Effects of Supplemental Phytase and Xylanase on Phytate Phosphorus Degradation, Ileal Protein and Energy Digestibility of a Corn-soybean-wheat Bran Diets in Broiler Chicks.

J. Pourreza^{1*} and H. L. Classen²

ABSTRACT

Effects of supplemental phytase and xylanase in a corn-soybean meal based diet containing 250 g/Kg wheat bran, were investigated. on broiler performance and phosphorus degradability and nitrogen digestibility. Phytase (500 and 1000 FTU) and xylanase (2700 and 5400 EXU) individually and in combination were added to the basal diet and given to 6 replicate groups (4 birds per replicate). At 21 days of age, birds were weighed and killed , ileal contents were collected and analyzed for P, N, chromic oxide and gross energy. Added phytase significantly (P<0.01) improved feed conversion ratio (FCR) (1.59 vs 1.62 g /g) and tibia ash (464.4 vs 444.3 g/Kg). Protein digestibility was increased (81.7 vs 79.4%) significantly (P<0.01) by 500 FTU/Kg added phytase. Supplemental xylanase significantly (P<0.01) improved FCR (1.58 vs 1.63 g/g) and protein digestibility (82.0 vs 80.4%). Phytate P degradability was significantly (P<0.02) improved by added phytase (41.4 vs 27.8%). Combination of supplemental phytase and xylanase had some beneficial effects on improving the nutritive value of diets containing wheat bran for broilers.

Keywords: Broiler, Phytase, Wheat bran, Xylanase.

INTRODUCTION

Wheat bran may be included in layer diets, but only in smaller amount in broiler diets owing to its high content of fiber and other antinutritional factors. Wheat and rice brans are two phosphorus-rich by-products that have feeding value for non-ruminant animals (Kratzer *et al.*, 1974; Farrell and Martin, 1998b). Phosphorus content of wheat bran is high (NRC, 1994) but most of it is present as phytate which is not available to poultry (Corley *et al.*, 1980). Phytic acid in wheat bran not only reduces P availability (<23%, Corley *et al.*, 1980), but also complexes with some other minerals and reduces their availability. In recent years, much attention has been focused on improving nutrient utilization of low quality or inferior feed ingredients with the aim of reducing feed costs (Farrell and Martin,1998a). Such improvement has been achieved by exogenous enzyme supplementation to the diets of poultry (Scott *et al.*, 1988, Farrell *et al.*, 1993;Yi *et al.*, 1996a;).

Non-starch polysaccharides, phytate, tannins and other antinutritional factors in cereal grains and their by-products reduce their digestibility and nutrient availability, and hence their feeding value (Annison *et al.* 1995). Results of many experiments indicated that enzyme supplementation of poultry diets improved the nutritional value of

¹ Department of Animal Science, Isfahan University of Technology, Isfahan, Islamic Republic of Iran, Email: Japour @ cc.iut.ac.ir, Fax: ++ 98313912254.

² Department of Animal and Poultry Science, University of Saskatchewan, 51 Campus Drive, Saskatoon, Saskatchewan, Canada, S7N 5A8.

^{*} Corresponding author

cereal grains and their by-products. Improvements in AME (Bedford *et al.*, 1998), starch digestibility (Choct and Annison, 1992), and phytate utilization (Simons *et al.*, 1990; Kornegay *et al.*, 1996) due to added enzymes have been reported.

Data on the effect of enzyme supplementation of wheat bran for poultry are limited. Addition of phytase and other enzymes could improve phytate degradability of wheat bran and such improvement may allow wheat bran to be used at higher levels in poultry diets and to supply considerable proportions of phosphorus requirements and also reduce feed cost and environmental pollution.

The objective of this study was to investigate the effects of different combinations of

Table 1. composition of the basal diet.

xylanase and phytase on performance, phosphorus, protein and energy utilization in broiler chickens fed on a high wheat bran basal diet.

MATERIALS AND METHODS

Birds and Diets

Two hundred and sixteen-day old male broiler chickens (Hubbard x Peterson) were weighed individually and divided into 54 groups of 4 (6 replicates per treatment) and each group was assigned to one of nine dietary treatments. Initial body weight of all replicates was nearly identical. The chicks were housed in battery cages and whole

Ingredient	g/kg
Wheat bran	250.0
Yellow com	375.0
Soybean meal (48% protein)	285.0
Canola oil	50.0
Dicalcium phosphate ^{<i>a</i>}	6.0
Limestone	17.0
Vitamins & minerals premix ^b	5.0
Sodium chloride	2.2
Choline chloride 70%	1.0
DL- Methionin	2.0
L- Lysine HCl	0.3
Coccidiostat	1.0
Growth promotant	0.5
Chromic oxide	5.0
Enzymes ^c	
Calculated analysis	
ME (Kcal/kg)	2910
Crude protein (g/kg)	209.0
Calcium (g/kg)	9.0
Non-phytate P (g/kg)	2.9
Methionine+cystine (g/kg)	8.15
Lysine (g/kg)	10.6
Linoleic acid (g/kg)	14.0
Crude fibre (g/kg)	47.0

^aDicalcium phosphate 220 g/Kg Ca and 187 g/Kg P.

^bSupplied per kilogram of diet: vitamin A (retinyl acetate+retinyl palmitate), 11000 IU;vitamin D, 2200 IU; vitamin E, 30 IU; menadione, 2.0 mg; thiamine, 1.5 mg; riboflavin, 6.0 mg; niacin, 60.0 mg; pyridoxine, 4.0 mg; vitamin B12, 0.02 mg; pantothenic acid, 10.0 mg; folic acid, 0.6 mg; biotin, 0.15 mg; iron, 80.0 mg; zinc, 80.0 mg; manganese, 80.0 mg; copper, 10.0 mg; iodine, 0.8 mg; and selenium, 0.3 mg.

^cNatuphos was added at the levels of 0.0, 0.1 and 0.2 g/Kg to provide 0.0, 500 and 1000 FTU/kg of phytase respectively. Avizyme 1300 was added at the levels of 0.0, 1 and 2 g/kg to provide 0.0, 2700 and 5400 EXU/kg of xylanase, respectively.

room brooding was used. Initial brooding temperature was 35°C and gradually reduced to 28°C by 21 d of age. Chicks had free access to feed and nipple drinkers during a 21 day experimental period.

A corn-soybean meal based diet contained 250g wheat bran/Kg (Table 1) supplemented with two kinds of phytase (Natuphos. 5000 FTU/g: BASF Corp., 3000# Continental Drive North, Mount Olive, NJ, U.S.A 07828) and xylanase (Avizyme 1300, 2700 EXU/g: Finnfeeds International, Box 777, Marlborough, Wiltshire, UK, SN8 1XN), each given individualy at three defferent levels (0, 500 and 1000 FTU/Kg phytase and 0, 2700 and 5400 EXU/Kg xylanase) and their combinations were tested. Dietary available P level was formulated at 2.9g/Kg, which was 30% below the current National Research Council (NRC, 1994) recommendations, to ensure a maximum response with phytase additions. Diets were otherwise formulated to meet the requirements of broiler chickens as established by NRC (1994).

Measurements and Analytical Methods

At 21 d of age, birds were weighed out and feed consumption and conversion during this period were measured out. All birds were killed by cervical dislocation and the lower section of ileal contents were gently squeezed, pooled and frozen for analysis.

The viscosity of fresh ileal contents was determined using a Brookfield (Brookfield Engineering Labs, Stoughton, MA 0207) viscometer (model LVDVII+Cp). The viscosity of five ileal samples were not determined because of the lack of supernatant after centrifugation, and those samples were therefore considered as missing values. Left tibia of two birds in each replicate were removed, pooled and stored at -20° C for tibia ash determination.

Ileal digesta were freeze-dried and then ground for gross energy, nitrogen, phytate and chromic oxide determination. Diets and digesta were ground to pass a 0.5 mm sieve. Chromic oxide was determined according to the method of Fenton and Fenton (1979). Gross energy was measured with a Parr 1720 Automatic Calorimeter (Parr Instrument Company, 21 Fifty Third Street, Moline, Illinois, U.S.A. 61265,) and nitrogen was measured using a nitrogen analyzer, lesed on the procedures of AOAC (1995). Apparent energy digestibility coefficients were calculated by the following formula.

Apparent digestible enegy = (GE/Cr203)d- $(GE/Cr203)i/(GE/Cr_{203})i$.

Where (GE/Cr203) d= the ratio of GE and Cr203 in diet and (GE/Cr203)i =the ratio of GE and Cr203 in ileal digesta.

Phytate was extracted by the method of Lehrfeld (1989) and measured using a Beckman (Beckman Instruments, 1045 Tristar Drive, Mississuaga, Ontario, Canada, L5T 1W5) HPLC anion-exchange chromatography with post-column reagent for detection (Rounds and Nielsen, 1993; Newkirk and Classen, 1997).

The bones were thawed and fat extracted in a Soxhlet extractor for 18 h with petroleum ether, then dried at 110°C for two h and ashed at 600°C for 24 h.

Statistical Analysis

All data were analyzed as a 3*3 factorial arrangement in a completely randomized design using the General Linear Model Procedure of SAS (1985) and if significant Duncan's multiple range test was used to compare means.

RESULTS

Body Weight, Feed Conversion and Tibia Ash

The effects of phytase and xylanase on body weight gain, feed intake, and feed conversion ratio (FCR), tibia ash and ileal viscosity are shown in table 2. Only FCR and tibia ash were improved significantly (P<0.01) by 500 FTU/Kg phytase. There

Table 2. Effects of wheat bran and dietary phytase and xylanase supplementation on body weight gain, feed consumption, feed conversion ratio, ileal digesta viscosity, tibia ash, phytate P degradability, energy and protein digestibility of male broiler chicks at 21 d of age.

Deit	Enzyme U/kg		Body	Feed	FCR	Viscos-		DE	Phytate P	
No	U/K	g	Wt.gain	intake		sity	ash		degrad-	digesti-
									ability b	ability ficient
	Distance V 1.		(.)	(.)	(F 1/)	((. /1 .)	(IZ - 1/1 -)		
	Phytase (FTU)	Xylanase (EXU)	(g)	(g)	(Feed/gian)	(ceps)	(g/kg)	(Kcal/kg)	%	%
1	0	0	499	828	1.66a ¹	4.02	437b	2723.2	22.2bc	80.2ab
2	500	0	523	831	1.60ab	2.95	466a	2830.0	46.5a	81.8a
3	1000	0	507	830	1.63ab	3.16	453ab	2727.2	43.4a	80.0ab
4	0	2700	484	780	1.61ab	2.92	455ab	2815.0	39.5ab	80.5a
5	0	5400	505	806	1.60ab	2.62	453ab	2727.0	10.7c	77.6b
6	500	2700	508	806	1.58ab	2.57	453ab	2878.0	43.2a	83.0a
7	500	5400	530	838	1.58ab	2.70	466a	2743.3	34.6ab	80.6a
8	1000	2700	528	861	1.63ab	2.52	452ab	2915.3	37.8ab	82.6a
9	1000	5400	511	800	1.56b	4.16	457ab	2871.7	24.9bc	81.7a
SE	1000	2100	± 21.4	30.0	0.022	1.22	8.0	63.8	5.7	0.97
Main e	effect			20.0	0.022	·· 	0.0	00.0	0.1	0.57
	se FTU/Kg									
1 11 9 144	0	0	496.2	829.5	1.62a	3.24	444.3b	2755.2	27.8b	79.4b
	500	0	520.3	825.1	1.59b	2.75	464.4a	2817.0	41.4a	81.7a
	1000	0	515.5	830.2	1.61ab	4.60	435.2ab		454.0ab	81.4a
SE			12.60	17.40	0.013	4.60	0.93	36.80	3.30	0.56
Xylan	ase EXU/Kg									
5	0	0	509.7	829.5	1.63a	3.40	452.0	2760.0	37.4a	80.4
	0	2700	506.6	816.0	1.61ab	3.67	453.3	2869.5	39.7a	82.0a
	0	5400	515.7	814.6	1.58b	4.44	458.4	2780.7	23.4b	80.0b
SE			12.60	17.40	0.0130	4.60	0.93	36.80	3.30	0.56
Probal	bility (P<)									
Phytas			NS	NS	**	NS	**	NS	*	**
Xylan			NS	NS	*	NS	NS	NS	**	*
2	se x xylanase		NS	NS	*	NS	*	NS	**	*

1 Values in each colums followed by the same letter are not significantly different.

* (P< 0.01)

** (P< 0.001) NS. Not significant

was no significant difference between 500 and 1000 FTU/Kg phytase levels. Supplemental xylanase (5400 EXU/Kg) improved FCR significantly (P<0.01). The interaction of phytase and xylanase regarding FCR and tibia ash was significant (P<0.05) (table 2).The best FCR was obtained with dietary combination of 1000 FTU/Kg and 5400 EXU/Kg of phytase and xylanase, respectively. Tibia ash from the birds that consumed diets containing phytase and xylanase had a higher ash content. There was no mortality during the experimental period.

DE, Protein Digestibility and Phosphorus Utilization

The effect of phytase supplementation on DE of diets was not significant, but added phytase at 500 FTU improved DE (table 2). The highest but not significant DE was obtained with 1000 FTU/Kg phytase and 2700 EXU/Kg xylanase respectively. Xylanase supplementation tended to improve dietary DE. The interaction of phytase and xylanase for dietary DE was not significant. The effects of phytase and xylanase on ileal protein digestibility were significant (P<0.01). Phytase added at 500 FTU/Kg improved protein digestibility significantly (P<0.01), but increasing the supplement to 1000 FTU/Kg had no further effect. Protein digestibility was improved significantly (P<0.01) by

2700 EXU/Kg xylanase. The difference between 5400 EXU/Kg and the control diet was not significant in this regard.

Supplemental phytase improved phytate P degradability significantly (P<0.02). Phytate P degradability was not improved significantly by xylanase. The interaction between phytase and xylanase regarding phytate P degradability was significant (P<0.01). Xylanase alone or in combination with 1000 FTU/Kg phytase significantly (P<0.01) reduced phytate P degradability. 5400 EXU/Kg Xylanase alone or in combination with 1000 FTU/Kg phytase decreased FCR, DE, protein digestibility and phytate P degradability when compared with 2700 EXU/Kg.

DISCUSSION

Phytase supplementation improved FCR and tibia ash, thus indicating a better utilization of phytate and other dietary constituents particularly energy and nitrogen. These results are in agreement with the previous findings in broilers (Kornegay et al. 1996; Yi et al. 1996a; Qian et al. 1996a; Sebastian et al. 1996; Cabahug et al. 1999; Ravindran et al. 2000), and in turkeys (Qian et al. 1996a; Yi et al. 1996b) suggesting an improvement in AME and N retention due to supplemental phytase. Farrell and Martin (1998a) found significant increases in dry matter, N and P retention by adding phytase to diets containing rice bran for ducks. Improvement in energy value of wheat due to supplemental phytase and xylanase individually and in combination was reported by Ravindran et al. (1999). This is in agreement with the results of our experiment.

Xylanase improved FCR, energy and protein digestibility. It seems that xylanase breaks the cell wall and liberates more nutrients for digestion and absorption. The combination of these two enzymes gave a better response regarding FCR, DE and protein digestibility. The low ileal viscosity found in this experiment (table 2) supported the idea that NSP's present in wheat bran contained no anti-nutritional factors and are not detrimental to broiler performance. Such a conclusion was reached by Farrell and Martin (1998b) with rice bran. This may be the reason for the lack of any significant interaction between phytase and xylanase regarding DE and body weight gain.

Improvement in phytate P degradability by the addition of phytase is in agreement with the findings of Farrell and Martine (1998b), Simons *et al.* (1990), Yi *et al.* (1996b) and Ravindran *et al.* (2000). The reason for the negative effects of 5400 EXU/Kg xylanase on phytate P degradation is unclear.

In conclusion, there was some beneficial response to phytase addition to all experimental diets with wheat bran. In addition, xylanase supplementation had some beneficial effects on FCR, DE and N digestibility. Added phytase improved DE and N digestibility in addition to P utilization. Feed intake was not increased by added phytase; therefore, improvements in FCR, DE and P utilization could be due to positive effects of supplemental phytase on these parameters. Combination of supplemental phytase and xylanase had some discauble benefits in improving the nutritional value of diets containing wheat bran for broilers.

ACKNOWLEDGEMENTS

The financial support of the Isfahan University of Technology is gratefully acknowledged. The authors wish to thank the Poultry Research staffs at the University of Saskatchewan for their technical assistance.

REFERENCES

- Annison, G., Moughan, P. J., and Thomas, D. V. 1995. Nutritive Activity of Soluble Rice Bran Arabinoxylanase in Broiler Diets. *Brit. Poult. Sci.*, 36: 479-488.
- Association of Official Analytical Chemists. 1980. Official Methods of Analysis. 13th ed. Washington, DC, AOAC.
- Bedford, M. R., Scott, T. A., Silverside, F. G., Classen, H. L., Swift, M. L., and Pack,

M. 1988. The Effect of Wheat Cultivar, Growing Environment, and Enzyme Supplementation on Digestibility of Amino Acids by Broilers. *Can. J. Anim. Sci.*, **78**: 335-342.

- Cabahug, C., Ravindran, V., Selle, P. H., and Bryden, W.L. 1999. Response of Broiler Chickens to Microbial Phytase Supplementation as Influenced by Dietary Phytic Aacid and Non-phytate Phosphorus Contents. I. Effects on *Bird Performance and Toe Ash*. Brit. Poult. Sci., 40: 660-666.
- Choct, M., and Annison, G. 1992. The Inhibition of Nutrient Digestion by Wheat Pentosans. *Brit. J. Nutr.*, 67: 123-132.
- Corley, J. R., Baker, R. D., and Easter, A. E. 1980. Biological Availability of Phosphorous in Rice Bran and Wheat Bran as Effected by Pelleting. *J. Anim. Sci.*, **50**: 286-292.
- Farrell, D. J., and Martin, E. A. 1998a. Strategies to Improve the Nutritive Value of Rice Bran in Poultry Diets. I. The Addition of Food Enzymes to Target the Non-starch Polysaccharide Fractions in Diets of chickens and Ducks Gave no Response. *Brit. Poult. Sci.*, **39**: 549-554.
- Farrell, D. J., and Martin, E. A. 1998 b. Strategeis to Improve the Nutritive Value of Rice Bran in Poultry Diets. III. The Addition of Inorganic Phosphorus and Phytase to Duck diets. *Brit. Poult. Sci.*, **39**: 601-611.
- Farrell, D. J., Martin, E. A., Dupreez, J. J., Bomgarts, M., Betts, M., Sudaman, A., and Thomson, E. 1993. The Beneficial Effects of a Microbial Food Phytase in Diets of Broiler Chickens and Ducklings. J. Anim. Physio. & Anim. Nutr. 69, 278-283.
- Fenton, T. W., and Fenton, M. 1979. Determination of Chromic Oxide in Feed and Feces. *Can. J. Anim. Sci.*, 58: 631-635.
- Kratzer, F. H., Earl, L., and Chiaravamont, C. 1974. Factors Influencing the Feeding Value of Rice Bran for Chicken. *Poult. Sci.*, 53: 1795-1780.
- Kornegay, E. T., Denbow, D. M., Yi, Z., and Ravindran, V. 1996. Response of Broilers to Graded Levels of Natuphos. Phytase Added to Corn-soybean-meal-based Diet Containing Three Levels Non-phytic Phosphorus. *Brit. J. Nutr.*, **75**, 839-852.
- Lehrfled, J. 1989. High-performance Liquid Chromatograph Analysis of Phytic Acid on a pH-stable, Macroporous Polymer Column. *Cereal Chem.*, 66: 510-515.

- 14. Newkirk, R. W., and Classen, H. L. 1997. In Vitro Hydrolysis of Phytate in Canola Meal with Purified and Crude Sources of Phytase. *Anim. Feed Sci. Techno.*, **72**: 315-327.
- National Research Council.1994. Nutrient Requirements of Poultry. 8th Revised edn (Washington, D. C., National Academy of Science).
- Qian, H., Veit, H. P., Kornegay, E. T., Ravindran, V., and Denbow, D. M. 1996a. Effects of Supplemental Phytase and Phosphorus on Histological and Other Tibia Bone Characteristics and Performance of Broilers Fed Semi-purified Diets. *Poult. Sci.*, 75: 618-626.
- Qian, H., Kornegay, E. T., and Denbow, D. M. 1996b. Phosphorus Equivalence of Microbial Phytase in Turkey Diets as Influenced by Calcium to Phosphorus Ratios and Phosphorus Levels. *Poult. Sci.*, **75**: 69-81.
- Ravindran, V., Selle, P. H., and Bryden, W.L. 1999. Effects of Phytase Supplementation, Individually and in Combination, with Glycanase, on the Nutritive Value of Wheat and Barley. *Poult. Sci.*, 78: 1588-1595.
- Ravindran, V., Cabahug, S., Ravindran, G., Selle, P. H., and Bryden, W.L. 2000. Response of Broiler Chickens to Microbial Phytase Supplementation as Influenced by Dietary Phytic Acid and Non-phytate Phosphorous Levels. II. Effects on Apparent Metabolizable Energy, Nutrient Degestibility and Nutrient Retention. *Brit. Poult. Sci.*, **41**:193-200.
- Rounds, M. A., and Nielsen, S. S. 1993. Anion-exchange High-performance Liquid Chromatography with Post-column Detection for the Analysis of Phytic Acid and other Inositol Phosphates. J. Chromatogr. A., 653: 148-152.
- 21. SAS, User's Guide: Statistics. Version 6. 4th ed., Vol. 2:SAS Institute, Cary, NC. 1686 pp.
- Scott, T. A., Silverside, F. G., Classen, H. L., Swift, M. L., Bedford, M. R., and Hall, J. W. 1998. A Broiler Chick Bioassay for Measuring the Feeding Value of Wheat and Barley in Complete Diets. *Poult. Sci.*, 77: 449-455.
- Sebastian, S., Touchburn, S. P., Chavez, E. R., and Lague, P. C. 1996. The Effects of Supplemental Microbial Phytase on the Performance and Utilization of Dietary Calcium, Phosphorus, Copper and Zinc in Broiler Chickens Fed Corn-soybean Diets.

Poult. Sci.,75: 729-736.

- Simons, P. C. M., Versteegh, H. A. G., Jongbloed, A. W., Kemme, P. A., Slump, P., Bos, K. D., Wolters, M. G. E., Beudeker, R. F., and Verschoor, G. J. 1990. Improvement of Phosphorus Availability by Microbial Phytase in Broilers and Pigs. *Br. J. Nutr.*, 64: 525-540.
- 25. Yi, Z., Kornegay, E. T., and Denbow, D. M. 1996a. Effect of Microbial Phytase on Nitro-

gen and Amino Acid Digestibility and Nitrogen Retention of Turkey Poults Fed Corn Soybean Meal diets. *Poult. Sci.*, **75**: 979-990.

26. Yi, Z., Kornegay, E. T., Ravindran, V., and Denbow, D. M. 1996b. Improving Phytate Phosphorus Availability in Corn and Soybean Meal for Broilers Using Microbial Phytase and Calculation of Phosphorus Equivalence of Phytase. *Poult. Sci.*, **75**: 240-249.

اثرات فیتاز و زایلاناز بر قابلیت استفاده از فسفر فیتاتی، قابلیت هضم پروتئین و انرژی قابل سوخت و ساز جیرههای حاوی سبوس گندم در جوجههای گوشتی

چکیدہ

اثرات مکمل های فیتاز و زایلاناز در جیره پایه ذرت – کنجاله سویا حاوی ۲۵درصد سبوس گندم بر عملکرد و قابلیت هضم فسفر و ازت و جوجه های گوشتی مورد بررسی قرار گرفت. در یک طرح کاملا " تصادفی به ترتیب سه سطح آنزیم فیتاز (۵۰۰ و ۱۰۰۰ واحد در کیلو گرم) و سه سطح انزیم زایلاناز (۲۷۰۰ و ۵۵۰ واحد در کیلو گرم) و ترکیب آنها به جیره پایه اضافه شد. هر یک از ۹ جیره آزمایشی به مدت ۲۱ روز به شش تکرار، در هر تکرار حاوی داده شد. در پایان آزمایش، جوجه های هر تکرار به صورت گروهی توزین و با جابجایی مهره گردن کشته شدند. محتویات ایلئوم جمع آوری و میزان ازت، فسفر، اکسید کرم و انرژی خام مورد بررسی قرار گرفت. مکمل فیتاز باعث بهبود معنی دار (۲۰/۰ > P) ضریب تبدیل غذا و خاکستر استخوان شد. قابلیت هضم پروتئین دراثر افزودن ۵۰۰ واحد فیتاز به هر کیلو گرم غذا به طور معنی داری (۲۰(۱ - ۷) افزایش یافت. مکمل زایلاناز، باعث بهبود معنی دار (۲۰/۰ > P) ضریب به طور معنی داری (۲۰(۱ - ۷) افزایش یافت. مکمل زایلاناز، باعث بهبود معنی دار (۲۰/۰ > P) ضریب به طور معنی داری (۲۰/۱ حاوی سبوس گندم برای جوجه های گوشتی بودند. داران ازت مغیری دار تبدیل غذا انرژی قابل هضم و قابلیت هضم پروتئین شد. مکمل فیتاز و زایلاناز دارای اثرات میدی در بهبود ازش غذایی جبره ای حاوی سبوس گندم برای جوجه می گوشتی بودند.