolus vulgaris* L.), two medicinal plants (*Hyssopus officinalis* L. and *Nigella sativa* L.) and two weeds (*Amaranthus retroflexus* L. and *Taraxacum officinale* F. H. Wigg) were separately treated with three concentrations of SiO$_2$ nanoparticles (400, 2,000, and 4,000 mg L$^{-1}$). The effects of these treatments on morphological and biochemical characteristics of the plants were assessed, including germination, root and shoot length, root and shoot fresh weight, root and shoot dry weight, photosynthetic pigments, total carbohydrates, total protein, total amino acid, and proline content. In the crops and medicinal plants, 400 mg L$^{-1}$ SiO$_2$ NPs significantly increased seed germination, root and shoot lengths, fresh weights (except for *H. officinalis*) and dry weights, photosynthetic pigments, total protein, and total amino acid (except for *H. officinalis*). In weeds, as SiO$_2$ NP concentration increased from 400 to 4,000 mg L$^{-1}$, germination, root and shoot lengths, fresh and dry weights, and photosynthetic pigments as well as total protein decreased. Total carbohydrates in all plants decreased significantly, except for *A. retroflexus* at 400 mg L$^{-1}$ SiO$_2$ NPs. In all plant species, with increasing SiO$_2$ NP concentration, proline content increased significantly. According to these results, a lower concentration of SiO$_2$ NPs can have beneficial effects on morphological, physiological, and biochemical characteristics of plants.

**Keywords:** Germination, Photosynthetic pigments, Total amino acid, Total carbohydrates, Total protein.

**INTRODUCTION**

Nanobiotechnology is one of the most important developing sciences. The uses of products from this technology have increased in agriculture, industry, medicine and the military (Gruère, 2012; Qu *et al*., 2013; Sharifi-Rad *et al*., 2014). Nanotechnology has many uses in all steps of processing, production, packaging, storing, and transport of agricultural products. The use of nanotechnology in agriculture also has environmental benefits and, as an interdisciplinary science, can be used as a powerful tool to empower the agricultural sector and in important cases such as crop production, use less pesticides and fertilizers to maintain crops for longer periods (Chinnamuthu and Boopathi, 2009; Mousavi and Rezaei, 2011).

With the rapid development of nanotechnology and its applications, nano-structured materials have been widely used in the fields of
biomedicine, pharmaceutical, and other industries (Maghbl et al., 2012; Cao et al., 2013; Rad et al., 2013a; Rad et al., 2013b). Nanometer silicon dioxide (nano-SiO$_2$) is one of the most popular nanomaterials used in industrial manufacturing, packaging, synthesis of high-molecule composite materials and ceramics, disease labeling, drug delivery, cancer therapy and biosensors (Sahoo et al., 2007). Most experiments to date that have assessed the effects of SiO$_2$ NPs on plants have considered physiological, biochemical or morphological characteristics of usually one or maximum two plants (Slomberg and Schoenfisch, 2012; Siddiqui and Al-Whaibi, 2014). Generally, studies on the effects of Si NPs on plants are limited and study on how the same levels of exposure to SiO$_2$ NPs would affect a wide range of crops using morphological, physiological and biochemical characteristics is lacking.

There are some important crops that can be included in such a study. For example, maize (Zea mays L.; Poaceae), one of the most widely cultivated crops, is a major component in the diet of many developing countries such as Iran and it is one of the crops with the most biotechnological potential for energy production and other industrial applications (McLaren, 2005). Also, common bean (Phaseolus vulgaris L.; Fabaceae), which is the most important grain legume and plays an important role in human nutrition as a valuable source of minerals, protein, fibres, calories and vitamins, is produced in a wide range of climatic conditions in Iran and elsewhere (Sadeghi and Cheghamirza, 2012). Medicinal plants can also be included in such a study: Hyssopus officinalis L. (hyssop; Lamiaceae), a perennial plant with a long history of traditional and medicinal uses, is an endemic Iranian species of the genus Hyssopus (Khazaie et al., 2008). Also, traditionally, H. officinalis – named Zufa in Iran – has been used as a tonic, antiseptic, cough reliever carminative and expectorant (Khazaie et al., 2008). In spite of having a somewhat bitter taste, H. officinalis is frequently used as a condiment and minty flavor in the food industry. Black seed (Nigella sativa L.; Ranunculaceae) has a long history of traditional medicinal use. It is also used as a flavouring agent and food additive in many countries, especially in developing countries (Sharifi-Rad et al., 2014). Black seed oil is reportedly beneficial due to its content of over 100 components such as vitamins, trace elements, and aromatic oils (Ali and Blunden, 2003). To further diversify the kind of crops for this type of research, weed plants can be studied. For example, redroot amaranth (Amaranthus retroflexus L.; Amaranthaceae) is a critically destructive weed distributed worldwide, particularly in farmlands, wastelands and gardens that is difficult to control due to its extreme vigor, flexibility and prolific seed production. Redroot amaranth can seriously influence the growth of crops and pollute crop seeds, causing immense losses to agricultural production (Costea et al., 2004). Also, dandelion (Taraxacum officinale F. H. Wigg; Asteraceae), a weed and perennial herbaceous plant that is native to the entire northern hemisphere, has several varieties and subspecies and grows as weed in wild, moist pastures in temperate areas (Chaitanya et al., 2013).

This study, the first of its kind for abovementioned set of plants, aimed to assess how the same levels of exposure to SiO$_2$ NPs would affect a wide range of crops using morphological, physiological and biochemical characteristics. The plants considered for the present study were two field crops (Zea mays L., Phaseolus vulgaris L.), two medicinal plants (Hyssopus officinalis L., Nigella sativa L.), and two weeds (Amaranthus retroflexus L., Taraxacum officinale F. H. Wigg) so as to have a wider and more representative assessment of the impact of SiO$_2$ NPs on plants.

**MATERIALS AND METHODS**

**Plant Materials and SiO$_2$ NP Treatments**

Seeds of maize (var. ‘KSC 704’), common bean (cv. ‘Naz’), hyssop (var. angustifolius), black cumin (cv. ‘Bafit’), redroot pigweed (var. retroflexus) and dandelion were obtained from the Iranian Agricultural Organization, Zabol City, Iran. SiO$_2$ NPs were purchased from Sigma–Aldrich (St. Louis, MO, USA; 99.5% purity; 10-20 nm in size; white; spherical shape; density of 3.8 g cm$^{-3}$). Figure 1 shows a Transmission Electron Microscopic (TEM) image of the tested NPs. The NPs were dispersed
Responses of Plants Exposed to SiO\textsubscript{2} Nanoparticles

Figure 1. Transmission Electron Microscopy (TEM) image of SiO\textsubscript{2} NPs used in this study (Sigma-Aldrich).

in distilled water at three concentrations (400, 2,000, and 4,000 mg L\textsuperscript{-1}) and then sterilized at 120°C for 20 minutes. An A KQ 5200DE model ultrasonicator (Shumei Instrument Factory, Kunshan, China), applied at 60 Hz for 30 minutes, was used for easy dispersion of NPs without precipitation. For the best dispersion, SiO\textsubscript{2} NPs were sufficiently shaken after sonication to break up agglomerates. For each SiO\textsubscript{2} NP treatment (400, 2,000, and 4,000 mg L\textsuperscript{-1}), each concentration was prepared separately, without dilution, by weighing NPs and dispersing them in distilled water. The SiO\textsubscript{2} NP suspensions were dispersed by sonication for 20 min before use. Seeds were surface sterilized by incubating for 20 minutes in 5% (w/v) sodium hypochlorite followed by four washes with sterile distilled water. Seeds were weighed directly with each concentration for 30 minutes. An ultrasonicator (Shumei Instrument Factory, Kunshan, China), applied at 60 Hz for 30 minutes, was used for easy dispersion of NPs without precipitation. For the best dispersion, SiO\textsubscript{2} NPs were sufficiently shaken after sonication to break up agglomerates. For each SiO\textsubscript{2} NP treatment (400, 2,000, and 4,000 mg L\textsuperscript{-1}), each concentration was prepared separately, without dilution, by weighing NPs and dispersing them in distilled water. The SiO\textsubscript{2} NP suspensions were dispersed by sonication for 20 min before use. Seeds were surface sterilized by incubating for 20 minutes in 5% (w/v) sodium hypochlorite followed by four washes with sterile distilled water. Seeds were weighed directly with each concentration for 30 minutes. An ultrasonicator (Shumei Instrument Factory, Kunshan, China), applied at 60 Hz for 30 minutes, was used for easy dispersion of NPs without precipitation. For the best dispersion, SiO\textsubscript{2} NPs were sufficiently shaken after sonication to break up agglomerates. For each SiO\textsubscript{2} NP treatment (400, 2,000, and 4,000 mg L\textsuperscript{-1}), each concentration was prepared separately, without dilution, by weighing NPs and dispersing them in distilled water. The SiO\textsubscript{2} NP suspensions were dispersed by sonication for 20 min before use. Seeds were surface sterilized by incubating for 20 minutes in 5% (w/v) sodium hypochlorite followed by four washes with sterile distilled water. Seeds were weighed directly with each concentration for 30 minutes. An ultrasonicator (Shumei Instrument Factory, Kunshan, China), applied at 60 Hz for 30 minutes, was used for easy dispersion of NPs without precipitation. For the best dispersion, SiO\textsubscript{2} NPs were sufficiently shaken after sonication to break up agglomerates. For each SiO\textsubscript{2} NP treatment (400, 2,000, and 4,000 mg L\textsuperscript{-1}), each concentration was prepared separately, without dilution, by weighing NPs and dispersing them in distilled water. The SiO\textsubscript{2} NP suspensions were dispersed by sonication for 20 min before use. Seeds were surface sterilized by incubating for 20 minutes in 5% (w/v) sodium hypochlorite followed by four washes with sterile distilled water.

Root and Shoot Length

After 14 days, root and shoot length were measured in mm with a ruler.

Fresh and Dry Weight of Seedling Shoots and Roots

The shoot and root FW of 30-day-old seedlings was assessed for all treatments since, by 30 days, plants had well developed leaf and root systems suitable for transplanting. The DW of shoots and roots was determined after placing the entire plantlets (with organs divided) in an oven for 48 hours at 75°C. Shoot and root DW were expressed as g pot\textsuperscript{-1}, measured with 10, 25, and 30 seedlings pot\textsuperscript{-1} for crops, medicinal plants and weeds, respectively, and four replicates per treatment.

Measurement of Photosynthetic Pigments

The content of photosynthetic pigments in control and treated plants was measured according to Lichtenthaler et al. (1987). Two hundred mg of randomly selected leaf tissue
was weighed and pulverized with a mortar and pestle in liquid nitrogen. Large pieces were completely pulverized with 80% acetone, and the final volume was brought to 25 mL. The resulting solution was centrifuged in a refrigerated Beckman GS-15R centrifuge (365702; bench-model; Ontario, Canada) at 4,800 rpm for 20 minutes. The supernatant was used to measure the content of chlorophyll (chl) $a$, $b$ and carotenoids. Light absorption of the plant extracts was determined by a Shimadzu A160 spectrofluorometer (Shimadzu, Japan) at 470, 645, 646.8, 663 and 663.2 nm.

**Total Carbohydrate Determination**

In this study, carbohydrate content was determined by the phenol-sulfuric acid method (Rao and Pattabiraman, 1989). Briefly, to 1 mL of each leaf aqueous extract, 50 µL of 80% phenol (Merck, Darmstadt, Germany) and then 3 mL of 98% sulfuric acid (Merck) were added. The leaf aqueous extract was prepared by crushing 2 g of fresh leaf tissue with 3 mL of distilled water then centrifuging the resulting solution for 10 minutes at 200 rpm, separating the supernatant and using it for the next steps of the experiment. The mixture was vortexed for 1 min, then, kept at room temperature for 30 minutes. The absorbance was read at 490 nm with the same spectrophotometer.

**Total Protein Determination**

In this study, total protein was extracted from the leaf tissue of each plant by homogenization with a mortar and pestle on ice at 4°C in an extraction buffer containing 50 mM Tris–HCl (pH 7.5), 2 mM EDTA, 0.5 mM EGTA, 1 mM PMSF and 1% Triton X-100. The homogenates were centrifuged in a Beckman GS-15R centrifuge at 14,000 rpm for 10 minutes at 4°C. The soluble protein concentration in the homogenate supernatant was determined using Bovine Serum Albumin (BSA) (Merck) as standard (Bradford, 1976).

**Total Amino Acid Assay**

Total amino acids were determined by reaction with ninhydrin using glycine as standard according to Sun et al. (2006). Briefly, to 500 mL solution of amino acids, 1 mL of 80% acetic acid and 1 mL of ninhydrin solution (2 mg ninhydrin in 50% ethanol) were added, then, mixed for 15 minutes. This mixture was placed at 100°C for 15 minutes and then at 70°C for 10 minutes. Finally, 5 mL of 2-propanol (50%) was added and absorbance was read at 570 nm. Different concentrations of the amino acid glycine were used to create a standard curve.

**Free Proline Amino Acid Content Assay**

Proline was extracted and measured according to the method of Bates et al. (1973). Leaf samples (0.8 g) were extracted with 3% sulphosalicylic acid. One mL of extracts was placed for 1 hour in boiling water. Then, 2 mL ninhydrin, 2 mL glacial acetic acid, and 4 mL of ice-cold toluene were added sequentially. Proline content was measured by a Shimadzu UV 1601 spectrophotometer at 520 nm and calculated as µmol g$^{-1}$ DW against a proline standard (Sigma-Aldrich).

**Statistical Analysis**

All data were analyzed as a completely randomized design with four replications. Data were expressed as means±Standard Error (SE). Means between treatments were separated by Analysis Of Variance (ANOVA) and then statistically significant differences between means were assessed by Duncan’s new Multiple Range Test (DMRT) and SPSS software version 11.5 (IBM SPSS, New York, USA) at $P \leq 0.05$.

**RESULTS**

**Effects of SiO$_2$ NPs on Seed Germination**

The effects of SiO$_2$ NPs on seed germination are shown in Figure 2. In all six plants, there
were significant differences between the control and all treatments: 400 mg L\(^{-1}\) SiO\(_2\) NPs significantly stimulated seed germination more than the control, but 2,000 and 4,000 mg L\(^{-1}\) SiO\(_2\) NPs significantly reduced seed germination relative to the control and 400 mg L\(^{-1}\) SiO\(_2\) NPs.

### Effects on Root and Shoot Length

The effects of SiO\(_2\) NPs on root and shoot length are shown in Table 1. In *Z. mays*, *P. vulgaris*, *H. officinalis* and *N. sativa*, there was a significant difference between the control and all treatments with 400 mg L\(^{-1}\) SiO\(_2\) NPs, which significantly stimulated root and shoot length more than the control. However, 2,000 and 4,000 mg L\(^{-1}\) SiO\(_2\) NPs significantly reduced root and shoot length relative to the control and 400 mg L\(^{-1}\) SiO\(_2\) NPs. In *A. retroflexus* and *T. officinale*, the control resulted in significantly longer roots and shoots than when any concentration of SiO\(_2\) NPs was used, except for 400 mg L\(^{-1}\) SiO\(_2\) NPs in *A. retroflexus*.

### Effects on Root and Shoot Fresh and Dry Weight

The effects of SiO\(_2\) NP concentration on root and shoot FW and DW are shown in Table 2. In both crops and in both medicinal plants, in most cases, there was a significant difference in root FW and DW between the control and all treatments: 400 mg L\(^{-1}\) SiO\(_2\) NPs significantly stimulated root and shoot FW and DW more than the control, but 2,000 and 4,000 mg L\(^{-1}\) SiO\(_2\) NPs significantly reduced root and shoot FW and DW relative to the control and 400 mg L\(^{-1}\) SiO\(_2\) NPs. Notable differences were for *H. officinalis* shoot FW, where the control and 400 mg L\(^{-1}\) SiO\(_2\) NPs were not significantly different, and for *H. officinalis* root DW, and *N. sativa* root FW and DW, in which 2,000 and 4,000 mg L\(^{-1}\) SiO\(_2\) NPs were not significantly different. The two weeds showed a different trend to the crops and medicinal plants, with the control resulting in significantly higher root and shoot FW and DW than all SiO\(_2\) NP concentrations, except for *A.*
Table 1. Effect of SiO₂ NPs on Root Length (RL) and Shoot Length (SL) (in mm) of six test species.

<table>
<thead>
<tr>
<th>SiO₂ (mg L⁻¹)</th>
<th>Zea mays</th>
<th>Phaseolus vulgaris</th>
<th>Hyoscyamus niger</th>
<th>Nigella sativa</th>
<th>Spinacia oleracea</th>
<th>Taraxacum officinale</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (0)</td>
<td>12.6 ± 0.09 b</td>
<td>29.3 ± 0.04 b</td>
<td>16.6 ± 0.12 b</td>
<td>19.4 ± 0.14 b</td>
<td>8.6 ± 0.09 b</td>
<td>12.4 ± 0.09 b</td>
</tr>
<tr>
<td>400</td>
<td>17.7 ± 0.14 a</td>
<td>37.4 ± 0.12 a</td>
<td>25.6 ± 0.09 a</td>
<td>13.2 ± 0.12 a</td>
<td>13.2 ± 0.14 a</td>
<td>19.2 ± 0.10 a</td>
</tr>
<tr>
<td>2000</td>
<td>9.4 ± 0.08 c</td>
<td>18.3 ± 0.09 c</td>
<td>7.4 ± 0.07 c</td>
<td>14.3 ± 0.08 c</td>
<td>4.1 ± 0.14 c</td>
<td>10.3 ± 0.09 c</td>
</tr>
<tr>
<td>4000</td>
<td>5.6 ± 0.1 d</td>
<td>14.6 ± 0.07 d</td>
<td>4.2 ± 0.10 d</td>
<td>11.2 ± 0.14 d</td>
<td>3.9 ± 0.07 c</td>
<td>9.8 ± 0.09 d</td>
</tr>
</tbody>
</table>

*Values indicate means ± SE. Different lower case letters indicate significant differences (DMRT) in RL and SL assessed separately for each plant species across treatments (P< 0.05). n= 4.

Table 2. Effect of SiO₂ NPs on root and shoot Fresh Weight (FW) and root and shoot Dry Weight (DW) (in g/pot) of six test species.

<table>
<thead>
<tr>
<th>SiO₂ (mg L⁻¹)</th>
<th>Zea mays</th>
<th>Phaseolus vulgaris</th>
<th>Hyoscyamus niger</th>
<th>Nigella sativa</th>
<th>Spinacia oleracea</th>
<th>Taraxacum officinale</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (0)</td>
<td>3.96 ± 0.00 b</td>
<td>1.86 ± 0.00 b</td>
<td>7.94 ± 0.01 b</td>
<td>3.84 ± 0.01 b</td>
<td>3.60 ± 0.14 b</td>
<td>1.92 ± 0.02 b</td>
</tr>
<tr>
<td>400</td>
<td>8.30 ± 0.09 a</td>
<td>4.23 ± 0.01 a</td>
<td>16.6 ± 0.09 a</td>
<td>6.22 ± 0.01 a</td>
<td>5.83 ± 0.14 a</td>
<td>2.70 ± 0.15 a</td>
</tr>
<tr>
<td>2000</td>
<td>3.43 ± 0.01 c</td>
<td>1.26 ± 0.00 c</td>
<td>6.87 ± 0.01 c</td>
<td>2.84 ± 0.01 c</td>
<td>2.10 ± 0.09 c</td>
<td>0.96 ± 0.00 c</td>
</tr>
<tr>
<td>4000</td>
<td>2.96 ± 0.01 d</td>
<td>0.90 ± 0.00 d</td>
<td>5.94 ± 0.01 d</td>
<td>1.80 ± 0.18 d</td>
<td>1.98 ± 0.04 c</td>
<td>0.40 ± 0.09 d</td>
</tr>
</tbody>
</table>

*Values indicate means ± SE. Different lower case letters indicate significant differences (DMRT) in FW and DW assessed separately for each plant species across treatments (P< 0.05). n= 4.
**Responses of Plants Exposed to SiO$_2$ Nanoparticles**

retroflexus shoot FW and T. officinale shoot DW.

**Effects on the Content of Photosynthetic Pigments**

The results related to photosynthetic pigments for all plants are shown in Table 3. In both crops and in both medicinal plants, there was a significant increase in all photosynthetic pigments (chl a, chl b, total chl, carotenoids) when 400 mg L$^{-1}$ SiO$_2$ NPs were used relative to the control and 2,000 and 4,000 mg L$^{-1}$ SiO$_2$ NPs. For the weeds, the trend was somewhat different: in all cases, either the control or 400 mg L$^{-1}$ SiO$_2$ NPs significantly increased the content of all photosynthetic pigments (or 2,000 and 4,000 mg L$^{-1}$ SiO$_2$ NPs significantly decreased the photosynthetic pigment content).

**Effects on Total Carbohydrate, Total Protein, Free Proline and Total Amino Acid Content of Leaves**

Total carbohydrates, total proteins, free proline and total amino acid content of leaves for all six plants are reported in Figures 3a-d, respectively. For all six plants, any concentration of SiO$_2$ NP significantly decreased the total carbohydrate content of leaves (Figure 3-a). However, for all six plants, 400 mg L$^{-1}$ of SiO$_2$ NPs significantly increased the total protein content of leaves relative to the control while higher concentrations of SiO$_2$ NPs (2,000 and 4,000 mg L$^{-1}$) caused a significant decrease (Figure 3-b). Free proline content was significantly enhanced in the presence of 4,000 mg L$^{-1}$ SiO$_2$ NPs relative to the control and other concentrations of SiO$_2$ NPs (Figure 3-c). The total amino acid content for all six plants followed the same trend as for total protein content, with 400 mg L$^{-1}$ of SiO$_2$ NPs significantly increasing the amino acid content of leaves relative to the control while higher concentrations of SiO$_2$ NPs (2,000 and 4,000 mg L$^{-1}$) caused a significant decrease in total amino acid content (Figure 3-d).

**DISCUSSION**

Plants need 16 essential elements for growth. Although Silicon (Si) is not within this group, it is a very important component of plants, and is treated as an inorganic component (Chen et al., 2000). Si is the second most abundant element in the earth’s surface, makes up more than 41% of the earth’s crust (Exley, 1998), and is absorbed by plants as Silicic acid [Si(OH)$_4$]. Si promoted the number of panicles, number of spikelets/panicle, tillers, grain yield, grain filling and quality of Oryza sativa (Savant, 1997) stem strength in turfgrasses (Hull, 2004), the FW and DW of both roots and shoots of Z. mays and Cucurbita moschata (Liang et al., 2007; Savvas et al., 2009) and the number of fruits in zucchini squash (Cucurbita pepo L. cv. 'Rival'), therefore, increasing agricultural productivity (Savvas et al., 2009). Thus, the advantageous effects of Si lie in its use by plants to enhance growth by reducing mineral toxicity, increasing resistance to biotic stresses, improving nutrient uptake, inducing a balance in plants and enhancing photosynthetic activity (Hull, 2004; Liang et al., 2007). Si affected the chl content in bread wheat (Triticum aestivum L.) (Gong et al., 2005) and total sugars, raffinose, sucrose and soluble sugars of the leaves of sugarcane (Saccharum officinarum L.) (Matichenkov and Calvert, 2002). It was proposed that the principal role of Si in increasing leaf chl arises from its upkeep of the chloroplast ultrastructure accompanied by an improvement of chl biosynthetic enzymes or a reduction of chl-degrading enzymes (Liang et al., 2007; Savvas et al., 2009).

Several studies also suggested that Si can reduce the content of protein carbonyl (oxidative proteins) and increase total soluble proteins in bread wheat (Gong et al., 2005; Gong et al., 2008). In addition, Si acts on mechanisms common to all plants by expressing plant signaling cascades (stress genes) as a natural defense reaction that translates into and activates a strategic signaling protein known as Mitogen-Activated Protein kinases (MAP-kinases) and Proline-Rich (PR) protein (Gong et al., 2008). PR proteins cause phosphorylation of the hydroxyl group on amino acid residues by transmitting information to the nucleus (Fauteux et al., 2005). Si binds to hydroxyl groups and may influence protein conformation or activity,
Table 3. Effect of SiO₂ NPs on photosynthetic pigments (chl a, b, a+b, carotenoids) (in mg g⁻¹ fresh leaf) of six test species.

<table>
<thead>
<tr>
<th>SiO₂ (mg L⁻¹)</th>
<th>Zea mays</th>
<th></th>
<th></th>
<th></th>
<th>Phaselus vulgaris</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>chl a</td>
<td>chl b</td>
<td>Total chl</td>
<td>Carotenoids</td>
<td>chl a</td>
<td>chl b</td>
<td>Total chl</td>
<td>Carotenoids</td>
</tr>
<tr>
<td>Control (0)</td>
<td>0.44 ± 0.01 c</td>
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<td>0.80 ± 0.02 b</td>
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<td>0.25 ± 0.01 b</td>
<td>0.20 ± 0.01 bc</td>
<td>0.45 ± 0.03 b</td>
<td>0.16 ± 0.00 b</td>
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<tr>
<td>400</td>
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<td>1.17 ± 0.00 a</td>
<td>0.62 ± 0.00 a</td>
<td>0.56 ± 0.00 a</td>
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<td>0.28 ± 0.01 b</td>
<td>0.22 ± 0.01 b</td>
<td>0.50 ± 0.02 b</td>
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<td>0.24 ± 0.01 d</td>
<td>0.23 ± 0.01 c</td>
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<td>0.26 ± 0.00 c</td>
<td>0.19 ± 0.00 c</td>
<td>0.15 ± 0.01 c</td>
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<td>0.20 ± 0.00 b</td>
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<table>
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<tr>
<th>SiO₂ (mg L⁻¹)</th>
<th>Hyoscyamus niger</th>
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<th></th>
<th>Nigella sativa</th>
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<tbody>
<tr>
<td></td>
<td>chl a</td>
<td>chl b</td>
<td>Total chl</td>
<td>Carotenoids</td>
<td>chl a</td>
<td>chl b</td>
<td>Total chl</td>
<td>Carotenoids</td>
</tr>
<tr>
<td>Control (0)</td>
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<td>0.28 ± 0.01 b</td>
<td>0.62 ± 0.01 b</td>
<td>0.21 ± 0.00 c</td>
<td>0.38 ± 0.01 b</td>
<td>0.32 ± 0.01 b</td>
<td>0.70 ± 0.02 b</td>
<td>0.17 ± 0.01 c</td>
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<tr>
<td>400</td>
<td>0.62 ± 0.01 a</td>
<td>0.36 ± 0.01 a</td>
<td>0.98 ± 0.02 a</td>
<td>0.42 ± 0.01 a</td>
<td>0.58 ± 0.01 a</td>
<td>0.38 ± 0.00 a</td>
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<td>0.52 ± 0.01 a</td>
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<tr>
<td>2000</td>
<td>0.38 ± 0.01 b</td>
<td>0.24 ± 0.01 b</td>
<td>0.62 ± 0.01 b</td>
<td>0.32 ± 0.00 b</td>
<td>0.32 ± 0.00 c</td>
<td>0.19 ± 0.01 c</td>
<td>0.51 ± 0.00 c</td>
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<td>4000</td>
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<td>0.17 ± 0.01 c</td>
<td>0.36 ± 0.01 c</td>
<td>0.18 ± 0.01 c</td>
<td>0.18 ± 0.00 d</td>
<td>0.12 ± 0.01 c</td>
<td>0.30 ± 0.02 d</td>
<td>0.14 ± 0.01 c</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>SiO₂ (mg L⁻¹)</th>
<th>Amaranthus retroflexus</th>
<th></th>
<th></th>
<th></th>
<th>Taraxacum officinale</th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>chl a</td>
<td>chl b</td>
<td>Total chl</td>
<td>Carotenoids</td>
<td>chl a</td>
<td>chl b</td>
<td>Total chl</td>
<td>Carotenoids</td>
</tr>
<tr>
<td>Control (0)</td>
<td>0.22 ± 0.00 a</td>
<td>0.24 ± 0.01 a</td>
<td>0.46 ± 0.01 a</td>
<td>0.18 ± 0.00 a</td>
<td>0.25 ± 0.00 a</td>
<td>0.21 ± 0.01 a</td>
<td>0.46 ± 0.01 a</td>
<td>0.16 ± 0.01 a</td>
</tr>
<tr>
<td>400</td>
<td>0.21 ± 0.00 a</td>
<td>0.16 ± 0.00 a</td>
<td>0.37 ± 0.01 b</td>
<td>0.16 ± 0.00 a</td>
<td>0.19 ± 0.00 b</td>
<td>0.14 ± 0.01 b</td>
<td>0.33 ± 0.00 b</td>
<td>0.12 ± 0.01 a</td>
</tr>
<tr>
<td>2000</td>
<td>0.11 ± 0.00 b</td>
<td>0.10 ± 0.00 a</td>
<td>0.21 ± 0.01 a</td>
<td>0.11 ± 0.00 b</td>
<td>0.10 ± 0.00 c</td>
<td>0.09 ± 0.00 c</td>
<td>0.19 ± 0.01 c</td>
<td>0.07 ± 0.01 b</td>
</tr>
<tr>
<td>4000</td>
<td>0.04 ± 0.00 c</td>
<td>0.05 ± 0.00 a</td>
<td>0.09 ± 0.01 d</td>
<td>0.06 ± 0.00 d</td>
<td>0.06 ± 0.00 d</td>
<td>0.03 ± 0.00 d</td>
<td>0.09 ± 0.00 d</td>
<td>0.04 ± 0.01 b</td>
</tr>
</tbody>
</table>

Values indicate means±SE. Different lower case letters indicate significant differences (DMRT) in photosynthetic pigments (assessed separately) for each plant species across treatments (P≤ 0.05), n= 4.
Figure 3. Effect of nano-SiO$_2$ nanoparticles (400, 2,000, and 4,000 mg L$^{-1}$) and the control (distilled water without nano-SiO$_2$) on leaf carbohydrate content (a), on total leaf protein content (b), on leaf proline (amino acid) content (c), and leaf total amino acid content (d) in six test species. Different lower case and capital letters show significant differences (DMRT) between treatments means for any one plant species at $P \leq 0.05$. $n = 4$. 
thus Si acts as a moderator and a potentiator of plant defense reactions against biotic and abiotic stresses (Fauteux et al., 2005).

Studies on the effects of Si NPs on plants are limited. In fact, most studies on the benefits and risks of NPs such as TiO$_2$ (Zheng et al., 2005), fullerene (Wang et al., 1999), Al$_2$O$_3$ (Yang and Watts, 2005), or zinc oxide (ZnO) (Lin and Xing, 2007) have focused on higher plants or lower plants such as fungi and algae. A common objective of all these studies was to control the uptake and subsequent effects, depending on the size of the NPs (Limbach et al., 2005; Chithrani et al., 2006). There are no studies on the effect of Si NPs on a wide range of plants.

The results of our study showed that 400 mg L$^{-1}$ SiO$_2$ NP improved seed germination in crops and medicinal plants while higher concentrations of SiO$_2$ NPs decreased seed germination, indicating its toxic effects (Figure 2). Due to the opportunistic nature of weeds, the testa may rupture more easily than that of crops and medicinal plants, thus exposing the endosperm to rupture more easily than that of crops and medicinal plants, while higher concentrations were toxic, decreasing these plant parameters; however, in weeds, all concentrations of SiO$_2$ NPs were phytotoxic (Table 1).

Mahmoodzadeh et al. (2013) reported that TiO$_2$ NPs affected root and shoot FW in wheat: root and shoot DW in the presence of 10 and 100 mg L$^{-1}$ increased these parameters but higher concentrations (1,000, 1,200, 1,500, 1,700, 2,000 mg L$^{-1}$) decreased them. This study also indicated some stimulatory action of SiO$_2$ NPs on root and shoot FW and DW, at least in crops and medicinal plants, with higher concentrations being toxic (Table 2).

Zamani and Moradshahi (2013) treated red algae (Rhodophyta) with 0, 25, 50, 100 and 200 µM of Ag NPs (i.e., nano-Ag). High concentrations of Ag NPs (100 and 200 µM) decreased growth, total carotenoids, and total chl content after 10 days of exposure. Wei et al. (2010) treated turpin (Scenedesmus obliquus L.) with 0, 25, 50, 100 and 200 µM of silica NPs. When treated with 25 µM of silica NPs, photosynthetic pigments (chl $a/b$ and carotenoids) increased as NP concentration increased, but decreased after 96-hours exposure. Jiang et al. (2012) reported that Ag NPs and AgNO$_3$ significantly reduced plant chl $a/b$ content. Unlike all these studies, our study shows that 400 mg L$^{-1}$ SiO$_2$ NPs significantly improved the content of all photosynthetic pigments for all six test species (Table 3), confirming the findings for other studies in which low and moderate levels of NPs increased photosynthetic pigment while high concentrations had a negative impact.

Nitrogen and phosphorus are essential elements for plants as they are related to cell division and growth, and treatment with metals decreased their uptake (Batty and Younger, 2003). These elements can affect the synthesis of photosynthetic pigments (Batty and Younger, 2003), so a decrease in chl could be related not only to increased degradation but also to decreased synthesis. Reduced content of
Responses of Plants Exposed to SiO₂ Nanoparticles

photosynthetic activity may be related to a decrease in access to light due to the accumulation of SiO₂ NPs on the surface of plant cell walls, causing a shading effect (Wei et al., 2010).

Salama (2012) reported that 60 mg L⁻¹ of Ag NPs increased carbohydrate content in P. vulgaris and Z. mays (57 and 62% more than the control) and that at 80 and 100 mg L⁻¹ of Ag NPs, carbohydrate concentration was reduced (19 and 18% for common bean and 28 and 31% for maize relative to the control). Our results show that in field crops, medicinal and weed plants, carbohydrate concentration decreased significantly as SiO₂ NP concentration increased, suggesting the toxic nature of SiO₂ NPs over a wide range of plants. Photosynthesis is a plant’s main metabolic pathway in which sugars are synthesized from CO₂, water, and light energy. These sugars or carbohydrates serve as the origin of energy for a plant’s other metabolic procedures. Thus, a low level of photosynthetic activity caused by SiO₂ NP stress can decrease carbohydrates directly, causing plant growth to be reduced.

Our results showed that with increasing SiO₂ NP concentration, there was a decrease in total protein content, with maximum values at 400 mg L⁻¹ SiO₂ NPs (Figure 3-b). At 60 mg L⁻¹, Ag NPs increased the protein content of P. vulgaris and Z. mays leaves (30 and 24%, respectively, more than the control) while 100 mg L⁻¹ significantly decreased the protein content (32 and 18%, respectively, less than the control) (Salama, 2012).

As SiO₂ NP concentration increased, proline content increased for all six crops (Figure 3-c). Proline accumulates in plant under a wide range of stress conditions such as high and low temperature, pathogen infection, anaerobiosis, nutrient deficiency, heavy metal toxicity, UV-irradiation, atmospheric pollution, salinity, water deficiency, high light intensity and extreme temperatures (Mansour, 2000). Proline accumulation in plant tissues has been proposed to result from a reduction in proline degradation, an intensification of proline biosynthesis, or a reduction in proline utilization or in the synthesis and hydrolysis of proteins. It protects plants under stress by stabilizing cell membranes by interacting with phospholipids, and by protecting folded protein structures against denaturation, functioning as a free radical scavenger, or serving as a source of energy and nitrogen.

As SiO₂ NP concentration increased, total amino acid content decreased (Figure 3-d). Amino acids are the precursors or activators of phytohormones and growth substances. Glycine and glutamic acid are fundamental metabolites in the formation of plant tissue and chl synthesis. These amino acids raise the chl content in plants which increases the absorption of light energy and leads to increased photosynthesis. As a result, if the amino acids content decreases, photosynthetic pigments, total protein, total carbohydrate content, and other related metabolic problems in plants can also be reduced.

CONCLUSIONS

The use of SiO₂ NPs at high concentrations can result in toxic effects on morphological, physiological, and biochemical characteristics of crop, medicinal and weed plants while, in select cases and for select parameters, a lower concentration (400 mg L⁻¹) can in fact be beneficial. Thus, caution is urged in the use and disposal of such materials into the environment. Regarding the toxic levels, future studies should focus on levels of uptake and retention, source-sink relations and the localization of SiO₂ NP sinks, the mechanism of phytotoxicity and uptake kinetics, and interactions within cells. However, since lower concentrations present positive aspects, concentrations of SiO₂ NPs should be optimized for each crop (a narrower range of 10-1,000 mg L⁻¹ should be tested next) in a bid to maximize yield and other ergonomically favorable factors. Ideally, toxicity tests should be conducted hand in hand with optimization trials.

REFERENCES


چکیده
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