Total Oxalate-Soluble Pectin Concentration in Asian Pear (Pyrus serotina Rehd) Fruit in Relation to Ripening, Storage and Internal Browning Disorder

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ABSTRACT

Oxalate Soluble Pectin (OSP) is related to fruit firmness with respect to the chelation of calcium ions with carboxyl groups of adjacent polyuronide chains. This study was carried out to explore the effect of time of fruit harvest, foliar spray with CaCl2 during growing season on OSP, Polygalacturonase Activity (PGA), fruit firmness and the effects on fruit shelf life, quality and Internal Browning (IB) disorder. Fruit samples were collected from Asian pear (Pyrus serotina Rehd.) trees ‘KS’9 and ‘KS’13 on European pear (Pyrus communis L.) seedling rootstocks. Fruits were picked on the 1st and 15th August, 2006 and stored at 1°C and 80-85% relative humidity (RH). Five stages of extraction were performed on fruit samples including one pre-harvest, one at the time of harvest, and three following fruit harvest. Results indicated that OSP increased slightly during fruit ripening as well as during storage. In both the studied cultivars, the level of OSP in fruit harvested fifteen days after Optimum Time for Harvest (OTH) was higher than that harvested at OTH. The level of OSP in fruit softened after harvest significantly increased starting from the time of harvest. The relationship between flesh firmness, IB and OSP concentration showed the highest correlations in the both of the studied cultivars among PGA. In conclusion, flesh firmness and IB were correlated with the extent of OSP concentration. In addition, fruit, sprayed with CaCl2 and harvested early, contained less OSP, PGA and IB following long-term storage.

Keywords: Firmness, Polygalacturonase activity, Softening, Soluble solid concentration, Storage, Titratable acids.

INTRODUCTION

One of the major disorders that develop during storage in pear, including Asian pears is internal browning (IB) of the flesh around core regions. It has been reported that mature fruits are more susceptible to senescence IB than immature ones. A determination of the optimum harvest time is an important task for preservation of the quality of Asian pear during the storage period. European and Asian pears that are picked either before or after the optimum harvest dates develop an inferior taste and are more sensitive to IB disorder, never developing the characteristics such as juicy texture of ripe fruit (Crisosto et al., 1994; Murayama et al., 1998; Arzani et al., 2008; 2009).

Pectin is one of the major components of the primary cell wall and of middle lamella in plant tissues. Pectic matrix provides an

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environment for the deposition, extension of the cellulose-glycan network and is the major adhesive material among cells (Willats et al., 2001). Pectin degradation plays an important role in fruit ripening, which leads to destruction of cellulose and hemicellulose and is accompanied by a decrease in fruit firmness (Duan et al., 2008). It has been reported that secretions of pectin-degrading enzymes, for example, endo- and exo-polygalacturonase accelerate scab and IB disorder in Asian pear (Faize et al., 2003). It seems that Asian and European pear fruit softening is related to changes in cell wall structure. These changes are related to an increase in soluble polyuronides accompanied by a decrease in insoluble polyuronides (Murayama et al., 2002). It has been reported that chelate-soluble pectin and sodium carbonate-soluble pectin are both closely related to fruit texture (Yang et al., 2006). Although, chelate-soluble pectin is much connected with the textural difference among the different fruit stages of the same cultivar, sodium carbonate-soluble pectin is more related to the textural difference among different fruit cultivars (Yang et al., 2009).

Quality Asian pears develop a buttery and juicy texture, associated with a reduction in extractable juice. In addition, the chelation of calcium ions with carboxyl groups of adjacent polyuronide chains is a key factor of fruit firmness with calcium deposition occurring in the junction zones in the middle lamella (Knee, 1993). Any increase in free pectin carboxyl groups might be expected to enhance the importance of calcium as a firmness-increasing agent (Villalobos and Mitcham, 2008).

Such calcium-binding agents as solutions of oxalate, Ethylene Diaminetetra Acetic Acid (EDTA) or polyphosphate have been employed to extract pectic acid from fruits (Pilnik and Voragen, 1971). For example, solutions of oxalic acid and ammonium oxalate are used to extract pectic acid in the form of oxalate-soluble pectin. Oxalate-soluble pectin exists as pectic acid that can bind calcium and form a cross-link structure (Yu et al., 1996; Aileen et al., 2008; Yadav et al., 2009). Wide variations in extraction times (10 min to 24 hours) and temperatures (0°C to boiling point) have been employed (Pilnik and Voragen, 1971). Oxalate-soluble pectin has been extracted from plant tissue (Aileen et al., 2008), and residue obtained from water-soluble pectin extracts of dry cell wall (Bouranis and Niavis, 1992), and alcohol-insoluble solids of fruits and vegetables (Ketsa et al., 1999; Panmanas et al., 2001; Yadav et al., 2009) under different conditions. These have caused the difference or misinterpretation of the data such as different correlation between the pectin concentration and firmness of the fruit, the contrasting comparison of solubilization of different kinds of pectin. Moreover, the time required to completely extract pectin was considerably long and might be unattainable.

The objective of the present research was to evaluate the effect of storage durations, CaCl₂ application and duration of postharvest on OSP concentration, pectin-degrading enzyme (polygalacturonase), internal browning (IB), firmness and the other related quality attributes of ‘KS’9 and ‘KS’13 Asian pears.

**MATERIALS AND METHODS**

**Samples**

Fruit samples were collected from Asian pear (*Pyrus serotina* Rehd.) trees ‘KS’9 and ‘KS’13 on European pear (*Pyrus communis* L.) seedling rootstocks, at the Research Orchard, Department of Horticultural Science, Tarbiat Modares University (TMU), Tehran, Iran. Preharvest foliar spray with calcium chloride (CaCl₂) at the concentration of 5 g L⁻¹ was applied by 2 week intervals from 30 days after full bloom (DAFB) until the time of fruit harvest. Asian pear fruits were picked on the 1st and 15th August, 2006 and stored at 1°C plus 80-85% relative humidity (RH). Samples were taken from 3 fruits and combined to form the substance for each replication.
Oxalate-Soluble Pectin (OSP) Extraction

Oxalate-soluble pectin (OSP) extraction was performed according to the method described by Panmanas et al. (2001). OSP was extracted from the alcohol-insoluble solids (AIS). The residue was Water Soluble Pectin (WSP) extract, so it was stirred in a solution of 0.25% ammonium oxalate and 0.25% oxalic acid for 1 minute. The OSP was extracted using a hot water bath at 95±2°C. The extraction times employed consisted of: 1 to 15 hours. (Figure 1). Three fruits (at each picking date) were used for each sample extract. OSP concentration was assessed and expressed as pectin concentration in AIS (µg mg⁻¹ AIS).

Internal Browning (IB) Evaluation in the Fruit

Internal browning (IB) in the fruit, including flesh and heart browning was assessed by counting the number of damaged fruits and expressed as percentage.

In addition, in order to determine the intensity of IB incidence according to the percentage of brownish area, affected fruit was cut into four equal longitudinal parts and scored as incidence intensity using the scale of 0 to 5. The nominated scores were: (0) without visible IB damage; (1) slightly visible IB damage with 1 to 10% of the area showing browning; (2) moderately visible IB damage with 11 to 33% of the area showing browning; (3) severe injury with 34 to 66% browning and (5) extreme injury with 67 to 100% browning (Ju et al., 2000).

Determination of Polygalacturonase (PG) Activity

Polygalacturonase (PG) activity was assessed through 2-cyanoacetamide spectrophotometric method (Gross, 1982). Fruit tissue (150 g) was placed in 300 ml of distilled water and homogenized in a Waring blender for 1 min. The homogenate was filtered through 6 layer cheesecloth and the residue resuspended in 300 ml of distilled water. The suspension was filtered and the residue resuspended in 150 ml of 1M NaCl. The acidity (pH) was adjusted to and maintained at 6.0 with 1N NaOH while the slurry slowly stirred at 4°C. After 3 hours the slurry was filtered through cheesecloth and the filtrate centrifuged at 9000xg for 15 minutes. Supernatant (1.5 ml) was de-salted on a sephadex G-25 column (10 ml bed volume) which was equilibrated in 50 mM Na-acetate (pH 4.4). The de-salted extract was used for polygalacturonase assay. The extraction procedures were carried out at 4°C. The assay of polygalacturonase activity was based on the hydrolytic release of reducing groups of galacturonic from polygalacturonic acid. Reaction mixture (0.2 ml of total volume) containing 37.5 mM Na-acetate (pH 4.4), 0.2% polygalacturonic acid (washed with 80% ethanol prior to use), and 50 µl of enzyme extract were incubated at 37°C for up to 3 hours. For quantifying the released reducing groups with 2-cyanoacetamide, reactions terminated with 1 ml of cold 100 mM of borate buffer (pH 9). Then 0.2 ml of 1% 2-cyanoacetamide were added, and the samples were mixed and immersed in boiling water bath for 10 minutes. After equilibration to 25°C the absorbance at 274 nm was determined using UNICAM 8620 UV-Vis spectrophotometer. A unit of PG activity is defined as the level of enzyme that can release 1 nmol (galacturonic acid) of reducing groups h⁻¹ at pH 4.4 and at 37°C.

Fruit Firmness, Soluble Solids Concentration (SSC) and Titratable Acids (TA) Measurements

Fruit firmness, SSC, and TA were assessed both at the time of fruit harvest and after cold storage (at weekly intervals). Firmness was evaluated through a penetrometer (CNS FARNELL) equipped with a 12 mm plunger at two locations per...
Figure 1. Extraction curve of oxalate-soluble pectin concentration of Asian pear (‘KS’13) extracted through hot water (95±2°C). Data are the means (3 fruits) with the error bar indicating the standard error.

Figure 2. Extraction rate of oxalate-soluble pectin of Asian pear (‘KS’13) extracted through hot water (95±2°C).

The changes in OSP from the fruit harvested at different stages are shown in Figure 2. The trend of extraction rates, changed with extraction time is shown in Figure 3. In the case of Asian pear, the power equation pattern best fitted the time(h) differences were considered statistically significant at \( P \leq 0.05 \). The REG procedure (SAS Institute) was used for multiple regression and correlation analysis.

RESULTS AND DISCUSSION

Changes of OSP and Fruit Firmness

The results were statistically evaluated through analysis of variance (ANOVA) and expressed as mean±standard error (SE). Mean comparisons were made using Duncan’s multiple range test (DMRT);
Table 1. Extracted total oxalate-soluble pectin concentration at different extraction times (preharvest, harvest and postharvest) of two Asian pear ‘KS’9 and ‘KS’13 fruits.

<table>
<thead>
<tr>
<th>Stage of storage</th>
<th>Total oxalate-soluble pectin (µg mg⁻¹ AIS)</th>
<th>‘KS’9</th>
<th>‘KS’13</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 (1 month before HT)</td>
<td>1 August</td>
<td>15 August</td>
<td>1 August</td>
</tr>
<tr>
<td>Control (without IB incidence)</td>
<td>96.32d*</td>
<td>105.15e</td>
<td>87.52d</td>
</tr>
<tr>
<td>CaCl₂ foliar spray</td>
<td>115.62c</td>
<td>123.26c</td>
<td>106.56c</td>
</tr>
<tr>
<td>Control (11–33% IB)</td>
<td>123.26c</td>
<td>139.50</td>
<td>142.50</td>
</tr>
<tr>
<td>Control (34–66% IB)</td>
<td>139.74</td>
<td>139.50</td>
<td>140.27</td>
</tr>
<tr>
<td>2 (HT)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3 (1 month after HT)</td>
<td>135.36a</td>
<td>142.30a</td>
<td>132.94a</td>
</tr>
<tr>
<td>4 (2 months after HT)</td>
<td>132.94a</td>
<td>139.74</td>
<td>142.50</td>
</tr>
<tr>
<td>5 (3 months after HT)</td>
<td>119.20c</td>
<td>113.70d</td>
<td>122.30b</td>
</tr>
</tbody>
</table>

a Harvest Time; b Internal Browning.
* Mean separation with columns by Duncan’s multiple range tests at 1% level.

Figure 3. Polygalacturonase activity in Asian pear cv. ‘KS’9 and ‘KS’13 during preharvest, at harvest and during postharvest: -1 (1 month before harvest) and 0 (harvest time, 1st Aug. 2006).

Extraction rates curve. The power equation is defined by \( y = at^b \), where \( y \) is the extraction rate, \( t \) the extraction time, with \( a \) and \( b \) as constants. Though, the OSP concentration extracted increased considerably with extraction time and picking dates. The OSP extraction rate was highly affected by extraction time, although the picking dates did not affect the extraction rates. These data indicate that the AIS of the fruit at any developmental stage could be used as a determination of half-extraction time. Panmanas et al. (2001) reported that to obtain the total OSP concentration, the extraction could be done until half-extraction time was reached. To get the total concentration, the OSP concentration at half-extraction time needs to be simply multiplied by two. According to the present data, the half-extraction time of Asian pear was 2.5 hours while total OSP concentrations were 115.62 (‘KS’9) and 106.56 µg mg⁻¹ AIS (‘KS’13) for the fruits harvested on 1st August 2006 and 123.26 (‘KS’9) and 115.42 µg mg⁻¹ AIS (‘KS’13) for the fruits harvested on 15th August 2006 respectively (Table 1). In both cultivars, late-harvested fruits on (15th August) contained more OSP than the early-harvested (1st August) fruit. In fruits picked...
from trees that were sprayed with CaCl₂, OSP was lower than those picked from non-sprayed trees in both of the studied cultivars, so such fruits exhibited higher firmness during the storage period. In addition, it was found that the low damaged fruits (11–33% IB) had a more OSP content than the sound fruits and than fruits with a high intensity of IB incidence (more than 33% IB) (Table 1).

Fruits in both cultivars softened during storage. A relatively high level of firmness was observed in ‘KS’₁₃ cultivar after a long-term storage. Also, ‘KS’₉ fruit softened faster than fruits of ‘KS’₁₃ cultivar fruit (Table 3). Fruits all softened during ripening, independent of cultivar, harvest time and storage period. The rate of softening was dependent on harvest date as well as on cultivar. In both of the studied cultivars, the later harvested fruits (15th August) suffered from a lower firmness than the early-harvested (1st August) ones and tended to soften faster. There was a relationship observed between flesh firmness and the extent of OSP in fruits before and after storage in both the studied cultivars. It has been reported that in pears, the level of OSP generally increases during normal ripening (Yoshioka et al., 1992; Murayama et al., 1998). Ahmed and Labavitch (1980) reported that galacturonic acid in the 80% ethanol perceptible portion of the supernatant fraction of pear fruit homogenate actually increased during fruit softening. Kim et al. (1991) also reported that a steady decline in cell wall galactosyl residues was accompanied by an increase in soluble galactose during tomato fruit ripening. In any case, OSP concentration increased when fruits were transferred to 20°C following a short-term storage. After long-term storage, fruits (in both cultivars) also softened during ripening. It has been reported that (Duan et al., 2008) fruit firmness of banana decreased rapidly from

Table 2. Polygalacturonase activity (PGA) in Asian pear cv. ‘KS’₉ and ‘KS’₁₃ in preharvest, harvest and postharvest duration and the effect of CaCl₂ foliar spray and internal browning on PGA.

<table>
<thead>
<tr>
<th>Stage of storage</th>
<th>PGA in ‘KS’₉ (nmol g⁻¹ h⁻¹)</th>
<th>PGA in ‘KS’₁₃ (nmol g⁻¹ h⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Harvest Time</td>
<td>Harvest Time</td>
</tr>
<tr>
<td></td>
<td>1 August 15 August</td>
<td>1 August 15 August</td>
</tr>
<tr>
<td>1.0 (1 month before HT)</td>
<td>16±1.0e*</td>
<td>21±1.4e</td>
</tr>
<tr>
<td>Control (without IB incidence)</td>
<td>19±0.8d</td>
<td>24±1.6d</td>
</tr>
<tr>
<td>0 (HT) CaCl₂ foliar spray</td>
<td>17</td>
<td>21</td>
</tr>
<tr>
<td>Control (11–33% IB)</td>
<td>26</td>
<td>25</td>
</tr>
<tr>
<td>Control (34–66% IB)</td>
<td>24</td>
<td>23</td>
</tr>
<tr>
<td>1.1 (1 month after HT)</td>
<td>26±1.7c</td>
<td>33±1.9c</td>
</tr>
<tr>
<td>2 (2 months after HT)</td>
<td>42±2.7a</td>
<td>39±1.9a</td>
</tr>
<tr>
<td>3 (3 months after HT)</td>
<td>33±2.1b</td>
<td>35±1.5b</td>
</tr>
<tr>
<td>4 (4 months after HT)</td>
<td>25±1.2c</td>
<td>24±1.1d</td>
</tr>
</tbody>
</table>

*Harvest Time; †Internal Browning.
*Mean separation with columns by Duncan’s multiple range tests at 1% level.
Table 3. Changes of fruit physicochemical properties of two Asian (`KS`9 and `KS`13) pear cultivars during five months of storage.

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Properties</th>
<th>Harvest time</th>
<th>Time (Month of storage)</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>0a</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Firmness</td>
<td>(N)</td>
<td>1Aug</td>
<td>44±1.2a</td>
<td>40±1.8ab</td>
</tr>
<tr>
<td></td>
<td></td>
<td>15Aug</td>
<td>37±0.5a</td>
<td>35±2.1a</td>
</tr>
<tr>
<td><code>KS</code>9</td>
<td>SSC (°Brix)</td>
<td>1Aug</td>
<td>13.2±0.5a</td>
<td>14.4±0.4a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>15Aug</td>
<td>14.1±0.3ab</td>
<td>15.2±0.2a</td>
</tr>
<tr>
<td></td>
<td>TA (mg/100g FW)</td>
<td>1Aug</td>
<td>33±0.6a</td>
<td>24±0.7b</td>
</tr>
<tr>
<td></td>
<td></td>
<td>15Aug</td>
<td>25±0.3a</td>
<td>18±0.5b</td>
</tr>
<tr>
<td>Firmness</td>
<td>(N)</td>
<td>1Aug</td>
<td>46±2.3a</td>
<td>42±1.7ab</td>
</tr>
<tr>
<td></td>
<td></td>
<td>15Aug</td>
<td>34±1.2a</td>
<td>28±0.9ab</td>
</tr>
<tr>
<td><code>KS</code>13</td>
<td>SSC (°Brix)</td>
<td>1Aug</td>
<td>14.5±0.5a</td>
<td>15.8±0.5a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>15Aug</td>
<td>15.1±0.3ab</td>
<td>17.2±0.5a</td>
</tr>
<tr>
<td></td>
<td>TA (mg/100g FW)</td>
<td>1Aug</td>
<td>28±0.7a</td>
<td>24±0.7b</td>
</tr>
<tr>
<td></td>
<td></td>
<td>15Aug</td>
<td>22±0.4a</td>
<td>20±0.6ab</td>
</tr>
</tbody>
</table>

a Time of fruit harvest.
b Mean separation within columns by Duncan’s multiple range tests at $P \leq 0.01$. Mean values±standard errors. NS, * and **; Nonsignificant or significant at $P \leq 0.05$ or 0.01, respectively.
the initial 11.6 to 1.8 N after 20 days of storage at 25°C.

Fruit softening is generally attributed to cellular wall structure, particularly due to solubilisation of pectin (Lohani et al., 2004). As shown in Table 1, OSP content increased significantly from fruit ripening (stages 1 to 3), indicating that solubilisation might occur. Also a number of reports indicate the conversion of water-insoluble to water-soluble pectin (Majumder and Mazumdar, 2002; Manrique and Lajolo, 2004; Yashoda et al., 2005), similar to the present research results during fruit ripening.

Wang et al. (1985) reported that ‘Eldorado’ pears usually remained firm and dry when transferred to 20°C after 36 weeks of storage at 0°C. Chen and Borgic (1985), reported that pear fruits did not ripen normally, never developing the buttery and juicy texture of ripe fruit with the OSP concentration of the fruit not changing, during 9 days of ripening after fruits were transferred to 20°C environment after prolonged storage at −1 or 0°C. Nevertheless, changes in OSP and softening during ripening and storage in the present research are in agreement with results from other works on pears (Ben-Arie and Sonego, 1979; Yoshioka et al., 1992).

Changes of Polygalacturonase Activity

Figure 3 and Table 2 show the changes in polygalacturonase (PG) activity during storage in two Asian pear cultivars determined by measuring the release of reducing sugars from polygalacturonic acid. In both the studied cultivars, PG activity increased from 1 month before to 2 months after harvest then decreased slightly in the early (1st August) and the late (15th August) harvested fruits. There was a relationship observed between OSP and polygalacturonase activity (PGA). For OSP concentration, higher OSP correlated with higher PG activity in Asian pears before and after storage (Figure 4). The degradation of pectic polysaccharides well corresponded with softening behaviors in the two Asian pears. Solubilization of pectic polysaccharides was observed in ‘KS’9 and ‘KS’13 Asian pears before 1 to 2 months after harvest (Table 1).

Involvement of polygalacturonase (PG) and/or Pectin Methyl Esterase (PME) in enzymatic disassembly of pectin polysaccharides, during fruit ripening, has been extensively investigated (Nikolic and Mojovic, 2007; Prasanna et al., 2007). Polygalacturonase and PME cooperatively regulate the pectin disassembly. Polygalacturonase hydrolyses α-1, 4-linked D-galacturonic acid, following de-esterification of pectin through PME. In addition, Pectate Lyase (PL) (Payasi et al., 2006) and bgalactosidase (Lazan et al., 2004) may play a combined effect on the disassembly of pectin polysaccharides during fruit ripening/softening. Non-enzymatic action might mediate in the polysaccharide solubilisation in plant tissues (Dumville and Fry, 2003). It has been reported that hydroxyl radical (·OH) was capable of cleaving polysaccharides present in cellular walls in a site-specific reaction, which led to cellular wall loosening and elongation of living coleoptile or hypocotyls in maize and in soybean. Enzymatic and non-enzymatic effects on pectin degradation are associated with modification of wall structure, including the modification in the composition, molecular weight, and structural characteristics of the pectin (Schweikert et al., 2002).

There are some reports on modification of cellular wall polysaccharides in apple (Scalzo, et al., 2005), papaya (Manrique and Lajolo, 2004) and other fruits (Sakurai and Nevins, 1997; Rose et al., 1998). However, variations in cell wall compositions could lead to differences in the softening-associated chemical modification for each fruit species. Huber and O’Donoghue (1993) have demonstrated that depolymerization of pectic polysaccharides in avocado is more extensive than that in tomato fruits, and correlates with the differences in flesh texture between these fruits. Similar
relationships between modes of pectin degradation and particular textural properties have also been revealed in other species (Yoshioka et al., 1992; Redgwell et al., 1997). These findings may suggest that the mode of pectin disassembly is one of the important factors determining fruit firmness and texture. Endo-PG and Exo-PG act on pectin-degrading enzymes for adjacent polyuronide chains that are a key factor for fruit firmness (Knee, 1993). In the present research, as with the rate of softening, the rate of increase in PG activity was greater in late-harvested than in the early-harvested fruits. PG activity determined through reducing sugar assay increased with ripening in the two Asian pears. Endo-PG activity has been identified in a number of ripening fruits and has been shown to correlate with an increase in soluble pectin during ripening (Hadfield and Bennett, 1998).

The present study demonstrates that PG can lead to significant solubilization of pectic polymers leading to an increase in OSP concentration. The diffusion freedom or mobility of wall-associated proteins has also been implicated as a means of regulating the activity of PG being released during the in vitro catalysis (Chun and Huber, 1997), with sustained pectin

**Figure 4.** The relationship between PG (Polygalacturonase) activity, OSP (Oxalate Soluble Pectin), IB (Internal Browning) and firmness of ‘KS’9 and ‘KS’13 Asian pears during storage.
solubilization requiring continued provision of fresh enzyme (Rushing and Huber, 1990), consistent with a relatively immobile status of the enzyme. Also The parallel interruption of PG deposition and pectin solubilization in silver-thiosulfate-treated tomato fruit reported Smith et al., 1989.

The biochemical bases for ripening and senescence associated with fruit softening are not yet fully understood. However, dissolution of the middle lamella and cell wall separation due to depolymerization and solubilization of pectin by PG as well as a loss of Ca$^{2+}$ have been suggested to contribute to fruit softening (Fischer and Bennett, 1991). The role that calcium plays in increasing cell rigidity has been related to its ability to bind with mid lamella pectate (Moraga et al., 2009). The firmness of the flesh of fresh-cut fruit products may be improved if treated with calcium compounds. Spraying of fruits with solutions of calcium chloride is very effective in maintenance of product firmness (Moraga et al., 2009). Calcium lactate has recently been shown to be as effective as the chloride form. On the other hand, a decrease in the respiration rate has been observed in samples treated with calcium. This may be related to the increase in the membrane rigidity which blocks the gas interchange to a delay in the arrival of the senescence or to the level of active water transport inhibition (Moraga et al., 2009). The results showed that in fruits sprayed with CaCl$_2$, PG activity was lower than in non sprayed trees in both cultivars, so such fruits benefited from a lower OSP and a higher firmness during storage period.

Conceivably the loss of bound Ca$^{2+}$ and the ability to form Ca$^{2+}$ cross-bridges might have neutralized any effects of reduced depolymerization of pectin on ripening-associated softening of transgenic fruits. Depending on the concentration of Ca$^{2+}$ ions, pectin has been suggested to form different types of aggregates (Aileen et al., 2008). Under low Ca$^{2+}$ levels, polygalacturonates have been suggested to form primary units of two chains in antiparallel configuration with about 50% of the carboxyl groups neutralized with Ca$^{2+}$. The excess Ca$^{2+}$ is weakly bound to sheet like aggregates formed with several primary units, which add little strength to the polygalacturonate gels (Aileen et al., 2008). This may explain why even after a significant loss of bound Ca$^{2+}$ detectable changes in fruit softening were not observed. Reduced PME and PG exhibit opposite effects on firmness during senescence of other fruits. An increase in fruit integrity has been shown in fruits with low PG activity. It is likely that the continuous action of several cell wall hydrolyses, including PG, on structurally altered cell walls results in the loss of fruit integrity during fruit senescence. An increase in PG activity due to reduced bound Ca$^{2+}$ levels may be partly responsible for the loss of tissue integrity (Fischer and Bennett, 1991).

PG activity in damaged fruits varied with the intensity of the damage. Slightly affected fruits (index 1) showed a significant increase in PG activity. Later, as the damage progressed (index 2 to 4), PG activity decreased (Table 2). In peach, a remarkable increase in PG activity during ripening was detected in some cultivars (Pressey and Avants, 1978). In European pear, activities of both exo and endo-PG were identified and characterized in ripening fruit (Pressey and Avants, 1978), although the relationship between each type of PG activity and softening was not established.

It has been reported that in the high-methoxyl (HM) form of pectin, more than 50% of the carboxyl groups are methylated, while in the low-methoxyl (LM) form less than 50% are methylated. The degree of methylation affects the gelling properties of pectin. High-methoxyl pectin gelation requires an acidic environment, pH< 3.5 and the presence of > 55% co-solute, usually sugar, whereas LM pectin gels over a wide range of pH values, either with or without sugar, and in the presence of Ca$^{2+}$ ions. The network structure of HM pectin depends on hydrophobic interactions and hydrogen bonds. In the present research it was found...
that sprayed fruits with solutions of calcium chloride were very effective in maintaining product firmness, low OSP, a decrease of PG activity and finally an increase of postharvest longevity in the studied Asian pear fruits.

**Changes of SSC, TA and Internal Browning**

Table 3 shows changes of physicochemical properties of the early (1st August) and late (15th August) harvested ‘KS’9 and ‘KS’13 Asian pears including firmness, SSC and TA. Relatively high levels of SSC were observed in ‘KS’13 cultivar (Table1). It is noteworthy that ‘KS’13 showed the higher SSC but contained a lower acid content. Asian pear SSC developed with fruit ripening, increasing slowly from harvest time (14.1 and 15.1° Brix in ‘KS’9 and ‘KS’13 harvest on 15th August) to 1 month after storage (15.2 and 17.2° Brix in ‘KS’9 and ‘KS’13 harvest on 15th August) and then decreased only slightly (Table 3). In fruits harvested on 1st August after 2, 3, 4 and 5 months of storage SSCs were higher in both cultivars. In addition, SSC decreased significantly from 1 month of storage to 5 months after storage in early and late harvested fruits. IB damaged fruits were higher in fruits harvested on 15th August during storage of either cultivar (Figure 5). In, ‘KS’13, fruits harvested on 15 August showed 5, 6, 10, 20 and 45% IB symptoms more than those harvested on 1st August after 1, 2, 3, 4 and 5 months of storage, respectively. IB can be avoided by early harvesting of Asian pear fruits (Figure 5). Crisosto et al. (1994) reported Asian pears grown under California conditions and picked 180 days after full bloom are likely to develop IB during storage. During storage, browning symptoms increased sharply in the more mature fruits (Veltman et al., 2000). In general, the browning tendency of Asian pear fruits increased with advancing maturity (Veltman et al., 2000). Those changes in IB values during storage are in agreement with the present results. In addition, our results indicated that fruit sprayed with CaCl₂ showed a lower IB value than the fruit of non sprayed trees in either cultivar. It has been reported that the chelation of calcium ions with carboxyl groups of adjacent polyuronide chains is a key factor of fruit firmness with calcium deposition occurring in the junction zones in the middle lamella (Knee, 1993). Any increase in free pectin carboxyl groups might be expected to increase the impact of calcium as a firmness-increasing agent (Van Buren, 1979). In the present research, there was a high correlation observed between IB value and the level of OSP in fruits in either

![Figure 5](image-url)  
**Figure 5.** Effect of picking date (1 and 15 August) on Internal Browning (IB) extent in Asian pear ‘KS’13 fruit.
one of the studied cultivars during storage (Figure 4). IB incidence was higher in fruits that contained a higher OSP prior to 2 months of storage.

In this study, relationships between SSC, TA as well as between internal browning and OSP concentration were investigated. Pectin is a naturally abundant polysaccharide, and an essential component in initial cell growth as well as in the ripening process. Holm et al. (2009) reported that sweetness and thickness of strawberry increased with increasing pure low-methoxyl pectin concentration. Usually, pectin compound contains a large level of sugar, sweetness with SSC differences due to varying sugar content. In the present research, the relative increase in SSC during ripening may account for the decrease in galactose and mannose contents and also relative increase in fructose content during ripening and as well a decrease of SSC during storage may account for the decrease of OSP. Similarly, this release of galactose from pectin fraction and/or the decreased pectin level have been reported during storage of tomato, apple, strawberry and peach (Duan et al., 2008).

In the present work, there existed a relationship among SSC, TA and IB. Higher SSC and TA correlated with lower IB in Asian pears before and after storage. Veltman et al. (2003) reported that such fruit disorders as internal browning in pears may develop because Adenosine Triphosphate (ATP) levels drop below levels needed for such cell maintenance activities as membrane repair. Pear cells subjected to anoxia exhibited an ATP synthesis threshold below which membrane lipids hydrolysed. The main source of ATP in fruits comes from sugars and organic acids. Some of such organic acids as Ascorbic and citric play an antioxidant effect on polyphenol oxidase activity (one of the most important factors affecting IB), which act against reactive O2 (Jimenez et al., 1997). In European pears, a relationship was found between AA content, harvest date and the susceptibility to browning during the storage period (Veltman et al., 2003). It has been reported that in ‘Conference’ pear there is a tendency of IB development when AA levels drop below a certain level.

In conclusion, the results indicated that the level of fruit OSP (softened after harvest) significantly increased with time elapsed from harvest. In fruits sprayed with CaCl2 and harvested earlier (1st August), OSP was lower than in those of non sprayed trees and harvested late (15th August) in both of the studied cultivars. Fruits in both cultivars softened during storage and the fruits harvested later (15th August) suffered from a lower fruit firmness than the early-harvested (1st August) ones. Higher OSP correlated with higher PG activity in Asian pears both prior to, and after storage. Nevertheless, in fruits harvested from sprayed trees with CaCl2, PG activity was lower than that in the fruits harvested from non sprayed trees in both of the studied cultivars. In addition, fruits picked from sprayed trees with CaCl2 and harvested on 1st August showed a lower percentage of IB incidence in comparison with those harvested from non-sprayed trees with CaCl2.

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REFERENCES


Asian Pear Fruit and Internal Browning Disorder


(Pyrus serotina Rehd.)

غلوظت پکتن انگزالات محلول گل در میوه گلابی آسیایی
در ارتباط با رسیدگی، انبارداری و عارضه فوهه ای شدن داخلی میوه

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چکیده
پکتن انگزالات محلول گل با سنتی یافته میوه ارتباط دارد و این همیستگی با توجه به کلاس پون های
کلسیم به گروه کربوکسیل مجاور زنجیره پلی پورونید بوجود می آید. این آزمایش به منظور بررسی اثر
زمان برداشت، محلول پاشی کلرور کلسیم در طول فصل رشد بر پکتن انگزالات محلول 
(PGA)، فعالیت آنزیم بیلی گالاکتروزاز (PGA) و نتیجه آن ها بر انبارداری، کیفیت و عارضه فوهه ای شدن
(Pyrus serotina Rehd.) در داخلی میوه انجام شد. یکین منظور میوه از درختان گلابی آسیایی

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ارقام 9 و 13 'KS' که بر روی پایه گلایه اروپایی (Pyrus communis L.) پوشش شده بودند جمع آوری گردید. میوه‌ها در دو تاریخ 20 مرداد و 5 شهریور 1385 برداشت و در دمای 18 درجه سانتی‌گراد در دسترس نگهداری شدند. بر روی میوه‌های برداشت شده یک مرحله استحراز شامل یک مرحله قبل از برداشت معمول میوه و یک مرحله در زمان برداشت و سه مرحله در طول دوره انبارداری انجام شد. نتایج نشان داد که OSP در طول زمان رسیدن و انبارداری به میزان کمی افزایش یافت. در هر دو رقم مورد مطالعه، میزان OSP در میوه‌هایی که پانزده روز در یک زمان مناسب برداشت، جمع آوری شده بودند بیشتر از میوه‌هایی که در زمان مناسب برداشت شده بودند بود. همچنین میزان OSP در میوه‌هایی که پانزده روز در بعد از برداشت نرم شده بود بطور معنی‌داری از زمان برداشت بیشتر بود. در هر دو رقم مورد مطالعه همیتک‌گی بالایی بین سفینی بافت میوه، IB مانند PGA و OSP مشاهده گردید. چنین می‌توان تجربه گیری نمود که سفینی بافت میوه و عارضه فهنه‌ای شدن داخلی میوه به میزان همیتک‌گی دارد. همچنین میوه‌هایی که با کلسیم محلول باشند و زودتر برداشت شده بودند پس از مدت طولانی انبارداری از میزان کمتری IB و PGA، OSP بروز می‌نماید.