

## ACCEPTED ARTICLE

### **Effects of Aloe vera gel based active coating functionalized with lemon peel essential oil on shelf life and quality attributes of cheese**

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#### **Abstract**

In this study, the effect of an edible aloe vera gel-coating containing lemon peel essential oil (0, 100, and 150 ppm) on the qualitative characteristics of cheese samples was examined. Treatments included 4 groups: control (without coating), aloe vera gel (AVG), AVG + 100 ppm lemon peel essential oil (EO), and AVG + 150 ppm lemon peel EO. These treatments were evaluated for 60 days in terms of physicochemical, textural, sensory, and microbial counting properties. The findings revealed that as storage duration increased, the acidity and salt increased while pH and moisture content decreased. In evaluating the sensory properties, the effect of treatments on all sensory properties except color scores was significant. Samples coated with AVG and 100 ppm lemon peel EO received the highest flavor scores (4.97). As the storage time increased, the hardness, chewiness and springiness of the cheese samples increased. The samples' adhesiveness was not affected by the storage duration. At the end of the storage time, the highest total microbial, mold and yeast counts were associated with the control cheese samples (5.37 and 4.62 log cfu/g, respectively) and the lowest amount was related to the samples coated with AVG and 150 ppm of lemon peel EO (3.92 and 3.76 log cfu/g, respectively). In general, the use of edible coating produced with AVG and lower concentrations of lemon peel EO (100 ppm and less) improved the appearance and the flavor of cheese samples during 60 days of storage.

**Keywords:** Aloe vera, Edible coating, Lemon peel essential oil, Cheese, Sensory evaluation, Textural properties.

#### **Introduction**

Nowadays, chemical preservatives have been proved to be harmful and consumers are more desired to use foods without preservatives or containing natural preservatives (Sambu *et al.*, 2022). Cheese

36 is a nutrient-dense dairy food, providing protein, fats, and minerals (Yerlikaya and Ozer, 2014).  
37 Cheese can be used as a main ingredient in meals, as a dessert and as a component of foods. The  
38 rapid growth of cheese consumption in the world particularly in European countries is due to its  
39 use in various foods (Gomes da Cruz *et al.*, 2009).

40 Due to its nutrients, cheese provides a favorable environment for the growth of several bacteria.  
41 Globally, there are a vast selection of different cheese varieties, and each one has a unique  
42 microbiological profile. The high nutritional value of cheese has led to extensive studies to improve  
43 the quantitative and qualitative properties of this product and the production of more marketable  
44 products (Trmčić *et al.*, 2016). Mold growth during the ripening and manufacturing of cheese is  
45 one of the common issues faced by cheese makers. This problem is also seen for sellers and  
46 consumers of this product during refrigeration. The use of herb materials has been considered for  
47 many years to prevent the growth of various microorganisms and molds. (Sengun *et al.*, 2008).

48 Some natural and edible film-forming materials can be used to preserve foods such as cheese. One  
49 of these natural ingredients is aloe vera gel (AVG). A clear and firm gel is extracted from the inner  
50 parts of the leaves of the aloe vera (*Aloe barbadensis* Miller). AVG is odorless, non-sticky and has  
51 a high absorption strength. More than 98% of aloe vera gel is made up of water, followed by  
52 polysaccharides (pectin, cellulose, hemicellulose, glucomannan, and acemannan), the acemannan  
53 being considered as the main functional component of AVG (Bozzi *et al.*, 2007). Aloe vera is also  
54 an excellent source of antioxidants, Vitamin C, Vitamin A, Vitamin E, Beta-carotene, Folic acid,  
55 Calcium, and Magnesium (Suriati, 2018). Aloe gel has high potential to be used in the food  
56 industry, one of which as an edible coating material (Suriati *et al.*, 2020). This gel is a  
57 polysaccharide coating and can prevent moisture loss of the product. Due to the presence of  
58 different chemicals including aloin, acemannan (Martinez-Romero *et al.*, 2018), anthraquinone,  
59 saponins (Ergun and Satici, 2012) and phenolic compounds such as chatechin hydrate, caffeic acid,  
60 ferulic acid, ellagic acid, and quercetin (Sumi *et al.*, 2019), AVG has antifungal and antimicrobial  
61 characteristics and inhibits the growth and proliferation of fungus.

62 Essential oils (EO) are another herbal component that can be utilized in edible films and coatings.  
63 Citrus peel EO is a mixture of more than 100 compounds, which is divided into two volatile parts  
64 (99-85% of the total essential oil), and the non-volatile part (1-15%). The volatile parts include  
65 monoterpenes (such as limonene) and sesquiterpene hydrocarbons and oxygenated derivatives  
66 [aldehydes (such as citral), ketones and acids (along with linear aldehydes), alcohols (such as

67 linalool)] and esters and the non-volatile parts include hydrocarbons, fatty acids, sterols,  
68 carotenoids, waxes, coumarins, and flavonoids (Bennici and Tani, 2004). Limonene is the main  
69 monoterpene compound of citrus essential oil and has antioxidant, antibacterial and antiviral  
70 properties (Espina *et al.*, 2011; Roy *et al.*, 2007). In general, Citrus peel EOs have potent  
71 antioxidant and antibacterial properties (Raspo *et al.*, 2020).

72 In a reported study, Shenbagam *et al.* (2023) investigated the effects of aloe vera gel-based edible  
73 coating (with or without incorporation of orange peel essential oil) on the postharvest shelf life and  
74 qualitative properties of button mushroom. The results showed that maximum concentration of  
75 orange peel essential oil (1500 µL/L) incorporated in the 50% aloe vera gel significantly improved  
76 the postharvest quality attributes of mushrooms and helped extend the shelf life of mushrooms up  
77 to 4 days as compared to the control.

78 This study's objective was to determine the effect of an edible coating made of AVG and various  
79 concentrations of lemon peel EO on physicochemical, sensory and textural properties as well as  
80 microbial profile (total microbial count and total mold and yeast counts) of cheese.

81

## 82 **Methods**

### 83 **Preparation of lemon peel essential oil**

84 20 kg of Mexican lemon peel (*Citrus aurantifolia*) was dried at an ambient temperature (25-38°C)  
85 and in the shade. The dried lemon peel was grounded and passed through a sieve (mesh 40). The  
86 EO was extracted by steam distillation over a Clevenger system (Aria Exir, Iran) for 4h. The  
87 obtained EO was dehydrated using sodium sulfate and stored at 4°C (Chanthaphon *et al.*, 2018).  
88 Lemon peel EO was yellow in color and yield of extracted EOs was 1.1% (w/w).

89

### 90 **Preparation of AVG**

91 Aloe vera leaves were collected from University of Jiroft Research Farm. The leaves were washed  
92 and their jagged edges were cut with a knife. The top layer of the leaf was removed lengthwise and  
93 the gel was carefully separated from the leaf. The gel parts were blended thoroughly and put  
94 through a clean metal sieve (mesh 20) to form a homogeneous solution and the extract was finally  
95 pasteurized at 65°C for 15 min (Martinez-Romero *et al.*, 2018). In this study, Aloe vera extract at  
96 a 100% concentration was used.

97

### 98 **Coating formulations and application**

99 The cheese samples were prepared in Kerman Pegah milk factory. To produce cheese, milk was  
100 pasteurized after fat standardization (3.5%) by HTST method and then concentrated at 50°C in  
101 ultra-filtration system until reaching 34% dry matter. Starter inoculation was done at 32-35°C.  
102 Then, rennet (12 ml per 400 g) was added and mixed well. The mixture was poured into containers  
103 and after passing through the coagulation tunnel (30°C for 20 min), salt (3%) was added. After  
104 that, it was sealed and placed in an incubator at 28°C until reaching a pH of 4.7. Then, it was  
105 transferred to the cold room and kept at 4°C until the experiments (Khani and Roufegari Nejad,  
106 2018). The cheese samples were cut into cubic specimens (3 ×3×3 cm<sup>3</sup>) and coated by immersion  
107 method. During this step, the cheese samples were immersed in the coating mixture (AVG with  
108 various concentrations of lemon peel EO (0, 100 and 150 ppm which was homogenized by a  
109 homogenizer at 1000 rpm) for 1 minute. The samples were incubated for about 8 hours under  
110 controlled temperature (12°C) and humidity (relative humidity of 85%) to dry all coatings  
111 (Henriques *et al.*, 2013). AVG and lemon peel EO created a colorless coating on the samples. The  
112 samples were then placed in sealed polypropylene containers and stored in the refrigerator (4°C)  
113 and evaluated at 15 days of intervals throughout 60 days of storage period.

114

## 115 **Experimental methodology**

### 116 **Determination of acidity**

117 The acidity of cheese samples was determined in terms of lactic acid and by titration with sodium  
118 hydroxide (0.1 N) using the equation 1 (Iranian International Standard No2852:1995).

119 Equation 1:  $\text{Acidity}\% = N \cdot 0.009 \cdot 100/M$

120 In this equation, N is the amount of sodium hydroxide 0.1 N consumed (ml) and M is the weight  
121 of the sample.

122

### 123 **pH measurement**

124 A digital pH meter (Metrohm, model 827, Switzerland) was used to determine the samples' pH  
125 levels (Iranian International Standard No2852:1995).

126

### 127 **Measurement of moisture content**

128 Cheese samples were placed in an oven at 102°C until they reached a constant weight (about 5 h).

129 The dried samples were weighed after cooling, and the amount of moisture loss was estimated  
130 using the equation 2 (Roy *et al.*, 2007):

131 Equation 2: Moisture loss rate= Weight before drying - Weight after drying / Weight before drying  
132 × 100

### 133 134 **Measurement of salt content**

135 Mohr method was used to determine the amount of salt. Titration was performed using silver  
136 nitrate solution (0.1 N) until an orange precipitate appeared. The percentage of salt was calculated  
137 as equation 3 (Dorosti *et al.*, 2011).

138 Equation 3: Amount of salt% = ml of consumed silver nitrate (ml) × silver nitrate N × 0.585

139

### 140 **Sensory evaluation**

141 Sensory properties of cheese samples were evaluated using a five-point hedonic test (very bad: 1  
142 to very good: 5) in the first and sixtieth days of storage. The evaluators were 50 people who were  
143 selected from the experts working in Pegah Kerman factory and students familiar with the  
144 characteristics of cheese. Samples (100 g packages) were removed from the refrigerator before the  
145 test and after reaching the ambient temperature in 30 g pieces were given to the evaluators.

146 Samples were assessed for their characteristics including flavor, odor, color, texture and overall  
147 acceptance. Mean data of the first and sixtieth days were reported (Beigomi *et al.*, 2013).

148

### 149 **Texture analysis test**

150 A texture analyzer equipment (model QTS25, FARNEL CNS, UK) and a cylindrical probe with a  
151 diameter of 36 mm were utilized for the texture profile analysis (TPA) test. The cheese samples  
152 were removed from the refrigerator before the test and after slicing (20 × 20 × 20 mm) up to 50%  
153 of the initial height (10 mm depth) was compressed by the machine. Each test was performed in at  
154 least three replications. The measured traits were: hardness, cohesiveness, adhesiveness,  
155 chewiness, springiness, and gumminess. It should be noted that the TPA test is a two-step test and  
156 these traits were defined according to the standard TPA curve (Hosseini *et al.*, 2013).

157

### 158 **Microbial tests**

#### 159 **1- Total microbial count**

160 The total microbial count was performed using a PCA (Plate Count Agar) at 37°C for 48 h. The  
161 number of bacteria in cheese samples was calculated as follows (Rezaei *et al.*, 2010).

162 Microbial content /g of cheese = number of colonies × inverse dilution coefficient × 10

163  
 164 **Mold and yeast count**  
 165 YGC (Yeast Extract Glucose Chloramphenicol) medium was used for mold and yeast (fungi) count  
 166 at 25°C for 48-72 h. After incubation, the obtained colonies were counted using the equation of the  
 167 previous section (Rezaei *et al.*, 2010).

168  
 169 **Statistical analysis**

170 The experiments were conducted in a factorial experiment based on completely randomized design  
 171 and the experimental data were analyzed with SPSS: 21 software. Factors included treatments (4  
 172 levels) and storage time (5 levels). The means were compared using the Duncan's multiple range  
 173 test with a 5% confidence level. All experiments were carried out in triplicate.

174  
 175 **Results and Discussion**

176 **The effect of treatments on the pH**

177 The findings in Table 1 demonstrate that the pH of cheese samples significantly reduced as storage  
 178 time was increased. The lowest pH reduction was observed for cheese samples coated with AVG  
 179 and 150 ppm of lemon peel EO. The pH of control treatment was found to be the lowest at the end  
 180 of the maintenance time, whereas the other treatments were not significantly different ( $p > 0.05$ ).

181  
 182 **Table 1.** The effect of treatments on the pH of samples.

Treatments	Day 1	Day 15	Day 30	Day 45	Day 60
Control	5.50±0.1 <sup>Ab</sup>	5.43±0.05 <sup>ABb</sup>	5.28±0.12 <sup>Cb</sup>	5.09±0.1 <sup>CDb</sup>	4.86±0.14 <sup>Ec</sup>
AVG	5.86±0.09 <sup>Aa</sup>	5.79±0.1 <sup>Aba</sup>	5.57±0.05 <sup>Ca</sup>	5.36±0.1 <sup>Da</sup>	5.10±0.06 <sup>Eab</sup>
AVG + 100 ppm EO	5.85±0.11 <sup>Aa</sup>	5.75±0.15 <sup>Aba</sup>	5.58±0.2 <sup>Ca</sup>	5.35±0.08 <sup>Da</sup>	5.16±0.1 <sup>Eab</sup>
AVG + 150 ppm EO	5.83±0.1 <sup>Aa</sup>	5.75±0.08 <sup>Aba</sup>	5.57±0.05 <sup>Ca</sup>	5.57±0.05 <sup>CDa</sup>	5.25±0.12 <sup>Da</sup>
P value	0.013	0.004	0.003	0.004	0.049

183 \* Mean values in each column that have different lower-case letters have a significant difference ( $p < 0.05$ ).

184 \* Numbers in each row that have different capital letters have a significant difference ( $p < 0.05$ ).

185  
 186 **The effect of the treatments on the acidity**

187 Table 2 shows that the acidity of the cheese samples was significantly influenced by the type of  
 188 coating used as well as the storage time. The treatment coated with AVG and 150 ppm EO and  
 189 control had the greatest and lowest acidity, respectively, on the sixtieth day. The acidity of the  
 190 treatments increased as storage duration increased, and this increase was significant in all

191 investigated treatments on all storage days. In cheese samples coated with AVG and 150 ppm of  
 192 lemon peel EO, minimal acidity changes were seen at the end of storage period.

193 **Table 2.** The effect of treatments on the acidity of samples.

Treatments	Day 1	Day 15	Day 30	Day 45	Day 60
Control	1.46±0.05 <sup>Ea</sup>	1.77±0.09 <sup>Da</sup>	2.11±0.2 <sup>Ca</sup>	2.66±0.09 <sup>Ba</sup>	3.26±0.1 <sup>Aa</sup>
AVG	1.26±0.1 <sup>Eb</sup>	1.56±0.1 <sup>Db</sup>	1.80±0.18 <sup>Cb</sup>	2.16±0.1 <sup>Bb</sup>	2.70±0.05 <sup>Ab</sup>
AVG + 100 ppm EO	1.23±0.08 <sup>Eb</sup>	1.55±0.1 <sup>Db</sup>	1.81±0.1 <sup>Cb</sup>	2.05±0.06 <sup>Bbc</sup>	2.66±0.11 <sup>Abc</sup>
AVG + 150 ppm EO	1.26±0.08 <sup>Eb</sup>	1.50±0.1 <sup>CDb</sup>	1.67±0.15 <sup>Cb</sup>	1.93±0.1 <sup>Bc</sup>	2.52±0.15 <sup>Ac</sup>
P value	0.004	0.004	0.033	0.044	0.004

194 \* Mean values in each column that have different lower-case letters have a significant difference ( $p < 0.05$ ).

195 \* Numbers in each row that have different capital letters have a significant difference ( $p < 0.05$ ).

196  
 197 By increasing the storage time, a reduction in pH values of all samples were observed which may  
 198 be related to the activity of lactic acid bacteria species owing to the metabolization of lactose to  
 199 lactate and produce acid (Dermiki *et al.*, 2008). Ramos *et al.* (2012) found that the pH of cheeses  
 200 coated with whey protein isolate, guar gum and antimicrobial substances decreased with increasing  
 201 storage time, and the coated cheeses had a higher pH than the control. Jamshidi *et al.* (2018), used  
 202 a coating of AVG and Persian gum in Iranian white cheese, and reported that during storage, the  
 203 pH decreased significantly while the acidity increased. Over time, the acidity of the various  
 204 treatments increased, indicating that an increase in lactic acid production by the bacteria may be  
 205 the main reason for this trend, which is definitely consistent with the decreasing trend observed in  
 206 pH during storage.

207 El-Sisi *et al.* (2015) showed that the acidity of chitosan-coated cheeses increased during storage.  
 208 A study also revealed that the acidity of cheddar cheese samples coated with whey protein  
 209 increased during ripening (Wagh *et al.*, 2013).

210 On the sixtieth day, the lowest amount of acidity was observed in cheese samples coated with AVG  
 211 and lemon peel EO, which could indicate the lowest bioavailability of lactic acid bacteria (starter  
 212 and non-starter) in these samples; Because more activity of lactic acid bacteria leads to more  
 213 decomposition of lactate and production of organic acids such as lactic acid and acetic acid, and  
 214 AVG and lemon peel EO probably due to antimicrobial activity decreased growth of these bacteria  
 215 in cheese samples (Wagh *et al.*, 2013).

216  
 217 **The effect of treatments on the salt content**

218 According to results of Table 3, the control had the most salt content at all storage times, while the  
 219 other treatments were not significantly different ( $p > 0.05$ ). The salt content of the treatments  
 220 enhanced with increasing storage period, although this rise was not significant in samples coated  
 221 with AVG and 150 ppm lemon peel EO ( $p > 0.05$ ).

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 232

**Table 3.** The effect of treatments on the salt content (%) of cheese samples.

Treatments	Day 1	Day 15	Day 30	Day 45	Day 60
Control	3.26±0.25 <sup>Ba</sup>	3.30±0.2 <sup>ABa</sup>	3.41±0.1 <sup>ABa</sup>	3.50±0.2 <sup>Aa</sup>	3.57±0.1 <sup>Aa</sup>
AVG	2.76±0.1 <sup>Bb</sup>	2.81±0.5 <sup>ABb</sup>	2.86±0.25 <sup>Ab</sup>	2.96±0.1 <sup>Ab</sup>	3.06±0.15 <sup>Ab</sup>
AVG + 100 ppm EO	2.73±0.3 <sup>Bb</sup>	2.80±0.5 <sup>ABb</sup>	2.86±0.21 <sup>Ab</sup>	2.93±0.09 <sup>Ab</sup>	3.07±0.15 <sup>Abc</sup>
AVG + 150 ppm EO	2.73±0.17 <sup>ABb</sup>	2.80±0.44 <sup>ABb</sup>	2.87±0.23 <sup>Ab</sup>	2.94±0.15 <sup>Ab</sup>	2.98±0.17 <sup>Ac</sup>
P value	0.007	0.002	0.001	0.001	0.001

233 \* Mean values in each column that have different lower-case letters have a significant difference ( $p < 0.05$ ).  
 234 \* Numbers in each row that have different capital letters have a significant difference ( $p < 0.05$ ).

235  
 236 It can be seen that all the coated samples have less salt than control, which is consistent with the  
 237 results of other researchers who have studied the effect of coating on the properties of cheese  
 238 (Ramos *et al.*, 2012; Yilmaz and Dagdemir, 2012). On the other hand, during the 60 days of storage,  
 239 the salt content of samples increased slightly as a result of weight loss due to the removal of  
 240 moisture from the cheese texture.

241  
 242 **The effect of treatments on the moisture content**

243 The effect of the treatments on the moisture content of cheese samples is shown in Table 4. The  
 244 results reveal that the coating treatments and storage period had a significant effect on the moisture  
 245 content. As storage time increased, the moisture content of samples decreased. Cheese samples  
 246 with coatings retained moisture significantly more than the control. There was no significant  
 247 difference in the moisture content of all treatments on the first day of storage ( $p > 0.05$ ).

248  
 249 **Table 4.** The effect of treatments on the moisture content (%) of samples.

Treatments	Day 1	Day 15	Day 30	Day 45	Day 60
------------	-------	--------	--------	--------	--------



Control	63.30±4.2 <sup>Aa</sup>	61.73±2.9 <sup>Ab</sup>	60.76±2.1 <sup>ABb</sup>	58.66±3.3 <sup>Bb</sup>	58.73±4.1 <sup>Bb</sup>
AVG	64.15±3.1 <sup>Aa</sup>	64.66±4.3 <sup>Aa</sup>	62.70±2.5 <sup>ABab</sup>	61.67±3.1 <sup>Ba</sup>	61.33±4.2 <sup>Ba</sup>
AVG + 100 ppm EO	64.60±4.0 <sup>Aa</sup>	64.73±4.2 <sup>Aa</sup>	63.00±2.1 <sup>Aa</sup>	62.08±3.5 <sup>ABa</sup>	61.40±3.8 <sup>Ba</sup>
AVG + 150 ppm EO	64.20±4.1 <sup>Aa</sup>	64.84±3.9 <sup>Aa</sup>	63.43±2.4 <sup>ABa</sup>	62.64±3.4 <sup>Ba</sup>	61.44±4.3 <sup>Ba</sup>
P value	0.006	0.032	0.007	0.000	0.033

250 \* Mean values in each column that have different lower-case letters have a significant difference ( $p < 0.05$ ).

251 \* Numbers in each row that have different capital letters have a significant difference ( $p < 0.05$ ).

252  
253 The cheese samples' moisture gradually decreased during the storage period as a result of some  
254 moisture being released from the texture of the cheese and the packaging to the outside. The  
255 difference between the coated samples is probably due to the composition of the coating as well as  
256 the kinetics of water influence and outflow into the various coatings (Pantaleão *et al.*, 2007).  
257 Jamshidi *et al.* (2018) reported that almost all cheeses coated with AVG and Persian gum showed  
258 higher moisture content than the control, which indicates the positive effect of coating on moisture  
259 retention in cheese during storage. Coating with aloe vera gel had a barrier property for moisture  
260 loss in several fruits such as peach (Mohammadi *et al.*, 2020), plum (Martinez-Romero *et al.*, 2018),  
261 grapes, fresh cut papaya (Farina *et al.*, 2020), and tomato fruit (Tzortzakis *et al.*, 2019).

### 262 263 **The effect of treatments on the sensory properties**

264 Table 5 shows that with the exception of the color index, the effects of the tested treatments on the  
265 sensory characteristics of samples are significant. The highest taste score was related to the  
266 treatment coated with AVG and 100 ppm of lemon peel EO + the lowest taste score was related to  
267 the control and the AVG and 150 ppm of lemon peel EO treatments. The highest and lowest odor  
268 scores were observed in AVG with 100 ppm of lemon peel EO treatment and control, respectively.  
269 Samples coated with AVG and different concentrations of EO did not show significant differences  
270 in terms of texture ( $p > 0.05$ ) and the lowest texture score was assigned to the control. In terms of  
271 general acceptance, AVG with 100 ppm of lemon peel EO treatment received the highest score.

272  
273 **Table 5.** The effect of treatments on the sensory properties of samples.

Treatments	Taste	Odor	Color	Texture	General acceptance
Control	4.36±0.1 <sup>c</sup>	4.45±0.05 <sup>c</sup>	4.92±0.1	4.53±0.15 <sup>c</sup>	4.42±0.08 <sup>cd</sup>
AVG	4.59±0.05 <sup>b</sup>	4.63±0.1 <sup>b</sup>	4.96±0.1	4.67±0.1 <sup>b</sup>	4.78±0.12 <sup>b</sup>
AVG + 100 ppm EO	4.97±0.06 <sup>a</sup>	4.89±0.12 <sup>a</sup>	4.96±0.15	4.91±0.05 <sup>Aa</sup>	4.95±0.06 <sup>a</sup>
AVG + 150 ppm EO	4.36±0.05 <sup>c</sup>	4.75±0.16 <sup>b</sup>	4.97±0.09	4.89±0.12 <sup>a</sup>	4.54±0.1 <sup>c</sup>
P value	0.001	0.001	0.56	0.000	0.003

274 \* Mean values in each column that have different lower-case letters have a significant difference ( $p < 0.05$ ).

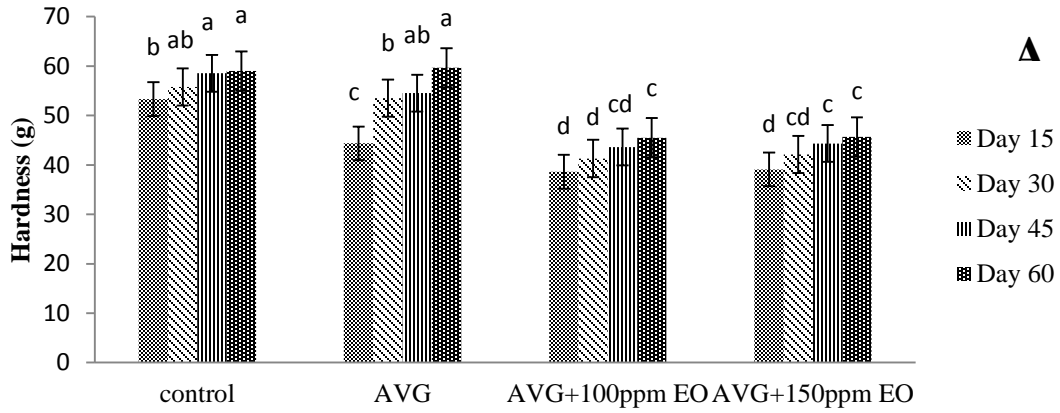
275  
276 Most sensory panelists reported a bitter taste for cheeses containing 150 ppm of lemon peel  
277 essential oil. According to research of Yilmaz and Dagdemir (2012), there were no significant  
278 differences in the color of cheese samples coated in beeswax compared to the control, which is  
279 consistent with the findings of this investigation.

280 Abbas *et al.* (2017) reported that adding 0.005 and 0.010  $\mu\text{l}$  of basil essential oil to UF soft cheese  
281 significantly improved the taste throughout the freshness of cheese and during the 60 days of  
282 storage time. According to this report, the desirability of samples containing low concentration  
283 (0.005  $\mu\text{l}$  per 100 ml) was higher than the samples containing high concentration (0.010  $\mu\text{l}$  / 100  
284 ml). Mohammadi *et al.* (2011) reported that 100 mg/kg of basil essential oil improved the odor,  
285 taste and acceptability of white cheese during the production and storage, however, the taste and  
286 acceptability of the cheese samples were adversely affected by the essential oil concentrations of  
287 150 and 200 mg/Kg.

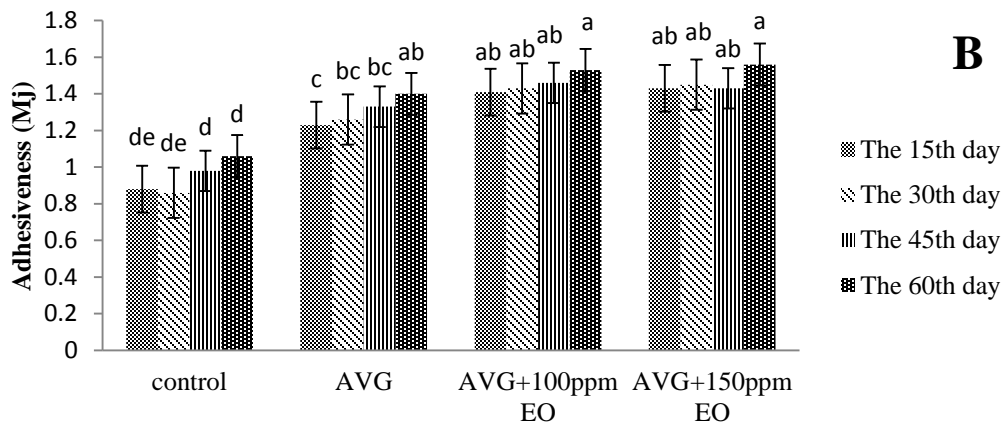
288 According to Otero *et al.* (2014), sheep cheese samples covered with edible films containing  
289 antimicrobial agents had improved sensory properties. The results of Pieretti *et al.* (2019) showed  
290 that cheese samples coated with alginate and low concentrations of oregano essential oil had better  
291 sensory acceptance than the control and higher concentrations of essential oil.

292  
293 **The effect of the treatments on the textural characteristics**  
294 The effect of the studied treatments on the textural characteristics of cheese samples is shown in  
295 Figure 1 (A-F).

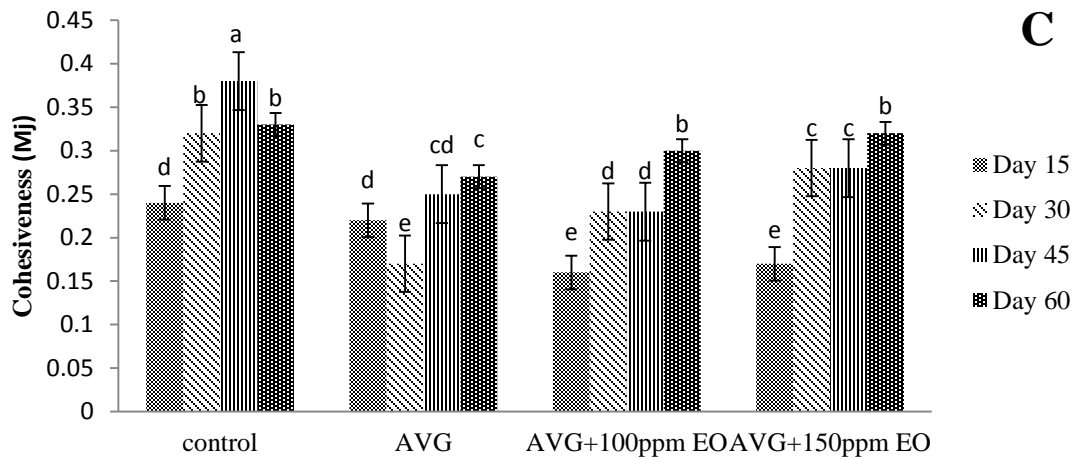
296  
297 **1- Hardness**  
298 According to Fig. 1(A), the coating and storage duration both significantly affected hardness. The  
299 hardness of the samples increased with storage time. On 60<sup>th</sup> day, the AVG with 100 and 150 ppm  
300 of lemon peel EO had the lowest hardness while the control and AVG treatments had the highest  
301 hardness.



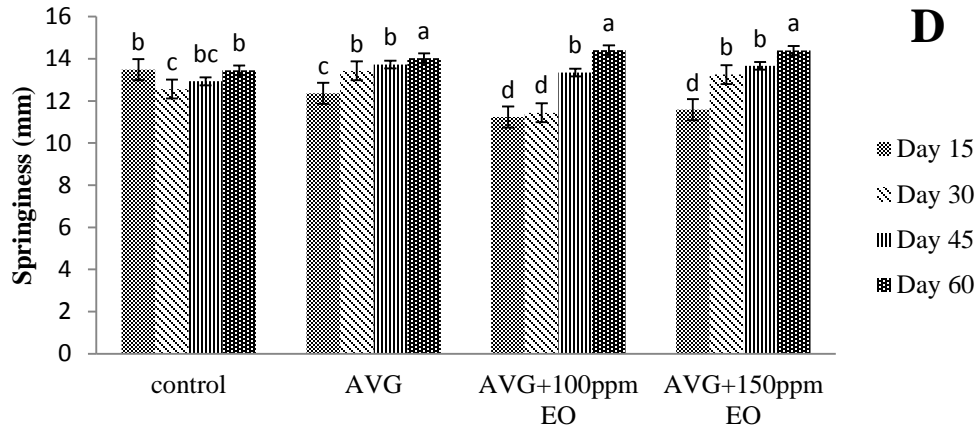
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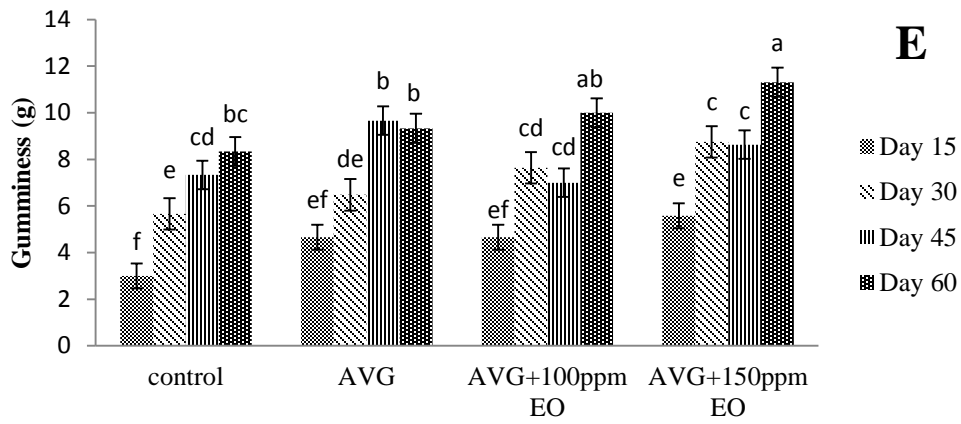
303



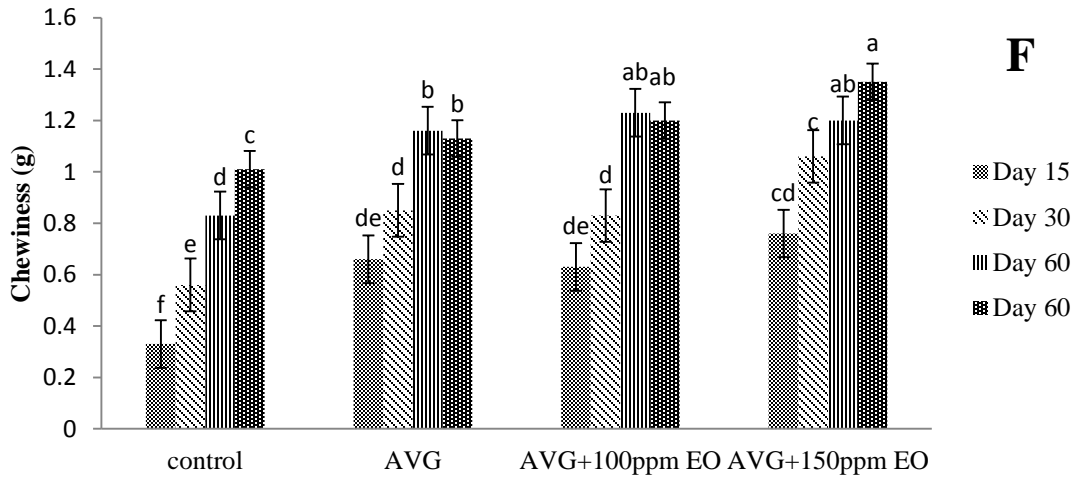
304



305



306



307

308 **Figure 1. The effect of treatments on the hardness (A), Adhesiveness (B), cohesiveness (C),**  
 309 **springiness (D), gumminess (E), and chewiness (F) of samples.**

310

311

312 **2- Adhesiveness**

313 Fig. 1(B), shows that although numerically the adhesiveness of the samples increased during  
314 storage, the storage time had no significant effect on the adhesiveness of samples. At the end of  
315 storage, samples coated with AVG and 150 ppm of lemon peel EO showed the highest  
316 adhesiveness, which did not show a significant difference with the AVG and 100 ppm of lemon  
317 peel EO treatment ( $p > 0.05$ ). The lowest adhesiveness was related to the control.

318  
319 **3- Cohesiveness**

320 Fig. 1 (C), shows that the AVG treatment had the lowest cohesiveness at the end of the storage  
321 period, with no other treatments significantly different ( $p > 0.05$ ).

322  
323 **4- Springiness**

324 According to Fig. 1(D), No particular trend in the springiness of samples during storage time was  
325 seen in the control. In other treatments, springiness of samples increased with increasing storage  
326 time. On the sixtieth day, the highest springiness was related to AVG treatments with 100 and 150  
327 ppm of lemon peel EO and the lowest amount of springiness was related to control.

328  
329 **5-Gumminess**

330 According to Fig. 1 (E), it can be seen that at the end of the storage, the highest and lowest  
331 gumminess were observed in the treatment coated with AVG+150 ppm lemon peel EO and the  
332 control, respectively. The gumminess of samples significantly increased as the storage time rose.

333  
334 **6- Chewiness**

335 Fig. 1(E) shows that the chewiness of samples increased as storage time increased. In the samples  
336 coated with AVG and AVG containing 100 and 150 ppm of lemon peel EO on the forty-fifth and  
337 sixtieth days, this enhancement was not significant ( $p > 0.05$ ). On the sixtieth day, the lowest  
338 amount of chewiness was observed in the control and samples coated with AVG and the highest  
339 amount of chewiness was observed in the samples coated with AVG and 150 ppm of lemon peel  
340 EO.

341 According to the findings of the textural characteristics, the hardness of the samples increased with  
342 increasing storage time, which may be related to moisture loss during storage. Another factor  
343 contributing to the samples' increased hardness during storage is likely an increase in protein-  
344 protein interactions (Bianchi *et al.*, 2021). It was also observed that the coated samples had less

345 hardness than the control. It seems that more moisture in the coatings and more hydration may  
346 reduce the hardness of the samples (Zhong *et al.*, 2014).

347 Pieretti *et al.* (2019) examined how rosemary and oregano EOs and alginate-based edible coatings  
348 affected the textural characteristics of fresh cheese, and they found that at the end of the storage  
349 period, the coated samples had less hardness than the control.

350 At the end of the storage time, the highest amount of adhesiveness was observed in the samples  
351 coated with AVG and 150 ppm lemon peel EO and the lowest amount of adhesiveness was related  
352 to the control. In the research of Wang *et al.* (2019), cheddar cheese samples coated with isolated  
353 whey protein nanofibrils and carvacrol showed more adhesiveness than uncoated samples.

354 The cohesiveness of the samples increased with increasing the storage time, and at the end of the  
355 storage, the treatment with the lowest cohesiveness was in the presence of AVG; the other  
356 treatments did not significantly differ from each other. In the study reported by Wang *et al.* (2019),  
357 the cohesiveness of coated cheese samples increased with increasing storage time, while no  
358 significant difference was observed in the other samples.

359 With increasing storage period, the chewiness of samples increased. This is in line with the  
360 hardness and gumminess properties. On the sixtieth day, the highest amount of chewiness was  
361 related to the treatment coated with AVG and 150 ppm of lemon peel EO. From a sensory point of  
362 view, it is perceived that more energy is needed to chew the coated samples. It was found that the  
363 chewiness of cheese samples coated with starch and carvacrol increased with increasing storage,  
364 and the coated samples had more chewiness than the control (López-Córdoba, 2021).

365

## 366 **The effect of treatments on microbial count of samples during storage**

### 367 **1- The total microbial count**

368 Table 6 shows the effect of the treatments on the total microbial count of the samples. This table  
369 shows that the total microbial count was significantly affected by both storage times and coatings.  
370 The total microbial count increased with increasing storage time. The sample coated with AVG  
371 and 150 ppm of lemon peel EO had the lowest microbial count. In general, the coated treatments  
372 showed less microbial count than the control.

373

374 **Table 6.** The effect of treatments on the total microbial count (log cfu/g) of samples.

Treatments	Day 1	Day 15	Day 30	Day 45	Day 60
Control	4.87±0.15 <sup>Ca</sup>	4.88±0.1 <sup>Ca</sup>	4.94±0.21 <sup>Ba</sup>	4.98±0.15 <sup>Ba</sup>	5.37±0.2 <sup>Aa</sup>
AVG	3.64±0.1 <sup>Db</sup>	3.73±0.14 <sup>Cb</sup>	4.60±0.15 <sup>Bb</sup>	4.64±0.11 <sup>Bb</sup>	4.82±0.1 <sup>Ab</sup>

AVG + 100 ppm EO	3.38±0.22 <sup>Dc</sup>	3.55±0.21 <sup>Cc</sup>	3.96±0.25 <sup>Bc</sup>	4.30±0.1 <sup>Ac</sup>	4.31±0.14 <sup>Ac</sup>
AVG + 150 ppm EO	2.71±0.12 <sup>Ed</sup>	2.92±0.2 <sup>Dd</sup>	3.73±0.1 <sup>Cd</sup>	3.81±0.25 <sup>Bd</sup>	3.92±0.21 <sup>Ac</sup>
P value	0.001	0.002	0.001	0.000	0.003

375 \* Mean values in each column that have different lower-case letters have a significant difference ( $p < 0.05$ ).

376 \* Numbers in each row that have different capital letters have a significant difference ( $p < 0.05$ ).

377

## 378 2- The total mold and yeast count

379 Table 7 shows that there is significant variation in the total number of mold and yeast in cheese  
 380 samples depending on the various treatments and storage time. The total amount of mold and yeast  
 381 increased with more storage time across all treatments, with the control having the highest levels.  
 382 The lowest amounts of mold and yeast were found in samples that had been coated with AVG and  
 383 EO.

384

385 **Table 7.** The effect of treatments on total mold and yeast count (log cfu/g) of cheese samples.

Treatments	Day 1	Day 15	Day 30	Day 45	Day 60
Control	3.34±0.2 <sup>Ea</sup>	3.82±0.13 <sup>Da</sup>	3.96±0.15 <sup>Ca</sup>	4.13±0.14 <sup>Ba</sup>	4.62±0.1 <sup>Aa</sup>
AVG	3.17±0.15 <sup>Eb</sup>	3.45±0.21 <sup>Db</sup>	3.81±0.11 <sup>Cb</sup>	4.01±0.1 <sup>Bb</sup>	4.15±0.2 <sup>Ab</sup>
AVG + 100 ppm EO	0.00 <sup>Dc</sup>	3.11±0.1 <sup>Cc</sup>	3.47±0.22 <sup>Bc</sup>	3.76±0.1 <sup>Ac</sup>	3.92±0.21 <sup>Ac</sup>
AVG + 150 ppm EO	0.00 <sup>Ec</sup>	0.00 <sup>Dd</sup>	3.06±0.1 <sup>Cd</sup>	3.47±0.12 <sup>Bd</sup>	3.76±0.15 <sup>Ad</sup>
P value	0.001	0.002	0.003	0.001	0.002

386 \* Mean values in each column that have different lower-case letters have a significant difference ( $p < 0.05$ ).

387 \* Numbers in each row that have different capital letters have a significant difference ( $p < 0.05$ ).

388

389 In general, the coated treatments showed less microbial, mold and yeast counts than the control.  
 390 Numerous studies have focused on the antibacterial effects of AVG and lemon peel EO (Nielsen  
 391 and Rios, 2000; Irshad *et al.*, 2011; Roy *et al.*, 2007).

392 Aloin and aloe-emodin are the two main components of aloe vera gel. Several researchers have  
 393 confirmed the antifungal and anti-bacterial properties with improved moisture and gas barrier  
 394 properties of aloe vera gel based edible coating (Ortega-Toro *et al.*, 2017)

395 AVG as a coating can create a physical barrier against microorganisms and reduce the occurrence  
 396 of microbial spoilage (Asghari and Khalili, 2014). AVG inhibits the germination and growth of  
 397 fungal mycelium and the inhibitory effect of its compounds on the activity of enzymes of  
 398 pathogenic fungi has been proven (Reynolds and Dweck, 1999). Saritha *et al.* (2010) reported that  
 399 the antimicrobial activity of AVG against gram-positive bacteria was higher than gram-negative  
 400 bacteria. Navarro *et al.* (2011) also reported that AVG controls the *Rhizopus stolonifer*, *Botrytis*  
 401 *cinerea* and *Penicillium digitatum*. Leitgeb *et al.* (2021) investigated the effect of two aloe vera

402 cultivars gel on different bacteria and fungi and reported that both aloe vera cultivars gel inhibited  
403 the growth of *Bacillus cereus*, *Staphylococcus aureus*, *E. coli*, *Pseudomonas aeruginosa*, *P.*  
404 *fluorescens* and *Candida albicans*, representatives of Gram-positive bacteria, Gram-negative  
405 bacteria, and fungi. The antibacterial properties of aloe vera are due to its constituents, which  
406 include saponins, acemannan, and anthraquinone derivatives. Therefore, the presence of these  
407 substances and antibacterial compounds in the AVG can reduce spread of germs in the treated  
408 samples (Ramasubramanian *et al.*, 2010).

409 Essential oils have different mechanisms in destroying microorganisms. These compounds enter  
410 the lipids of cell membranes and mitochondria, and this causes a difference in the structure of cells  
411 and their greater permeability, resulting in the release of ions and other cell contents. The release  
412 of large amounts of cellular contents or the release of vital molecules and ions causes cell death  
413 (Pauli, 2006). There are several reports about the antimicrobial effect of citrus EOs and extracts  
414 (Chanthaphon *et al.*, 2018; Tan *et al.*, 2011). Antimicrobial properties of lemon peel EO are related  
415 to its active ingredients. Limonene is the main monoterpene compound of lemon peel and other  
416 citrus EOs, which has antibacterial and antiviral properties (Espina *et al.*, 2011; Roy *et al.*, 2007).  
417 Artiga-Artigas *et al.* (2017) studied the antimicrobial effect of edible coating containing different  
418 concentrations of oregano EO on low-fat cheese. Their results showed that coatings containing  
419 oregano EO significantly reduced the microbial population during storage.

## 420 421 **Conclusion**

422 The use of edible coating of AVG and lemon peel EO on cheese improved the appearance and  
423 prevented textural changes during storage. Lower concentrations (50 and 100 ppm) of lemon peel  
424 EO were suitable for obtaining cheeses with better sensory properties. The coatings maintained  
425 properties such as moisture, pH, hardness, etc. The lowest microbial, mold and yeast counts were  
426 observed in the treatments coated with AVG and 150 ppm of lemon peel EO. In general, the coated  
427 treatments showed less microbial, mold and yeast counts than the control.

## 428 429 **References**

- 430 1- Abbas, H.M., Assem, F.M., Zaky, W.M., Kassem, J.M. and Omer, E.A. 2017. Antioxidant,  
431 rheological and sensorial properties of ultra-filtrated soft cheese supplemented with basil  
432 essential oil. *Int. J. Dairy Sci.*, **12(5)**: 301-309.
- 433 2- Artigas, M.A., Acevedo-Fani, A. and Martín-Belloso, O. 2017. Improving the Shelf Life of



- 434 Low-Fat Cut Cheese using Nanoemulsion-Based Edible Coatings Containing Oregano  
435 Essential Oil and Mandarin Fiber. *Food Control.*, **76**: 1–12.
- 436 3- Asghari, M.R. and Khalili, H. 2014. The effect of *aloe vera* gel on polyphenol oxidase  
437 activity, qualitative properties and shelf life of cherry fruit. *J. Horticl. Sci.*, **28** (3): 399-406.
- 438 4- Beigomi, M., Ghods Rohani, M., Mohammadifar, M.A., Hashemi, M., valizadeh, M. and  
439 Ghanati, K. 2013. Comparison of textural and sensory characteristics of ultrafiltrated white  
440 cheese produced by paneer bad (*Withania coagulans*) protease and fungal rennet. *Iran. J.*  
441 *Nutr. Sci. Food Technol.*, **8**(1): 253-262.
- 442 5- Bennici, A. and Tani, C. 2004. Anatomical and ultrastructural study of the secretory cavity  
443 development of *Citrus sinensis* and *Citrus limon*: evaluation of schizolysigenous ontogeny.  
444 *Flora.*, **199**: 464 -475.
- 445 6- Bianchi, A., Mallmann, S., Gazoni, I., Cavalheiro, D. and Rigo, E. 2021. Effect of Acid  
446 Casein Freezing on the Industrial Production of Processed Cheese. *Int. Dairy J.*, **118**:  
447 105043
- 448 7- Bozzi, A., Perrin, C., Austin, S. and Arce Vera, F. 2007. Quality and authenticity of  
449 commercial *aloe vera* gel powders. *Food Chem.*, **103**(1): 22–30.
- 450 8- Chanthaphon, S., Chanthachum, S. and Hongpattarakere, T. 2018. Antimicrobial activities  
451 of essential oils and crude extract from tropical citrus spp. against food-related  
452 microorganism. *J Sci Technol.*, **30** (1): 125-131.
- 453 9- Dermiki, M., Ntzimani, A., Badeka, A., Savvaidis, I.N. and Kontominas, M.G. 2008. Shelf-  
454 life extension and quality attributes of the whey cheese. *LWT-Food Sci. Techol.*, **41**(2): 284-  
455 294.
- 456 10- Dorosti, S., Bazmi, A., Ghanbarzadeh, B. and Ayaseh, A. 2011. Effect of brine  
457 concentration on the physicochemical properties of Iranian White cheese. *J. Food Sci.*  
458 *Technol.*, **8**(30): 1-10.
- 459 11- El-Sisi, A.S., Mohamed Gapr, E.S. and Kamaly, K.M. 2015. Use of Chitosan as an Edible  
460 Coating in RAS Cheese. *Biolife.*, **3**(2): 564-570.
- 461 12- Espina, L., Somolinos, M., Loran, S., Conchello, P., Garcia, D. and Pagan, R. 2011.  
462 Chemical composition of commercial Citrus fruit essential oils and evaluation of their  
463 antimicrobial activity acting alone or in combined processes. *Food Control*, **22**: 896- 902.
- 464 13- Farina, V., Passafiume, R., Tinebra, I., Scuderi, D., Saletta, F. and Gugliuzza, G. 2020.

- 465 Postharvest application of aloe vera gel-based edible coating to improve the quality and  
466 storage stability of fresh-cut papaya. *J. Food Qual.*, **200(1)**: 8303140
- 467 14- Gomes da Cruz, A., Buriti, F.C.A., Batista de Souza, C.H., Fonseca Faria, J.A. and Isay  
468 Saad, S.M. 2009. Probiotic cheese: health benefits, technological and stability aspects.  
469 *Trends Food Sci. Technol.*, **20(8)**: 344-354.
- 470 15- Henriques, M., Santos, G., Rodrigues, A., Gomes, D., Pereira, C. and Gil, M. 2013.  
471 Replacement of conventional cheese coatings by natural whey protein edible coatings with  
472 antimicrobial activity. *J. Hyg. Eng. Des.*, **3**: 34-47.
- 473 16- Hosseini, M., Habibi Najafi, M.B. and Mohebbi, M. 2013. Assessment of physico-chemical  
474 and sensory properties of imitation cheese containing whey protein concentrate and  
475 enzyme-modified Lighvan cheese. *Iran. J. Nutr. Sci. Food Ind.*, **8(2)**: 91-102.
- 476 17- Iranian International Standard. No2852:1995, 1st revision. Milk and milk  
477 products Determination of titrable acidity and value pH –Test method.
- 478 18- Irshad, S., Butt, M. and Younus, H. 2011. In Vitro antibacterial activity of Aloe Barbadensis  
479 Miller (Aloe vera). *Pharma.*, **1(2)**: 59-64 .
- 480 19- Jamshidi, F., Rahimi, S. and Fadaei Noghani, V. 2018. The Effect of Edible Aloe vera Gel-  
481 Persian Gum Film on Iranian White Cheese Properties. *Iran. J. Nutr. Sci. Food Technol.*,  
482 **13 (1)**: 63-74.
- 483 20- Leitgeb, M., Kupnik, K., Knez, Ž. and Primožič, M. 2021. Enzymatic and Antimicrobial  
484 Activity of Biologically Active Samples from *Aloe arborescens* and *Aloe barbadensis*. *J.*  
485 *Biol.*, **10(8)**: 765-771.
- 486 21- Khani, A. and Roufegari Nejad, L. 2018. Low fat UF-feta cheese production containing  
487 xanthan gum. *J. Food Ind. Res.*, **29(1)**: 155-167.
- 488 22- López-Córdoba, A. 2021. Feasibility of Using Carvacrol/Starch Edible Coatings to Improve  
489 the Quality of Paipa Cheese. *Polym.*, **13(15)**: 1-11.
- 490 23- Martinez-Romero, D., Paladines, D., Valverde, M., Guillén, F., Zapata, P. J. and Valero, D.  
491 2018. Rosehip oil added to aloe vera gel as postharvest coating of ‘Songría’ plums and  
492 ‘President’ prunes. *Acta Hortic.*, **1194(4)**: 321–325..
- 493 24- Mohammadi, K., Karim, G., Hanifian, Sh., Tarinejad, A. and Gasemnezhad, R. 2011.  
494 Antimicrobial effect of *Zataria multiflora* Boiss. Essential oil on *Escherichia coli* O157:H7  
495 during manufacture and ripening of white brined cheese. *J. Food Hyg.*, **1(2)**: 69-78.

- 496 25- Mohammadi, L., Hassanzadeh Khankahdani, H. and Tanaka, F. 2020. Effect of aloe vera  
497 gel combined with basil (*Ocimum basilicum* L.) essential oil as a natural coating on  
498 maintaining post-harvest quality of peach (*Prunus persica* L.) during storage. IOP  
499 Conference Series: Earth and Environmental Science, **594(1)**: 012008
- 500 26- Navarro, D., Díaz-Mula, H.M., Guillén, F., Zapata, P.J., Castillo, S., Serrano, M., Valero,  
501 D. and Martínez-Romero, D. 2011. Reduction of nectarine decay caused by *Rhizopus*  
502 *stolonifer*, *Botrytis cinerea* and *Penicillium digitatum* with Aloe vera gel alone or with the  
503 addition of thymol. *Int. J. Food Microbiol.*, **151**: 241–246.
- 504 27- Nielsen, V. and Rios, R. 2000. Inhibition of fungal growth on bread by volatile components  
505 from spices and herbs and the possible application in active packaging with special  
506 emphasis on mustard essential oil. *J. Food Microbiol.*, **60 (2-3)**: 219-29.
- 507 28- Otero, V., Raquel, B., Santosa, J., odríguez-Calleja, M.R., Nerín, C. and García-López,  
508 M. 2014. Evaluation of two antimicrobial packaging films against *Escherichia coli*  
509 O157:H7 strains in vitro and during storage of a Spanish ripened sheep cheese (Zamorano).  
510 *Food Control.*, **42**: 296-302.
- 511 29- Ortega-Toro, R., Collazo-Bigliardi, S., Roselló, J., Santamarina, P. and Chiralt, A. 2017.  
512 Antifungal starch-based edible films containing aloe vera. *Food Hydrocoll.*, **72(2)**: 1–10
- 513 30- Pantaleão, I., Pintado, M.E. and Poças, M.F. 2007. Evaluation of two packaging systems  
514 for regional cheese. *Food Chem.*, **102 (2)**: 481–487.
- 515 31- Pauli, A. 2006.  $\alpha$ -Bisabolol from chamomile-A specific ergosterol biosynthesis inhibitor. *J.*  
516 *Aromathe.*, **16**:5-21.
- 517 32- Pieretti, G.G., Pinheiro, M.P., Scapim, M.R.D.S., Mikcha, J.M.G. and Madrona, G.S. 2019.  
518 Effect of an Edible Alginate Coating with Essential Oil to Improve the Quality of a Fresh  
519 Cheese. *Acta. Sci. Technol.*, **41**: 36402.
- 520 33- Ramasubramanian, T.S., Sivakumar, V.T. and Thirumalai, A.V. 2010. Antimicrobial  
521 activity of *Aloe vera* (L.) Burm. against pathogenic microorganisms. *J. Bio. Sci. Res.*, **4(2)**:  
522 251–258.
- 523 34- Ramos, O.L., Pereira, J.O., Silva, S.I., Fernandes, J.C., Franco, M.I., Lopes-da-Silva, J.A.,  
524 et al. 2012. Evaluation of antimicrobial edible coatings from a whey protein isolate base to  
525 improve the shelf life of cheese. *J. Dairy Sci.*, **95(11)**: 6282-92.
- 526 35- Raspo, M.A., Vignola, M.B., Andreatta, A.E. and Juliani, H.R. 2020. Antioxidant and

- 527 antimicrobial activities of citrus essential oils from Argentina and the United States. *Food*  
528 *Biosci.*, **36(3)**: 27-38.
- 529 36- Rezaei, M., Yahyaei, M., Parviz, M. and Khodaei motlagh, M. 2010. A Survey of microbial  
530 contamination in Traditional Cheese distributed in Markazi Province in. *Iran. J. Health*  
531 *Environ.*, **7(1)**: 115-121.
- 532 37- Reynolds, T. and Dweck, A.C. 1999. Aloe vera leaf gel. A review updatr. *J.*  
533 *Ethnopharmacol.*, **21**: 68- 89.
- 534 38- Roy, B.C., Hoshino, M., Ueno, H., Sasaki, M. and Goto, M. 2007. Supercritical Carbon  
535 Dioxide Extraction of the Volatiles from the Peel of Japanese Citrus Fruits. *J. Essent. Oil*  
536 *Res.*, **19**: 78-84.
- 537 39- Sambu, S., Hemaram, U., Murugan, R. and Alsofi, A.A. 2022. Toxicological and  
538 Teratogenic Effect of Various Food Additives: An Updated Review. *Biomed. Res. Int.*,  
539 **24(1)**: 1-11.
- 540 40- Saritha, V., Anilakumar, K.R. and Khanum, F. 2010. Antioxidant and antibacterial activity  
541 of Aloe vera gel extracts. *Int. J. Pharm. Biol. Sci.*, **1**: 376–384.
- 542 41- Sengun, I., Yaman, D. and Gonul, S. 2008. Mycotoxins and mould contamination in cheese:  
543 a review. *World Mycotoxin J.*, **1(3)**: 291 – 298
- 544 42- Shenbagam A., Kumar, N., Rahul, K., Upadhyay, A., Gniewosz, M. and Kieliszek, M.  
545 2023. Characterization of Aloe Vera Gel-Based Edible Coating with Orange Peel Essential  
546 Oil and Its Preservation Effects on Button Mushroom (*Agaricus bisporus*). *Food Bioproc.*  
547 *Tech.*, **131(4)**: 1-22.
- 548 43- Sumi, F.A., Sikder, B., Rahman, M.M., Lubna, S.R., Ulla, A., Hossain, M.H., Jahan, I.A.,  
549 Alam, M.A., and Subhan, N. 2019. Phenolic Content Analysis of Aloe vera Gel and  
550 Evaluation of the Effect of Aloe Gel Supplementation on Oxidative Stress and Fibrosis in  
551 Isoprenaline-Administered Cardiac Damage in Rats. *Prev. Nutr. Food. Sci.*, **24(3)**: 254-  
552 264.
- 553 44- Suriati, L., Utama, I.M.S., Harjosuwono, B.A. and Gunam, B.W. 2020. Stability Aloe Vera  
554 Gel as Edible Coating. *Earth Environ. Sci.*, **411(2)**: 1-6.
- 555 45- Suriati, L. 2018. Studies the Resistance to Oxidation and the Changes Phases against the  
556 Characteristics of Physicochemical Aloe vera Gel. *J. Bio. Chem. Research.*, **35(2)**: 670-  
557 679.

- 558 46- Tan, Q., Ai, M. and Minh, N. 2011. Volatile constituents of essential oil from citrus sinensis  
559 grown in tine giant province, Vietnam. *Asian J. Food Agro. Ind.*, **4(3)**: 183-186
- 560 47- Trmčić, K., Chauhan, A., Kent, D.J., Ralyea, R.D., Martin, N.H., Boor, K.J. and Wiedmann,  
561 M. 2016. Coliform detection in cheese is associated with specific cheese characteristics,  
562 but no association was found with pathogen detection. *J. Dairy Sci.*, **99(8)**: 6105-6120.
- 563 48- Tzortzakis, N., Xylia, P. and Chrysargyris, A. 2019. Sage essential oil improves the  
564 effectiveness of aloe vera gel on postharvest quality of tomato fruit. *Agron.*, **9(10)**: 635-  
565 643.
- 566 49- Wagh, Y.R., Pushpadass, H.A., Magdaline Eljeeva Emerald, F. and Surendra Nath, B.  
567 2013. Preparation and characterization of milk protein films and their application for  
568 packaging of Cheddar cheese. *Food Sci. Technol.*, **51(12)**: 3767-3775.
- 569 50- Wang, Q., Yu, H., Tian, B., Jiang, B., Xu, J., Li, D., Feng, Z. and Liu, C. 2019. Novel  
570 Edible Coating with Antioxidant and Antimicrobial Activities Based on Whey Protein  
571 Isolate Nanofibrils and Carvacrol and Its Application on Fresh-Cut Cheese. *J. Coat*, **9(9)**:  
572 583-591.
- 573 51- Yerlikaya, O. and Ozer, E. 2014. Production of probiotic fresh white cheese using co-  
574 culture with Streptococcus thermophilus. *Food Sci. Technol.*, **34(3)**: 1-10.
- 575 52- Yilmaz, F. and Dagdemir, E. 2012. The effects of beeswax coating on quality of Kashar  
576 cheese during ripening. *Int. J. Food Sci. Tech.*, **47**: 2582-2589.
- 577 53- Zhong, Y., Cavender, G. and Zhao, Y. 2014. Investigation of different coating application  
578 methods on the performance of edible coatings on Mozzarella cheese. *LWT Food Sci.*  
579 *Technol.*, **56**: 1-8

580

581 **تأثیر پوشش فعال مبتنی بر ژل آلونته ورا با اسانس پوست لیمو بر ماندگاری و ویژگی های کیفی پنیر**

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588 در این مطالعه تأثیر پوشش خوراکی ژل آلوئه ورا حاوی اسانس پوست لیمو (0، 100 و 150 پی پی ام) بر ویژگی های کیفی نمونه های

589 پنیر مورد بررسی قرار گرفت. تیمارها شامل 4 گروه کنترل (بدون پوشش)، ژل آلوئه ورا، ژل آلوئه ورا و 100 پی پی ام اسانس پوست

590 لیمو و ژل آلوئه ورا و 150 پی پی ام اسانس پوست لیمو بود. این تیمارها به مدت 60 روز از نظر خواص فیزیکوشیمیایی، بافتی، حسی و

591 شمارش میکروبی مورد ارزیابی قرار گرفتند. یافته ها نشان داد که با افزایش مدت نگهداری، اسیدیته و نمک افزایش و pH و رطوبت

592 کاهش یافت. در ارزیابی ویژگی های حسی، تأثیر تیمارها بر تمامی ویژگی های حسی به جز امتیاز رنگ معنی دار بود. نمونه های پوشش

593 داده شده با ژل آلوئه ورا و 100 پی پی ام پوست لیمو بالاترین امتیاز طعم (4/97) را دریافت کردند. با افزایش زمان نگهداری، سختی،

594 قابلیت جویدن و فتری بودن نمونه های پنیر افزایش یافت. چسبندگی نمونه ها تحت تأثیر مدت زمان نگهداری قرار نگرفت. در پایان زمان

595 نگهداری، بیشترین شمارش کل میکروبی، کپک و مخمر مربوط به نمونه های پنیر شاهد (به ترتیب 5/37 و  $\log \text{cfu/g} 4/62$ ) و

596 کمترین مقدار مربوط به نمونه های پوشش داده شده با ژل آلوئه ورا و 100 پی پی ام پوست لیمو (به ترتیب 3/92 و  $\log 3/76$   $\text{cfu/g}$ )

597 . به طور کلی استفاده از پوشش خوراکی تهیه شده با یل آلوئه ورا و غلظت های کمتر اسانس پوست لیمو (100 پی پی ام و

598 کمتر) باعث بهبود ظاهر و طعم نمونه های پنیر طی 60 روز نگهداری گردید.

599 واژگان کلیدی: آلوئه ورا، پوشش خوراکی، اسانس پوست لیمو، پنیر، خواص بافتی، ارزیابی حسی.

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