Evidence for Physiological Vascular Occlusion in Stems of Cut Gerbera cv. Hongyan

R. Wang¹*, X. Zheng¹, and X. Xu¹

ABSTRACT

The aim of this paper was to assess the effect of catechol (CH), as a peroxidase inhibitor, and 8-hydroxyquinoline citrate (HQC), as a bacterial inhibitor, on keeping the quality of Gerbera Jamesonii cv. Hongyan cut flowers, known to be limited by vascular occlusion. Blockage in the stem xylem vessels of cut gerbera was studied using light and scanning electron microscope. The results showed that some stem xylem vessels of the cut flowers were blocked during the vase period, and the blockage consisted of some amorphous or physiological deposition and rod-shaped bacteria located within the 5cm stem end of the cut flower. In addition, CH (1.0mM) or 8-HQC (0.45mM) decreased the blockage of stems and reduced the bacterial growth in the vase solution, but extended the vase life without statistical significance. The combination of CH (0.5 or 1.0 mM) and 8-HQC (0.45mM) decreased the blockage and inhibited the bacteria more than CH or 8-HQC alone, and extended the vase life significantly (P ≤ 0.05).

Keywords: Cut flowers, Catechol, Gerbera jamesonii, Bacterium inhibitor, Vase life, Xylem blockage.

INTRODUCTION

Many works on wilting mechanism and preservation of cut flowers have been reported (Podd et al., 2002; van Doorn and Vasilier, 2002; Damunupola et al., 2010; Ahmad et al, 2013). The vase life of cut flowers is determined by the phenotypexpostharvest conditions interaction (Fanourakis et al., 2013). However, insufficient water uptake due to the xylem occlusion is one of the direct reasons for wilting during the vase period (Ieperen et al., 2002).

Gerbera jamesonii and its hybrids are well known for their variable shapes and colors in the world (Solgi et al., 2009; Liu et al., 2009). But, the vase life of cut gerbera is often shortened by stem bending due to insufficient water uptake (van Meeteren, 1978). Stem end blockage is a major factor in the imbalance between water uptake and water loss from cut flowers (He et al., 2009). Some studies suggested that the blockage of cut gerbera flower is caused by bacteria (microbes) (van Meeteren, 1978; Put, 1990; He et al., 2009) or bacteria and decay products (Liu et al., 2009). Bacterial inhibitors such as 8-HQC, silver nanoparticles or salicylic acid could extend vase life of cut flowers, whose vascular occlusion is caused by bacteria (Marousky, 1971; Loubaud and van Doorn, 2004; Solgi et al., 2009; Mashhadian et al, 2012). HQC, as an antibacterial agent, may also promote flower longevity by acidifying the vase solution, or influencing the activity of some enzymes, or their chelating properties (Edrisi et al., 2012).

Physiological deposition such as lignin, suberin, mucilage, gums or tyloses (Loubaud and van Doorn, 2004, van Doorn and Cruz,
2000) and air emboli or cavitations (van Meeteren et al., 2006) may also decrease the vase life of some cut flowers. The air emboli or cavitations may be not serious if cut flowers are put into water immediately after cutting (Ieperen et al., 2002). Thus, physiological inhibitors such as CH and S-carvone extended the vase life of cut chrysanthemum or Grevillea flowers, whose stem blockage was due to physiological deposition (van Doorn and Vaslier, 2002; Loubaud and van Doorn, 2004; He et al., 2006).

The blockage can be directly seen with the light microscopy technique, but this technique could not distinguish blockage caused by physiological deposition or bacteria (He et al., 2009; Liu et al., 2009), which can be directly observed with scanning electron microscopy (SEM). Most previous studies have reported the vascular occlusion of some cut flowers indirectly by deduction and not by observation with SEM. Only a few studies were performed using SEM (Clerkx et al., 1988; Macnish et al., 2008; He et al., 2009). Further observation with SEM can contribute to determine whether physiological deposition or bacterial growth occurred in stem of cut flowers during vase period.

In this research, we studied the effect of CH, as a physiological inhibitor, on the vase life of cut gerbera, and observed the stem vessel blockage with light microscope and SEM. The objectives of this study were to investigate: (1) how CH, as a physiological inhibitor, affects the vase life of cut gerbera flower, (2) what materials other than bacteria block the vascular stem of cut gerbera during vase period, and (3) what are the influences of the blockage on the vase life of cut gerbera flower?

### MATERIALS AND METHODS

#### Plant Materials

*Gerbera Jamesonii* cv. Hongyan flowers were obtained from a commercial grower in Linyi, located at 35°03′N, in the south of Shandong province, China. The flowers were grown in a greenhouse and harvested in the morning on May, 2011. Cut flowers were covered with plastic films and transported within 4 hours to the laboratory in Jiangsu Normal University, where the flower stems were re-cut in distilled water to a length of 25 cm. Thirty-two flowers were selected for the vase life experiment and 3 flowers for fresh stem structure observation.

#### Experimental Designs and Treatments

The experiment was conducted in the laboratory at 18–22°C, 35–55% RH, and 15 μmol m⁻² s⁻¹ light intensity (cool white fluorescent tubes) under a daily light period of 12 hours. Vase solutions were freshly prepared at the beginning of the experiment and not renewed in the course of study. The experiment was carried out in a factorial design. The vase solutions included distilled water, 0.45 mM 8-HQC with 0.06M sucrose, 0.5 and 1.0 mM CH with 0.45 mM 8-HQC and 0.06M sucrose (Table 1). A complementary experiment containing CH at 1.0 mM was conducted to verify the effect of CH alone on cut gerbera flowers. CH was obtained from a Chemical company (Alfa Aesar Chemical Co. Ltd. Tianjin, China). Sucrose was used as a food source or for water balance maintenance of vase flowers (Solgi et al., 2009).

The cut flowers were immediately immersed individually into a 150 ml conical

### Table 1. Prescription for preserving cut gerbera flowers.

<table>
<thead>
<tr>
<th>Components</th>
<th>Treatments</th>
<th>water</th>
<th>HQC</th>
<th>HQC and 0.5 mM CH</th>
<th>HQC and 1.0 mM CH</th>
</tr>
</thead>
<tbody>
<tr>
<td>water</td>
<td>Distilled water</td>
<td>0.45mM 8-HQC</td>
<td>0.5mM catechol+ 0.45 mM</td>
<td>1.0 mM catechol+ 0.45 mM</td>
<td></td>
</tr>
<tr>
<td></td>
<td>+0.06M sucrose</td>
<td>8-HQC +0.06M sucrose</td>
<td></td>
<td>8-HQC +0.06M sucrose</td>
<td></td>
</tr>
</tbody>
</table>
Vascular Occlusion of Cut Gerbera

beaker containing 100 ml solution after recut. Every treatment included 3 replications for stem structure observation and 5 replications for the other experiments. Vase mouths were covered with small masses of absorbent cotton to minimize water evaporation and prevent contamination. The vase life, fresh weight (FW), relative fresh weight (RFW), and water balance (WB) of the flowers were assessed daily during the vase period.

**Vase Life and Relative Fresh Weight**

The vase life was judged to the end when the cut flowers were dropping or their petals were moderately wilting (Macnish et al., 2008). The FW (g) of cut flowers was daily recorded and the RFW was calculated by the formula:

\[ RFW(\%) = \frac{FW_t}{FW_0} \times 100 \]  

Where, \( t \) = Day 0, 1, 2, etc., respectively, \( FW_t \) is the fresh weight at day \( t \), \( FW_0 \) is the fresh weight at day 0 (He et al., 2006).

The WB was calculated by the formula:

\[ WB = U - L \]  

Where, \( U \) is water uptake and \( L \) is water loss of the cut flowers. And \( U = SC_{t,i} - SC_i \), where, \( SC_i \) is total weight of the solution and conical beaker at \( t \) = Day 1, 2, 3, etc., respectively, and \( SC_{t,i} \) is the weight of the solution and conical beaker on the previous day; \( L = SCF_{t,i} - SCF_i \), where, \( SCF_i \) is total weight of the same solution and conical beaker and flowers at \( t \) = Day 1, 2, 3, etc., respectively, and \( SCF_{t,i} \) is the weight of the same solution and conical beaker and flowers on the previous day.

**Bacterial Counts**

Bacterial counts in the vase solutions were examined using the plate count method (Liu et al., 2007) on the 5th day. The vases were agitated to stir their solution. A 0.2 ml of the diluted sample solution was pipetted onto a plate count agar (beef extract peptone medium). The beef extract peptone medium consisted of beef extract (0.3 g), peptone (1 g), sodium chloride (0.5 g), agar (2 g), and distilled water (100 ml), and its pH value was 7.2. The bacteria on the agar were incubated for 24 hours at 37°C and the bacterial colonies were enumerated.

**Light Microscopy and Scanning Electron Microscopy**

To observe the blockage of the fresh stem on day 0 and the stems of different treatments on the 5th day a 2 cm segment (denoted the basal 0–2 cm segment) and a 1 cm segment (denoted the basal 5–6 cm segment) were excised and fixed in formalin-acetic acid (FAA). For the light microscopy observation, the specimens in the FAA were rinsed with distilled water, and, then, for paraffin sectioning through a gradual concentration series of Ethyl alcohol, as a dehydration agent, and Dimethylbenzene, as a transparence agent, and with Safranin T and fast green as double staining agents (Li, 1996). Photographs were taken by the light microscope with YM310 DV. For the SEM observation, the specimens in the FAA were rinsed in distilled water, and dehydrated in a gradual series of 70, 80, 95, and 100% (v/v) Tert-butyl alcohol for 10 minutes every step and 100% Tert-butyl alcohol 3 times (Gu, 2006). The dehydrated specimens were frozen and dried in a Hitachi ES-2030 freeze dryer (Hitachi Ltd., Tokyo, Japan), then sputter-coated with gold in a Hitachi E-1010 ion sputter (Hitachi Ltd., Tokyo, Japan). The transverse section of the stem base was observed under the Hitachi S-3400N SEM (Hitachi Ltd., Tokyo, Japan).

**Statistical Analyses**

The data were analyzed by relative analysis of variance (ANOVA) with the
RESULTS

Vase Life

At the end of vase life, the stems in distilled water and in HQC alone bent over at 15-20 cm from stem bottom (Figure 1-B), and in 0.5 or 1.0 mM CH solution with HQC, the stem broke at points 10 cm from the bottom and the stem base turned brown (Figure 1-C). The petals of cut gerbera flowers in different treatments did not wilt.

Compared to the control (distilled water), the vase solution of 0.5 or 1.0 mM CH with HQC extended the vase life of flowers significantly, and the longest vase life was obtained at 1.0 mM (Table 2). HQC alone extended the vase life, but not significantly (Table 2), and CH alone at 1.0 mM had no effect on the vase life (data not shown).

Relative Fresh Weight and Water Balance

The treatments of 0.5 and 1.0 mM CH with HQC both delayed loss of the RFW during the vase period, and 1.0 mM CH best maintained the RFW (Figure 2-A) and the WB (Figure 2-B).

Bacterial Numbers

Compared to the control (distilled water), different treatments decreased bacterial numbers of vase solutions significantly (Table 2). CH (0.5 or 1.0 mM) with HQC reduced bacterial numbers more than HQC alone did.

Stem Blockages

Observation with the light microscopy showed that the xylem vessels at the 0–2 cm stem base in water were blocked by some
material dyed into dark green (Figure 3-A). HQC or CH (0.5 mM or 1.0 mM) with HQC decreased the blockage (Figure 3, B-D). No blockage was observed in the fresh stem. Since SEM image shows the results more clearly, the data with light microscopy are not shown.

Observation with SEM showed that the xylem vessel of fresh stems had no occlusion (Figure 4-A). On the 5th day of the vase life, some mixture of amorphous matter and rod-shaped bacteria was observed in the xylem vessels of stems in distilled water (Figure 4, C–F), but few in 1.0 mM CH with HQC treatment (Figure 4-B).

In the basal 5–6 cm segment, no blockage was observed in all treatments, which were the same as that of the fresh stem, therefore, the data are not shown.

**DISCUSSION**

The vascular occlusion of gerbera stems has been considered to be mainly due to microbial proliferation (van Meeteren, 1978; Put, 1990; He et al., 2009, Liu et al., 2009). In the present study, besides the rod-shaped bacteria, some amorphous matter was observed in the vascular vessels of gerbera stems. This amorphous matter might be mucus or gums due to wound reaction rather than lignin or suberin or tyloses, because lignin or suberin could be dyed red by Safranine, and tyloses are membrane-bound invaginations (Tagne et al., 2002) or the membrane-bound cell bodies into vessels. It was during the vase period that the amorphous matter was produced, because no blockage was observed in the vessels of fresh stems. This result suggested
that the vascular occlusion of gerbera stems was apparently due to physiological process involving oxidation. Tagne et al. (2002) observed that the deposition occurred in maize xylem vessels after fungal invasion and considered the depositions were gums and gels, which were part of the host defense response. In this paper, the depositions may be a response to wound, and could be prevented by combining CH with HQC.

No blockage was observed in the basal 5-6 cm stem, which suggested that blockage of the gerbera flowers was located within the stem base of 5cm. This result was similar to the findings on chrysanthemum (van Doorn and Vaslier, 2002), Bouvardia Domestica cv. Van Zijverden (Vaslier and van Doorn, 2003), Grevillea (He et al., 2006) and gerbera (He et al., 2009).

CH combined with HQC decreased bacterial numbers of vase solutions and reduced blockage (Figure 3-D) more than HQC alone did, which implied that CH as a peroxidase inhibitor (Srivastava and van Huystee, 1977; Wang et al., 1991; van Doorn and Vaslier, 2002) suppressed the microbial proliferation, or increased the effect of HQC as bacterial inhibitor. Maddox et al. (2010) presented that CH was effective in inhibiting a pathogenic bacterium (Xylella fastidiosa). Alternatively, Loubaud and van Doorn (2004) considered that 8-HQC, as a bacterial inhibitor, suppressed the plant-induced xylem occlusion. Therefore, CH or HQC could not only be referred to as a physiological inhibitor, but also an antibacterial agent in the preservation of cut flower. The mechanism of CH or HQC acting on the vase life of cut flowers still needs more studies.

Water loss of cut gerbera flowers was aggravated by lower air humidity in the test room than recommended (Fanourakis et al., 2013), therefore, in this study, the vase life of cut gerbera in distilled water was shorter than the findings of Solgi et al. (2009).

CONCLUSIONS

CH at 1.0 mM combined with HQC extended the vase life of the gerbera flowers significantly, while CH or HQC alone had a weak effect. Both CH with HQC and HQC alone reduced bacteria counts of the vase solution and decreased the occlusion of the cut flower stems. The occlusion occurred within 5 cm of the stem base, and consisted of not only bacteria, but also amorphous mucilage or...
gums involving wound-induced physiological processes of the cut flowers.

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REFERENCES


Hongyan

شواهدی برای گرفتگی (انсадاد) آوندی در ساقه زیریای بریده کالیبوار

چکیده

از آنکه گرفتگی آوندی عامل اصلی عامل اصلی زیریای بریده را محدود می‌کند، هدف پژوهش حاضر ارزیابی اثر 8-hydroxyquinoline citrate (HQC) پازدارنده باکتری در حفاظت کیفیت زیریای بریده برگ است. برای مطالعه گرفتگی Jamesonii cv. Hongyan پازدارنده باکتری در حفاظت کیفیت زیریای بریده در آوند جوبی ساقه زیریای میکرووسکوب نوری و میکروسکوب الکترونی اسناده شد. نتایج نشان داد بعضی آوندهای ساقه گل بریده در طی زمان در گلدان دچار گرفتگی می‌شدن، این گرفتگی ناشی از رسوش های آمورفوس (بی شکل) با فیزوپولیزوم در آوند برای رشد باکتری‌ها لوله ای شکل در 5 ساعت متی ای شده‌اند. برای هر یک از مواد به نتیجه‌ی بود همچنین، تریکب مرزی عامل گلدانی زیریا می‌باشد. پذیره می‌باشد.

References