

The Effect of Salinity on Growth, Inorganic Ions and Dry Matter Partitioning in Sugar Beet Cultivars

A. R. Dadkhah^{1*} and H. Grrifiths²

ABSTRACT

This study investigated intra-specific variation in the growth parameters and dry matter partitioning of sugar beet cultivars at different levels of salinity (0, 50, 150, 250 and 350 mM NaCl + CaCl₂ in 5:1 molar ratio) using four cvs, one British (Madison) and three Iranian cvs (7233-P₁₂, 7233-P₂₁ and 7233-P₂₉). The plants were grown in a controlled greenhouse environment for about 18 weeks. Although growth parameters such as leaf area and dry matter accumulation were stimulated or unaffected at a low level of salinity (50 mM), higher salt concentrations significantly decreased all growth traits. At a high level of salinity, cv P₂₉ showed a significantly higher leaf area and total dry matter than others at 16 weeks of salt treatment. Pre-dawn leaf-water potential (Ψ_L) and shoot water content (SWC) were decreased by salinity. Cvs Madison and P₁₂ had significantly lower Ψ_L (more negative) and shoot water content at high levels of salt treatment (250 and 350 mM), while P₂₉ and P₂₁ showed higher Ψ_L (less negative) and shoot water content at the same salt concentrations. Analysis of ion accumulation revealed that Na⁺ and Cl⁻ contents were greatly increased in leaves under saline conditions. However, salt tolerant P₂₉ had lower Na⁺ and Cl⁻ concentrations than other cvs at high levels of salinity. The root dry matter of sugar beet cvs at the lowest and the highest salt concentrations reduced by 23.3% and 89.8%, respectively compared to the non-stressed plants, while shoot dry matter decreased by 1.1% and 77% at the same salt concentrations, respectively, after 16 weeks of salt treatment. The pattern of dry matter partitioning to different parts of the plant was changed by salt stress and the dry matter allocated to the storage roots was reduced by 6% and 18% at the lowest and the highest salt concentrations, respectively, at 16 weeks after salinisation.

Keywords: Dry matter partitioning, Growth, Salinity, Sugar beet.

INTRODUCTION

Salinity is one of the major constraints on crop production in numerous parts of the world, especially in arid and semi-arid regions, where soil and water-borne salts become concentrated due to inputs of irrigation water and high rates of evapotranspiration (Kingsbury *et al.*, 1984). Plants vary in their ability to cope with salinity, being capable of growth in a wide diversity of habitats ranging from non-saline environments to salt marshes. Differences in salt tolerance exist not only between species but also amongst

genotypes of certain species. This latter aspect attracts increasing studies on the mechanism of salt tolerance and applied research such as adaptation of crop species to saline soils (Polyakoff-Mayber and Lerner, 1999).

Sugar beet is a glycophytic member of *Chenopodiaceae* (Ghoulam and Fares, 2001). It is sensitive to elevated salinity at germination and the early seedling phase of development (Ghoulam and Fares, 2001) and tolerant at later growth stages (Katerji, 1997). The predominant influence of salinity stress on plants is growth suppression. It has

1. Ferdowsi University, Mashhad, Islamic Republic of Iran.

2. University of Cambridge, Cambridge, England.

* Corresponding author.



been reported that the fresh and dry weight of roots and leaves of sugar beet varieties were reduced with increasing salt concentration of irrigation water (Higazi *et al.*, 1995; Yassen *et al.*, 1998; Mekki and El-Gazzar, 1999). Growth reduction under salt stress may be because of osmotic stress due to lowering of the external water potential or the effect of ion toxicity on metabolic processes (De-Herralde *et al.*, 1998). Under salt stress, plants have evolved complex mechanisms allowing for adaptation to osmotic adjustment by the accumulation of compatible solutes such as glycinebetaine and proline (Yeo, 1998; Bohnert *et al.*, 1999) and lowering the toxic concentration of ions in the cytoplasm by restriction of Na^+ influx or its sequestration into the vacuole and/or its extrusion (Binziel *et al.*, 1988). In a saline environment, plants uptake excessive amounts of Na^+ at the cost of K^+ and Ca^{2+} (Rengle, 1992) and can lead to nutrient deficiencies (Muhammad *et al.*, 1987; Francois and Donovan, 1991). Sodium accumulation could disrupt membrane integrity and inhibit the transport of nutritional ions into the root and up to the shoot (Cramer *et al.*, 1994). As the main sugar beet growing areas in Iran are affected by salinity and this problem causes a serious handicap for cultivation and production of this agricultural crop, more tolerant sugar beet varieties must be selected and recommended for the saline areas. Accurate selection requires an understanding of the mechanisms involved in salt tolerance in this species. The objective of this present work is to assess four sugar beet varieties for their salt tolerance and to gain more information on dry matter partitioning and inorganic ions accumulation under salt stress.

MATERIALS AND METHODS

Plant Materials and Experimental Conditions

Four varieties of sugar beet (*Beta vulgaris* L.) three of Iranian origin (7233-P₁₂, 7233-P₂₁, 7233-P₂₉) plus one of British origin

(Madison) were used. The seeds of each cultivar were sown in a shallow polyethylene tray filled with vermiculite grade 0.2-1mm, and the seedlings were transplanted into 15 cm diameter plastic pots containing washed sand and with a saucer under them to prevent leaching after irrigation. Six weeks later, the plants were transplanted into 32 cm diameter plastic pots. Five salinity treatments 0, 50, 150, 250 and 350 mM ($\text{NaCl} + \text{CaCl}_2$ in 5:1 molar ratio) were imposed through irrigation from the time that most plants were at the 4-leaf stage. Salt was added to a modified Hoagland nutrient solution (Mass and Poss, 1989) in increments of 50 mM day⁻¹ to avoid salt shock. The plants were kept in a warm glasshouse where the temperature ranged between 15-35 °C for about 18 weeks. Relative humidity was 35% and 55% during the day and night, respectively.

Measurement of Growth Parameters and Leaf Water Potential

Growth parameters were measured as leaf area, number of leaves per plant and dry matter accumulation. Plants were harvested at 16 weeks after salt treatment and were separated into leaf lamina, petioles, stem and roots. The fresh weight of the aerial parts was determined immediately after harvesting. These components were dried for 48 hours at 75 °C in a conventional oven and weighed. Dry matter partitioning in the lamina, petioles, stem and root was determined after drying the whole samples. Leaf area was determined using a leaf area meter (Delta-T Devices Ltd., Cambridge, U.K). Pre-dawn leaf water potential (Ψ_1) of the youngest fully expanded leaves was measured by thermocouple psychrometers and a microvoltmeter Model 85 (J.R.D.Merrill Special Equipment, Logan, Utah, USA). Pre-dawn leaf water potential was determined on detached leaf discs (0.5 cm²) placed into a water bath at 25 °C for approximately 2 hours to equilibrate before readings were taken. To determine leaf wa-

ter content, the youngest fully expanded leaves were harvested early morning and the fresh weight of leaves was obtained, oven dried at 75 °C for 48 hours and reweighed.

Analysis of Inorganic Ions

For Na⁺ and K⁺ analysis, samples of dried leaves and roots were ashed in a furnace for 24 hours at 500 °C. The ash was dissolved in 50% hydrochloric acid, diluted in distilled water and filtered through a sheet of Whatman No. 2 filter paper. The Na⁺ and K⁺ contents were determined by flame photometer. Chloride (Cl⁻) determinations were made on a distilled water extract of the dried sample, shaken for 24 hours and then filtered through a 90mm Whatman No.2 filter paper. The amount of Cl⁻ was measured by ion-exchange chromatography using a DIONEX Model DX 500, fitted with a CD20 conductivity Detector, IP 25 pump and AS 14 Ion Exchange Column.

Data Analysis

This experiment was carried out based on a randomized complete block design. The data for all parameters were analysed using the analysis of variance procedure of the Statistical Analysis System (SAS) soft ware, version 6.12. Means were compared using a Duncan's multiple range test at 0.05 percent probability level for all the comparisons.

RESULT AND DISCUSSION

Leaf Dynamics

Live leaf area was greatly reduced by high levels of salinity (Table 1). Live leaf area of plants at the highest salinity (350 mM) was decreased by 77% (average of all 4 cvs) compared to the leaf area of non-stressed plants at 16 weeks after salinisation (Table 1). The leaf area of cvs P₂₉ and P₂₁ were not reduced by the low level of salinity (50

mM), while the leaf area of cv Madison was decreased by about 11.9% compared with non-stressed plants at 16 weeks of salinisation. The best cv, which had the least reduction (61.8%) in leaf area and the highest area (1923 cm²) at the highest level of salinity was P₂₉ (Table 1), while cv Madison showed the greatest reduction (83.8%) in leaf area and the lowest area (793 cm²) due to salinity at the same level of salt treatment (Table 1). Leaf numbers per plant decreased significantly as salinity increased in all cvs. However, a low level of salt treatment (50 mM) slightly decreased the leaf number per plant in all cvs, except cv P₂₉ which had more leaves at a low level of salinity in comparison with its control. This increase was not, however, statistically significant. Cv Madison had the greatest reduction in leaf numbers due to salinity amongst cvs at 16 weeks of salt treatment (Table 2). However, the number on leaves of Madison was higher than on other cultivars at all levels of salinity but its total leaf area was less than others. Therefore, it can be concluded that Madison genetically produces more, but smaller leaves. P₂₉ had the smallest reduction in leaf number due to salt at 16 weeks. The results show that leaf number was less affected than leaf area by salinity. It is suggested that most of the reduction in plant leaf area was caused by the inhibition of leaf expansion. This is consistent with the results of previous research, which showed that high levels of salinity decreased leaf area due to a combination of a decrease in cell number and in cell size (Brugnoli and Lauter, 1991; De-Herralde *et al.*, 1998). Munns and Termaat (1986) demonstrated that, for a given amount of NaCl transport to the shoot, reduction in leaf expansion results in the same proportional increase in the leaf NaCl concentration. Salt stressed barley plants produced smaller leaf areas, which caused a higher Na⁺ accumulation in a specific leaf area (Munns, 1985). Witkowski and Lamont (1991) reported that plants might reduce water loss by reducing their evaporation surface. Therefore, leaves tend to be smaller and thicker under saline conditions.



Table 1. Mean live leaf area of sugar beet plant ($\text{Cm}^2 \text{ plant}^{-1}$) and percentage of reduction at different levels of salinity. Each number is the average of six replications. Numbers followed by the same letter are not significantly ($P \leq 0.01$) different according to Duncan's multiple range test (different upper case and lower case letters show significantly differences within columns and rows, respectively).

Treatment		0 mM			50 mM			150 mM			250 mM			350 mM		
Variety		Actual		Reduction %	Actual		Reduction %	Actual		Reduction %	Actual		Reduction %	Actual		Reduction %
		Actual	Reduction %		Actual	Reduction %		Actual	Reduction %		Actual	Reduction %		Actual	Reduction %	
Madison		4889 ^A _a	0		4306 ^B _a	11.9		2366 ^B _b	51.6		1294 ^B _b	73.5		793 ^C _d	83.8	
P ₂₁		5370 ^A _a	0		5454 ^A _a	-1.6		2662 ^A _b	50.4		1743 ^{AB} _c	67.5		1092 ^B _d	79.7	
P ₂₉		5030 ^A _a	0		5370 ^A _a	-6.8		2650 ^A _b	47.3		2201 ^A _c	56.2		1923 ^A _c	61.8	
P ₁₂		5780 ^A _a	0		5714 ^A _a	1.1		3030 ^A _b	47.6		1695 ^{AB} _c	70.7		1036 ^B _d	82.1	
Average		5267	0		5211	1.1		2677	49.2		1733	67.1		1211	77	

Table 2. Mean number of leaves per sugar beet plant and percentage of reduction at different levels of salinity. Each number is the average of six replications. Numbers followed by the same letter are not significantly ($P \leq 0.01$) different according to Duncan's multiple range test (different upper case and lower case letters show significantly differences within columns and rows, respectively).

Treatment		0 mM			50 mM			150 mM			250 mM			350 mM		
Variety		Actual		Reduction %	Actual		Reduction %	Actual		Reduction %	Actual		Reduction %	Actual		Reduction %
		Actual	Reduction %		Actual	Reduction %		Actual	Reduction %		Actual	Reduction %		Actual	Reduction %	
Madison		32.7 ^A _a	0		26 ^A _b	20.5		23.3 ^A _b	28.7		17.3 ^A _c	47.1		14.5 ^A _d	59.3	
P ₂₁		22.3 ^B _a	0		21.7 ^A _a	2.7		16.3 ^B _{ab}	26.9		13.7 ^B _{ab}	38.6		11.3 ^B _b	49.3	
P ₂₉		17.7 ^B _{ab}	0		21.3 ^A _a	-20.3		17.7 ^B _{ab}	0		15 ^A _b	15.2		14.3 ^A _b	19.2	
P ₁₂		23.7 ^B _a	0		21 ^A _a	11.6		20.3 ^{AB} _a	14.3		14 ^B _b	40.9		13.7 ^{AB} _b	42.2	
Average		24.1	0		22.5	3.4		19.4	17.5		15	35.5		13.4	40	

Dry Matter Production and Partitioning

Shoot dry weight was decreased by salinity ($P \leq 0.0001$) (Table 3). The shoot dry weight of cv P₂₉ was stimulated at low level of salinity (50 mM). Cultivar P₂₉ had a higher dry weight than other cvs at high level of salinity, which can be attributed to greater leaf area (Table 1).

Root dry weight was also decreased by salinity (Table 4). An increasing shoot / root ratio with increasing salt concentration (Figure 1) showed that the growth rate of the

shoot/root ratio of Madison at all levels of salinity was less than in other cvs, possibly because of production of more root dry weight and less shoot dry weight than others (Tables 3,4) and selection by its breeders for diversion of a high proportion of assimilate to the root. A possible reason for dry matter reduction could be the greater reduction in uptake and utilization of mineral nutrients by plants under salt stress. Pessarakli and Tucker (1985, 1988) reported that the total nitrogen uptake of cotton plants decreased with increasing salinity, reflecting primarily

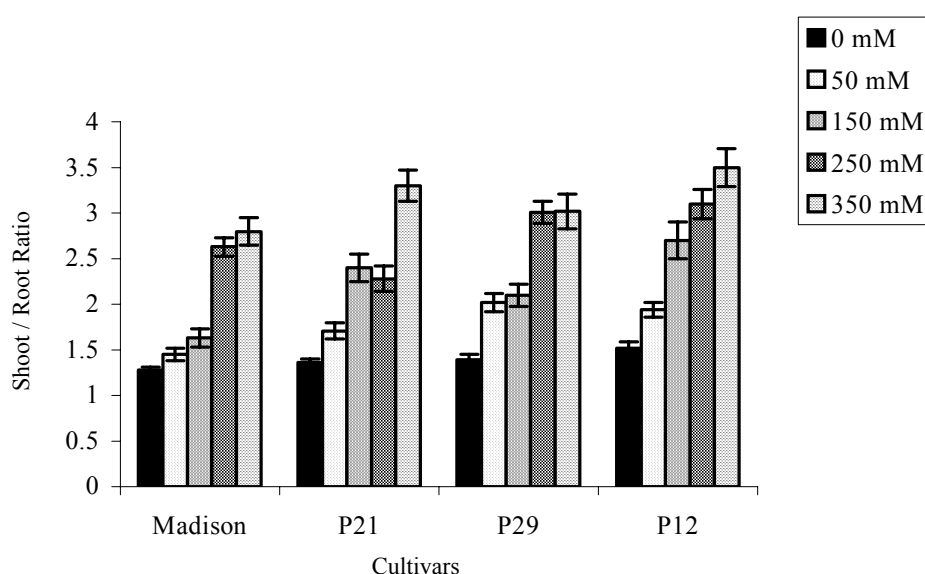


Figure 1. The shoot: root ratio of four sugar beet cultivars at 16 weeks after salinisation under different salt treatments. The values are mean of six replications (\pm S.E.).

shoot was not reduced as much as that of the root. It seems that root growth in sugar beet plants is more sensitive than shoot growth because, despite stimulation of shoot dry weight at low level of salinity (Table 3), root dry weight decreased by 23.3% at 16 weeks after salinisation (Table 4). This might be due to the type of sugar beet root (storage root) and also to water deficiency caused by concentration of salt in the growth medium. Abdollahian-Noghabi (1999) found that the shoot/root ratio of *Beta vulgaris* increased under drought stress conditions. The

a dry matter reduction. The uptake of nitrogen (N) in salt stressed plants might be competitively limited by Cl^- (Aslam *et al.*, 1984, Ward *et al.*, 1986).

The partitioning of photosynthetic materials among different organs is an important component in any description of crop growth. However, it has been given only limited attention to date, and many crop models include only simple definitions in which fixed fractions are partitioned to different organs throughout specific phases of development (Weir *et al.*, 1984). Salt stress



Table 3. Mean shoot dry weight (g.plant⁻¹) of sugar beet plant and percentage of reduction at different levels of salinity. Each number is the average of six replications. Numbers followed by the same letter are not significantly ($P \leq 0.01$) different according to Duncan's multiple range test (different upper case and lower case letters show significantly differences within columns and rows, respectively).

Treatment →		0 mM		50 mM		150 mM		250 mM		350 mM	
Variety ↓		Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %
Madison		60.3 ^A _a	0	57.0 ^A _a	5.5	35.3 ^A _b	41.5	16.6 ^B _c	11.5 ^B _d	81	
	P ₂₁	67.4 ^A _a	0	63.6 ^A _a	5.9	38.4 ^A _b	43	21.3 ^A _c	14.8 ^{AB} _c	78	
P ₂₉		64.7 ^A _a	0	69.4 ^A _a	-5.6	32.5 ^A _b	50.5	23.5 ^A _c	20.8 ^A _c	68.3	
	P ₁₂	72.4 ^A _a	0	72.8 ^A _a	-0.6	38.6 ^A _b	46.7	18.5 ^{AB} _c	14.7 ^{AB} _c	79.7	
Average		66.2	0	65.7	1.1	36.2	45.5	20	15.5	77	

Table 4. Mean root dry weight (g.plant⁻¹) of sugar beet plant and percentage of reduction at different levels of salinity. Each number is the average of six replications. Numbers followed by the same letter are not significantly ($P \leq 0.01$) different according to Duncan's multiple range test (different upper case and lower case letters show significantly differences within columns and rows, respectively).

Treatment →		0 mM		50 mM		150 mM		250 mM		350 mM	
Variety ↓		Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %
Madison	P ₂₁	49.4 ^A _a	0	39.4 ^A _b	20.2	21.6 ^A _c	55.9	6.3 ^A _d	87.3	4.1 ^B _e	91.7
		49.6 ^A _a	0	37.2 ^A _b	25.0	16.0 ^B _c	67.7	6.5 ^A _d	87.0	4.5 ^B _e	90.9
P ₂₉	P ₁₂	46.7 ^A _a	0	34.3 ^A _b	26.6	15.5 ^B _c	66.8	7.8 ^A _d	83.3	6.9 ^A _d	85.2
		47.7 ^A _a	0	37.5 ^A _b	21.4	14.4 ^B _c	69.8	6.0 ^A _d	87.4	4.2 ^B _d	91.2
Average		48.4	0	37.1	23.3	16.9	65.1	6.7	86.1	4.9	89.8

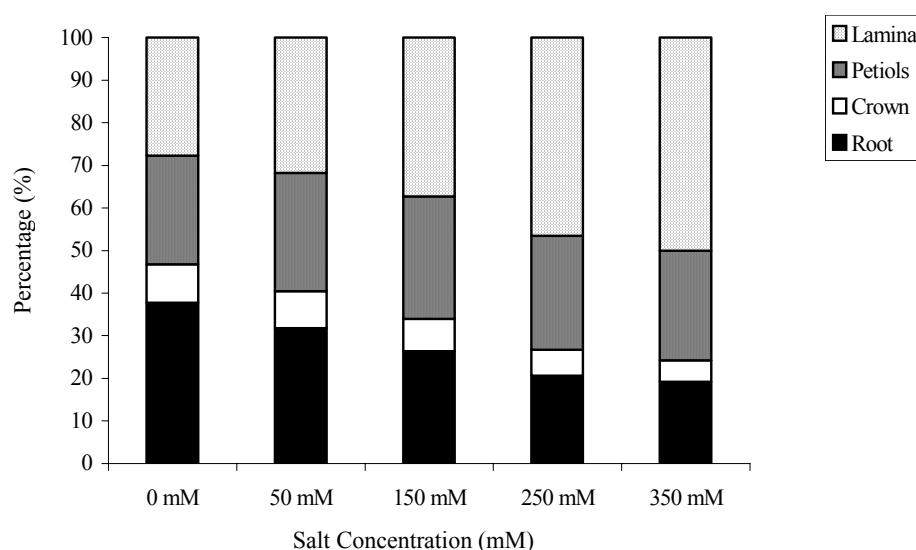


Figure 2. Overall effect of salinity on dry matter partitioning to leaf lamina, petiole, crown and storage root of four sugar beet cultivars at 16 weeks after salt treatment.

altered the pattern of dry matter partitioning in sugar beet plants (Figure 2). Despite 6% reduction in sugar beet root dry weight due to low level of salinity (50 mM), the lamina and petioles dry weight in sugar beet were stimulated by 3% and 2%, respectively. These results demonstrate that the high level of salinity reduced the proportion of dry weight allocated to the storage root by 18%. Conversely, high level of salinity increased the proportion of dry matter allocated to sugar beet lamina by 22%. These results are in agreement with Abdollahian-Noghabi and Williams (1998) who obtained similar results for dry matter allocation in sugar beet root under drought stress conditions.

Milford *et al.* (1988) found no evidence of a discontinuity in the partitioning between shoot and root at any stage in crop development under normal conditions, and they concluded that there is normally a progressive shift in partitioning to the storage root as the crop develops. Clover (1997) found that water deficiency had a variable affect of dry matter partitioning.

Leaf Water Potential (Ψ_L) and Shoot Water Content (SWC)

Shoot water content and leaf water potential were decreased by salinity (Tables 5 and 6). The shoot water content (SWC) of plants grown at low level of salinity (50 mM) was greater than in the control but not significantly different, except for P₁₂. The SWC of cvs Madison and P₁₂ were significantly decreased by increasing the salt concentration but no significant differences were observed in SWC between the control and any of the salinity levels in P₂₁ and P₂₉. Also with declining SWC, the values for leaf water potential (Ψ_L) reduced. Madison and P₁₂ had significantly the lowest Ψ_L (more negative, with values of -2.7 and -2.9 Mpa, respectively) at high levels of salt treatment (250 and 350 mM), while P₂₉ and P₂₁ had higher levels (less negative) at the same salt concentrations (Table 6). The percentage reduction of Ψ_L in Madison at high levels of salinity was more than twice that compared to P₂₁.

Salt treatment induced a reduction in shoot water content and Ψ_L . This reduction was

Table 5. Mean shoot water content (gH₂O/g dry weight) of sugar beet plant and percentage of reduction at different levels of salinity. Each number is the average of six replications. Numbers followed the same letter are not significantly (P ≤0.01) different according to Duncan's multiple range test (different upper case and lower case letters show significantly differences within columns and rows, respectively).

Treatment →		0 mM		50 mM		150 mM		250 mM		350 mM	
Variety ↓		Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %
Madison		7.55 ^A	0	7.70 ^{AB}	-1.99	6.80 ^A	9.67	6.30 ^{AB}	16.42	5.80 ^A	23.10
P ₂₁		6.95 ^A	0	6.99 ^B	-0.58	6.50 ^A	9.91	6.30 ^B	10.50	6.20 ^A	11.10
P ₂₉		7.39 ^A	0	7.80 ^{AB}	-6.10	7.10 ^A	3.65	6.60 ^A	10.40	6.50 ^A	12.60
P ₁₂		7.10 ^A	0	8.20 ^A	-15.2	6.75 ^B	5.10	6.60 ^A	6.89	5.50 ^A	23.10
Average		7.25	0	7.67	-6.03	6.78	6.33	6.46	10.06	5.99	17.45

Table 6. Mean leaf water potential (Mpa) of sugar beet plant and percentage of reduction at different levels of salinity. Each number is the average of six replications. Numbers followed the same letter are not significantly (P ≤0.01) different according to Duncan's multiple range test (different upper case and lower case letters show significantly differences within columns and rows, respectively).

Treatment →		0 mM		50 mM		150 mM		250 mM		350 mM	
Variety ↓		Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %	Actual	Reduction %
Madison		-1.05 ^A	0	-1.90 ^{AB}	80.95	-2.01 ^A	91.43	-2.40 ^{AB}	128.57	-2.70 ^A	157.14
P ₂₁		-1.40 ^A	0	-1.90 ^B	35.71	-2.10 ^A	50.0	-2.20 ^B	57.14	-2.50 ^A	78.57
P ₂₉		-1.10 ^A	0	-1.20 ^{AB}	9.09	-1.80 ^A	63.63	-2.00 ^A	81.81	-2.20 ^A	100
P ₁₂		-1.30 ^A	0	-1.70 ^A	30.77	-2.20 ^B	69.23	-2.70 ^B	107.69	-2.90 ^A	123.08
Average		-1.21	0	-1.68	38.84	-2.03	67.77	-2.34	93.39	-2.55	110.74

more important in the less tolerant variety, Madison, than in the more tolerant one, P₂₉. The decrease in SWC indicated a loss of turgor that resulted in limited water availability for the cell extension process (Katerji *et al.*, 1997). Salinity strongly reduced leaf water potential in all cvs. However, cvs P₂₁ and P₂₉ had significantly higher (less negative) Ψ_L than Madison and P₁₂ at high levels of salinity at 16 weeks of salt treatment. This may be attributed to a greater water loss in Madison and P₁₂ at higher levels of salinity or could be due to the transport of more Na⁺ and Cl⁻ to the leaves under saline conditions

(Heuer and Plaut, 1989).

Inorganic Ion Concentration

The presence of salt in the rooting medium induced a remarkable increase in the Na⁺ concentration in leaves and roots ($P \leq 0.0001$) (Figure 3). There were significant differences in leaf sodium concentration among cvs at different levels of salinity. As can be seen in Figure 3, cv P₂₉ had a significantly higher Na⁺ content in its leaves at the lowest salinity (50 mM) than cvs Madison and P₁₂. However, at the highest salinity

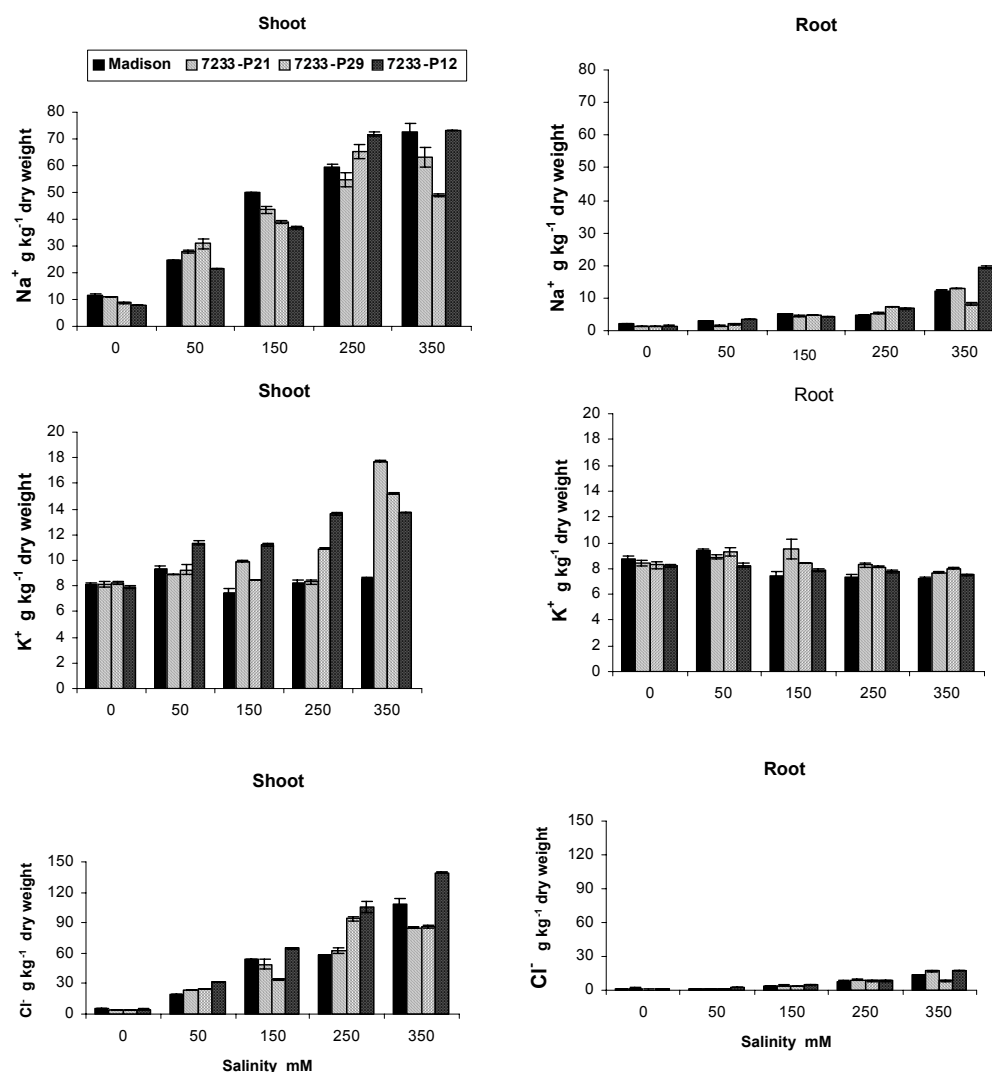


Figure 3. Concentration of Na⁺, K⁺ and Cl⁻ in shoot and root of four sugar beet varieties submitted to five salt treatments for 16 weeks. The values are means of six replications (\pm S.E.).



level cv P₂₉ had significantly lower sodium content while Madison and P₁₂ had the highest Na⁺. Leaf potassium (K⁺) concentration significantly decreased with increasing salinity to around half of the control values at 350 mM (Figure 3). Cl⁻ contents in leaves of all sugar beet varieties increased with increasing salt concentration (Figure 3). There were no significant differences in leaf Cl⁻ concentration between different cvs in control plants. However, P₁₂ had a significantly higher leaf Cl⁻ content than others at all levels of salt treatment. At high levels of salinity cvs P₂₉ and P₂₁ had significantly lower Cl⁻ content in leaves than other cvs. The four varieties tested accumulated more Na⁺ and K⁺ and Cl⁻ ions in the leaves than in the roots under salinity (Figure 3). These varieties are qualified as includers (Yeo and Flowers, 1985). Accumulation of inorganic ions, predominantly Na⁺ and Cl⁻, plays an important role in the process of osmotic adjustment. There is substantial evidence that plants of moderate to high salt tolerance may accumulate large amounts of salt under saline conditions, contributing to osmotic adjustment (Al-Kateeb, 1997; Ashraf and Ahmad, 2000; Ghoulam *et al.*, 2002). The increase in Na⁺ and Cl⁻ concentrations in the salinised expanded leaves of all cultivars was not similar. Cultivar P₂₉ (salt tolerant) accumulated less Na⁺ and Cl⁻ in its cells at high levels of salinity. Schachtment and Munns (1992) reported that the lower rate of Na⁺ accumulation in expanding leaves of *Triticum* species might be due to higher leaf expansion and/or a mechanism that limits the transport of uptake of Na⁺ by roots. The lower Na⁺ and Cl⁻ contents in P₂₉ at high levels of salinity can be attributed to its higher leaf expansion than other cvs at high salinity. Another possibility might be that this cv absorbs Na⁺ and Cl⁻ up to a threshold level and then changes to being a salt excluder.

CONCLUSION

This assessment of the effect of salinity on

the growth parameters in four sugar beet cvs allows us to conclude that all of the parameters considered were affected by salinity with a cultivar difference. Moreover, the Madison cv was the least tolerant and P₂₉ was the most tolerant cv. Water loss and inorganic ion accumulation were involved in the osmotic adjustment. Salt stress altered the pattern of dry matter partitioning in sugar beet. Dry matter allocated to the shoot was increased under saline conditions, suggesting that root growth of sugar beet is more sensitive to salinity than shoot growth and that economic yield of sugar beet is likely to be adversely affected by saline conditions.

REFERENCE

1. Abdollahian-Noghabi, M. (1999). Ecophysiology of Sugar Beet Cultivars and Weed Species Subjected to Water Deficiency Stress. Ph.D dissertation, The University of Reading, Reading.
2. Abdollahian-Noghabi, M., and Froud-Williams, R. J. (1998). Effect of Moisture Stress and Re-watering on Growth and Dry Matter Partitioning in Three Cultivars of Sugar Beet. *Aspects Appl. Biol.* **52**: 71-78.
3. Al-khateeb, S.A. (1997). Effect of NaCl and Na₂SO₄ on Growth, Ion Relations, Water Relations and Gas Exchange of Two *Atriplex* Species. University of Reading, Reading. pp. 230.
4. Ashraf, M. and Ahmad, S. (2000). Influence of Sodium Chloride on Ion Accumulation, Yield Components and Fiber Characteristics in Salt-tolerant and Salt-sensitive Lines of Cotton. *Field Crop Res.*, **66**: 15-127.
5. Aslam, M., Huffaker, R.C. and Rains, D.W. (1984). Early Effects of Salinity on Nitrate Assimilation in Barley Seedlings. *Plant Physiol.*, **76**: 321-325.
6. Binziel, M. L., Hess, F.D., Bressan, R.A. and Hasegawa, P.M. (1988). Intercellular Compartmentation of Ions in Salt Adapted Tobacco Cells. *Plant Physiol.* **86**: 607-614.
7. Bohnert, H.J., Su, H. and Shen, B. (1999). Molecular Responses of Salinity Tolerance. In: *Molecular Responses to Cold, Drought, Heat and Salt Stress in Higher Plants*, Shinzaki, K. and Yamaguchi-Shinozaki, K.

- (eds) University of Arizona, pp. 29-60.
8. Brugnoli, N. and Lauteri, M. (1991). Effect of Salinity on Stomatal Conductance, Photosynthesis Capacity and Carbon Isotope Discrimination of Salt Tolerant (*Gossypium hirsutum* L.) and Salt Sensitive (*Phaseolus vulgaris* L.) C₃ Non-halophytes. *Plant Physiol.*, **95**: 628-635.
 9. Clover, G.R.G. (1997). *Effects of Beet Yellows Virus and Drought on Growth of Sugar Beet*. University of Nottingham.
 10. Cramer, G. R., Alberico, G. J. and Schmidt, C. (1994). Leaf Expansion Limits Dry Matter Accumulation of Salt-stressed Maize. *Aust. J. Plant Physiol.*, **21**: 663-674.
 11. De-Herralde, F., Biel, C., Save, R., Morales, M. A., Torrecillas, A. and Alarcon, J. J. (1998). Effect of Water and Salt Stresses on the Growth, Gas Exchange and Water Relations in *Argyranthemum coronopifolium* Plants. *Crop Sci.*, **139**: 9-17.
 12. Francois, L. E., and Donovan, T. J. (1991). Calcium Deficiency of Artichoke Buds in Relation to Salinity. *Hortscience* **26**: 549-553.
 13. Ghoulam, C. and Fares, K. (2001). Effect of Salinity on Seed Germination and Early Seedling Growth of Sugar Beet (*Beta vulgaris* L.). *Seed Sci. Technol.*, **29**: 357-364.
 14. Ghoulam, C., Foursy, A. and Fares, K. (2002). Effects of Salt Stress on Growth, Inorganic Ions and Proline Accumulation in Relation to Osmotic Adjustment in Five Sugar Beet Cultivars. *Environ. Exp. Bot.*, **47**: 39-50.
 15. Hang, A.N. and Miller, D.E. (1986). Response of Sugar Beet to Deficit, High-frequency Sprinkler Irrigation. II: Sugar Beet Development and Partitioning to Root Growth. *Agron. J.*, **78**: 305-312.
 16. Higazy, M. A., Shehata, M. M., and Allam, A. I. (1995). Free Proline Relation to Salinity Tolerance of Three Sugar Beet Varieties. *Egypt. J. Agr. Res.* **13(1)**: 175-190.
 17. Heuer, B. and Plaut, Z. (1989). Photosynthesis and Osmotic Adjustment of Two Sugar Beet Cultivars Grown Saline Conditions. *J. Exp. Bot.*, **40**: 437-440.
 18. Katerji, N., van Hoorn, J.W., Hamdy, A., Mastroilli, M. and Moukarzel, E. (1997). Osmotic Adjustment to Sugar Beets in Response to Soil Salinity and its Influence on Stomatal Conductance, Growth and Yield. *Agr. Water Manage.*, **34**: 57-69.
 19. Kingsbury, R.W., Epstein, E. and Percy, R.W. (1984). Physiological Responses to Salinity in Selected Lines of Wheat. *Plant Physiol.*, **74**: 417-423.
 20. Mass, E. V., and Poss, J. A. (1989). Salt Sensitivity of Wheat at Various Growth Stages. *Irrig. Sci.*, **10**: 29-40.
 21. Mekki, B. B. and EL-Gazzar, M. M. (1999). Response of Root Yield and Quality of Sugar Beet (*Beat vulgaris* L.) to Irrigation with Saline Water and Follar Potassium Fertilization. *Ann. Agr. Sci.* **44(1)**: 213-225.
 22. Milford, G.F.J., Travis, K.Z., Pocock, T.O., Jaggard, K.W. and Day, W. (1988). Growth and Dry Matter Partitioning in Sugar Beet. *J. Agr. Sci.*, **110**: 301-308.
 23. Muhammad, S., Akbar, M. and Neue, H. U. (1987). Effect of Na/Ca and Na/K Ratios in Saline Culture Solution on the Growth and Mineral Nutrition of Rice (*Oryza sativa* L.). *Plant Soil* **104**: 57.
 24. Munns, R. (1985). Na⁺, K⁺ and Cl⁻ in Xylem Sap Flowing to Shoots of NaCl Treated Barley. *J. Exp. Bot.*, **36**: 1032-1042.
 25. Munns, R. and Termaat, A. (1986). Whole-Plant Responses to Salinity. *Aust. J. Plant Physiol.*, **13**: 143-160.
 26. Pessaraki, M. and Tucker, T. C. (1985). Uptake of Nitrogen-15 by Cotton under Salt Stress. *Soil Sci. Soc. Am. J.* **49**: 149-152.
 27. Pessaraki, M. and Tucker, T. C. (1988). Dry Matter Yield and Nitrogen-15 Uptake by Tomatoes under Sodium Chloride Stress. *Soil Sci. Soc. Am. J.*, **52**: 698-700.
 28. Polyakoff-Mayber, A. and Lerner, H. R. (1999). Plant in Saline Environments. In: "Handbook of Plant and Crop Stress". M. Pessaraki, (Ed.), Marcel Dekker, New York. 125 pp.
 29. Rengel, Z. (1992). The Role of Calcium in Salt Toxicity. *Plant, Cell Environ.*, **15**: 625-632.
 30. Schachman, D.P. and Munns, R. (1992). Sodium Accumulation in Leaves of *Triticum* Species that Differ in Salt Tolerance. *Aust. J. Plant Physiol.*, **19**: 331-340.
 31. Ward, M.R., Aslam, M. and Haffaker, R.C. (1986). Enhancement of Nitrate Uptake and Growth of Barley Seedlings by Calcium under Saline Conditions. *Plant Physiol.*, **80**: 820-824.
 32. Weir, A.H., Bragy, P.L., Porter, J.R. and Rayner, J.H. (1984). A Winter Wheat Crop Simulation Model without Water or Nutrient Limitations. *J. Agr. Sci.*, **102**: 371-382.
 33. Witkowski, E. T. F. and Lamont, B. B.



- (1991). Leaf Specific Mass Confounds Leaf Density and Thickness. *Oecologia* **88**: 486.
34. Yassen, B. Y. and Jurgees, J. A. (1998). The Response of Sugar Beet Leaf Growth and its Ionic Composition to Sodium Chloride. *J. Agr. Water Res. Res., Soil and Water Resour.* **7** (1), 47-59.
35. Yeo, A.R. (1998). Molecular Biology of Salt Tolerance in the Context of Whole Plant Physiology. *J. Exp. Bot.*, **49**: 915-929.
36. Yeo, A. R. and Flowers, T. J. (1983). Vertical Differences in the Toxicity of Sodium Ions in Rice Leaves. *Physiol. Plant.*, **59**: 989-995.

تأثیر تنش شوری بر رشد، میزان یونها و توزیع ماده خشک در ارقام چغندر قند

ع. ر. دادخواه و ح. گریفیتز

چکیده

این تحقیق بمنظور بررسی تفاوت‌های درون گونه ای واریته‌های چغندر قند، در پارامترهای رشد و توزیع ماده خشک تحت تیمارهای مختلف شوری (۰، ۵۰، ۱۵۰، ۲۵۰ و ۳۵۰ میلی مول مخلوط نمک‌های کلرورسیدیم و کلرور کلسیم به نسبت مولی ۵ به یک) با استفاده از چهار واریته، یک واریته انگلیسی (مادیسون) و سه واریته ایرانی (۷۲۳۳-P۱۲، ۷۲۳۳-P۲۱ و ۷۲۳۳-P۲۹) انجام شد. آزمایش در گلخانه تحت شرایط کنترل شده بمدت ۱۸ هفته اجرا گردید. اگر چه پارامترهای رشد از قبیل سطح برگ گیاه و تجمع ماده خشک در سطح پایین شوری (۵۰ میلی مول) در واریته‌های مختلف چندان تحت تأثیر قرار نگرفتند، غلظت‌های بالاتر شوری بطور معنی داری همه صفات رشد را کاهش دادند. در بالاترین سطح شوری (۳۵۰ میلی مول) واریته P۲۹ بطور معنی داری سطح برگ و ماده خشک بیشتری نسبت به سایر واریته‌ها در هفته شانزدهم پس از اعمال شوری داشت. پتانسیل آب برگ (Ψ_1) قبل از طلوع خورشید و محتوی آب اندام‌های هوایی (SWC) با افزایش شوری کاهش یافتند. واریته‌های مادیسون و ۷۲۳۳-P۱۲ در سطوح شوری بالا (۲۵۰ و ۳۵۰ میلی مول) بطور معنی داری Ψ_1 (منفی تر) و SWC کمتری داشتند، در حالیکه P۲۹ و P۲۱ Ψ_1 و SWC بیشتری در همین سطوح شوری داشتند. میزان یونهای سدیم (Na^+) و کلر (Cl^-) در شرایط شوری افزایش یافت، اما واریته P۲۹ کمترین میزان سدیم و کلر را نسبت به سایر واریته‌ها در سطوح شوری بالا داشت. در شانزده هفته پس از اعمال شوری ماده خشک ریشه واریته‌های چغندر قند در پایین‌ترین و بالا ترین سطوح شوری بترتیب ۲۳/۳ و ۸۹/۸ درصد در مقایسه با شاهد کاهش یافت، در حالیکه ماده خشک اندام‌های هوایی در همین سطوح شوری بترتیب ۱/۱ و ۷۷ درصد کاهش نشان دادند. تنش شوری الگوی توزیع ماده خشک به قسمتهای مختلف گیاه را تغییر داد بطوریکه تخصیص ماده خشک به ریشه گیاه در پایین‌ترین (۵۰ میلی مول) و بالاترین (۳۵۰ میلی مول) سطوح شوری پس از شانزده هفته بترتیب ۶ و ۱۸ درصد در مقایسه با شاهد کاهش یافت.