

## Haplotype and Genetic Diversity of mtDNA in Indigenous Iranian Sheep and an Insight into the History of Sheep Domestication

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### ABSTRACT

The archaeological evidence suggests that Iran has been one of the first origins of sheep domestication in the world. This study aims to investigate the genetic diversity of indigenous Iranian sheep breeds using mitochondrial DNA (mtDNA) and to explore the evolutionary history of sheep domestication in Iran. Single Nucleotide Polymorphism (SNP) markers in the control region of mtDNA were used to genotype the unrelated sheep samples of Zel and Lori-Bakhtiari breeds which were collected from or near the center of the sheep domestication, using the Sequenom MassARRAY platform. Phylogenetic analysis of the mitochondrial SNPs classified all animals into either of two haplogroups A and B. The population differentiation ( $F_{ST}$ ) and gene flow ( $N_m$ ) statistics were 0.054 and 4.715 respectively, indicating a low mitochondrial genetic differentiation and high gene flow between two sheep breeds. The Analysis of Variation (AMOVA) showed that around 97% of the total genetic diversity is distributed within the two breeds. Further analysis using SNP haplotyping identified nine different haplotypes within the animals; eight haplotypes were present in the Zel, while only four were seen in the Lori-Bakhtiari breed. Two haplotypes, designated H<sub>1</sub> and H<sub>3</sub>, were present at higher frequencies in both breeds. Haplotypes H<sub>5</sub>, H<sub>6</sub>, H<sub>7</sub>, H<sub>8</sub> and H<sub>9</sub> were found as population-specific in the Zel, and haplotype H<sub>2</sub> only occurred in Lori-Bakhtiari breed. The existence of two common Haplotypes (H<sub>1</sub> and H<sub>3</sub>) in the animals suggest that the two Iranian breeds from strikingly different geographical regions, may share a common ancestry, and these haplotypes could be the origin haplotypes while the population specific haplotypes developed later.

**Keywords:** Genetic diversity, Haplogroups, Mitochondrial genome, Phylogenetic study, Sheep evolution.

### INTRODUCTION

Domestication of livestock represents a crucial step in human history. The rise of civilisations could not happen without the domestication of plants and animals. Sheep (*Ovis aries*) is the first grazing

animal known to be domesticated (Chessa *et al.*, 2009). The archaeozoological evidence suggests that the domestication of sheep occurred during the Neolithic revolution approximately 9000 years ago (Kijas *et al.*, 2009), in a region in northern Iraq and nearby regions close to Zagros

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mountains in Iran (Zeder, 1999). This means that the indigenous Iranian sheep breeds were among the first to be domesticated in the world. The wide geographical distribution patterns of retrovirus insertion site polymorphisms suggested that there were two major waves of sheep migrations: firstly, the spread of early domestic sheep presumably from the center of domestication, followed by the second wave of sheep (with improved traits) migration from Southwest Asia around 5,000 years BP (Chessa *et al.*, 2009). Therefore, it is of essential interest to study the major native sheep breeds of Iran, Lori-Bakhtiari and Zel, using mitochondrial DNA (mtDNA) approach in an attempt to understand the history of domestication and migrations.

The genetic history of sheep has been investigated based on three major sources of genomic variations, namely the autosomes, *Y* chromosome and mitochondrial genome (Agaviezor *et al.*, 2012). Presently, mtDNA has received much attention because its markers provide important phylogenetic information in relation to genetic diversities. The rate of mtDNA mutations is about 5 to 10 times faster than nuclear DNA, and its genes do not usually recombine (Upholt *et al.*, 1977). Mammalian mtDNA is deemed to strictly follow maternal inheritance and is highly variable within a species, so mtDNA provides an important resource for phylogenetic studies. Based on mtDNA sequence, four distinct haplogroups (namely A, B, C and D) have been found in the domestic sheep, indicating four maternal lineages. The main haplogroups A and B are both found in Asia, while B dominates in Europe (Guo *et al.*, 2005; Cinkulov *et al.*, 2008). Haplogroup C has been found in Portugal, Turkey, the Caucasus and China (Tapio *et al.*, 2006; Chen *et al.*, 2006; Pardeshi, 2007; Zhao *et al.*, 2011). Haplogroup D, present in Rumanian Karachai and Caucasian

animals, is possibly related to the haplogroup A (Pariset *et al.*, 2011). In contrast to cattle, the sheep haplogroups hardly correlate with geography.

Sheep production is the most important component of the Iranian livestock industries, with a population of 50 million animals which are well adapted to the harsh arid of sub-tropical areas (Mokhtari *et al.*, 2015). Two main Iranian indigenous sheep breeds were investigated in the present study. The Lori-Bakhtiari sheep is one of the most common breeds, with a population of more than 1.7 million animals. It is well adapted to the hilly and mountainous Bakhtiari region which stretches out to the southern Zagros Mountains (Figure 1). The animals are kept mostly in villages under semi-intensive systems. Relative to other Iranian fat-tail breeds, Lori-Bakhtiari is a large animal, having the largest fat-tail by girth and weight (Vatankhah and Talebi, 2008). Zel is the only thin-tail Iranian breed; it is present largely on the northern slopes of the Elburz mountain range near the Caspian Sea (Moradi *et al.*, 2012), and constitutes around 3% of the Iranian sheep population. This breed is also known as the Aryan breed since the historical evidence shows that the Aryans, who were living in these areas, attempted to domesticate these animals. It is a general belief that the first domesticated sheep were thin-tail and the fat-tail animals were developed later (Moradi *et al.*, 2012). Hence, Zel breed is also known as a contour of wild and domesticated strains in Iran.

Very little work has been done on the genetic background and phylogenetic history of Iranian sheep breeds using mtDNA. The aim of the present study was to examine and describe the genetic diversity and phylogeny of indigenous Lori-Bakhtiari and Zel sheep breeds in Iran. With the animal samples collected from or near the center of domestication, we hope to shed some light on the



**Figure 1.** (a) Traditional distributions of the two Iranian breeds (Lori Bakhtiari and Zel) used in this study. Examples of a fat-tailed Lori-Bakhtiari animal (b) and a thin-tailed Zel animal (c).

domestication event and on the history of Iranian sheep.

## MATERIALS AND METHODS

### Animal Samples

Blood samples of 150 Zel (134 females and 16 males) and 107 Lori-Bakhtiari (89 females and 18 males) animals were collected from farms distributed across the traditional rearing areas of each breed. Blood were collected in EDTA vacutainers by jugular venepuncture. The animals were from groups which were in the registration and recording system of the Animal

Breeding Centre of Iran. Two criteria were used when selecting the experimental animals, namely the animals must be unrelated and from diverse geographical regions. While sampling pedigreed animals we ensured that they had no common antecedents, and in non-pedigreed animals we typically selected 4-5 animals of different ages from each group. The Zel breed animals were collected from the northern province of Mazandaran located in the south of the Caspian Sea. The Lori-Bakhtiari animals were from the Chaharmahal and Bakhtiari Province, which are located in the south western part of Iran close to the Zagros Mountain ranges.



## SNP-Multiplex Design, DNA Extraction and Genotyping

In the 1,180-bp sequence of the mtDNA control region, there are 39 SNP markers which differentiate between mtDNA haplotypes A and B (Wood and Phua, 1996). These markers were used in the construction of a SNP-multiplex utilizing the MassARRAY Assay Design 4.0 software (Sequenom Inc., San Diego, CA, USA). The mtDNA sequence reported in Wood and Phua (1996) was used for designing the primers. We obtained a multiplex consisting of ten SNP markers (Table 1).

Total DNA, consisting of genomic and mitochondrial DNA, was extracted from whole blood using a modified salting out protocol (Moradi *et al.*, 2012). DNA samples were diluted to 50 ng  $\mu\text{l}^{-1}$  concentration. Genotyping with the SNP-multiplex was performed following the Sequenom's recommended protocol, based on the iPLEX<sup>TM</sup> genotyping assay (Oeth *et al.*, 2005). Reactions were carried out in a 384-well plate, using 25 ng of DNA per animal. The reaction products were transferred onto chips and analysed in a MassARRAY Compact 96 mass spectrometer.

### Data Analysis

Nucleotide variations in the ten SNP sites of the 257 Lori-Bakhtiari and Zel animals were analysed and aligned using MEGA v.5.1 software, with a Kimura 2-parameter (with transitions and transversions) model and a bootstrap (1,000 replications) test (Tamura *et al.*, 2011). DnaSP 5.1 software (Rozas *et al.*, 2009) was used to calculate haplotype, indices of nucleotide variation and haplotype structure including nucleotide diversity ( $\pi$ ), number of haplotypes (nh), Haplotype diversity (Hd) and average number of nucleotide substitutions (Dxy) per site between breeds. To evaluate the fit of major models of nucleotide substitutions based on obtained haplotypes, MEGA v.5.1 software

**Table 1.** Sequences of primers for SNP markers.<sup>a</sup>

| SNP   | Nucleotide variants | Sequence of PCR-primer A       | Sequence of PCR-primer B        | Sequence of extension primer |
|-------|---------------------|--------------------------------|---------------------------------|------------------------------|
| SNP01 | C/T                 | ACGTTGGATGGGAGCCTCAGTAGATCTAAC | ACGTTGGATGGGCTCTAGAAITGAAGAG    | AGTAGATCTAACTAATTTCTCCCTACA  |
| SNP02 | T/C                 | ACGTTGGATGGAAGCGTTGCTAGTCAACTG | ACGTTGGATGGGTTACTTCACGTCAGCTAC  | TACTCTGTTGGGATGGC            |
| SNP03 | C/T                 | ACGTTGGATGACCAGGAACTGCGTTTAC   | ACGTTGGATGGTAGGACATCTCGGAAGAG   | TGCGTTACTAGAAGTAGA           |
| SNP04 | G/A                 | ACGTTGGATGGTCCGATATGTAATTTG    | ACGTTGGATGCTACTAGTTTTCGCACTTG   | CCATATGTAATTTGACACCAATTAC    |
| SNP05 | G/A                 | ACGTTGGATGGCGTTTAGCCGTTCTGTTG  | ACGTTGGATGTCACGCCACAGAACTAATC   | TAATAATGATAAGTGTGGGGACTAG    |
| SNP06 | A/G                 | ACGTTGGATGAAAGCCCATGTAGAAGCTCC | ACGTTGGATCGGTATCATGCCATATCCCTCC | GTAGAAGCTCCAATTGC            |
| SNP07 | G/A                 | ACGTTGGATGGGCTAGGATAAATTTTCGGG | ACGTTGGATGCTTTCACACGAGAAAATGCAC | AAGTAGACTGGTAGTATATG         |
| SNP08 | T/C                 | ACGTTGGATGGTTGGAGATCTCAGGTGTTG | ACGTTGGATGAGCCCTTACAAGCAATCC    | TGGTAGAATAATCCGATGTC         |
| SNP09 | T/C                 | ACGTTGGATGGACATAAGATATGTTGGGTT | ACGTTGGATGCCACATAAACCCTAC       | TGTTGGTTTATGAACCGTC          |
| SNP10 | G/A                 | ACGTTGGATGGGGAAGCGTGTAAAAATGG  | ACGTTGGATGCCCTATAATGGTAGCATGG   | GGGGGAAAAAAGAAATATAAAATG     |

<sup>a</sup> All the ten markers were designed and used as one SNP-multiplex.

was applied and phylogenetic relationships among mtDNA haplotypes were obtained by Maximum Likelihood (ML) as implanted in this software. An Analysis of Molecular Variance (AMOVA) was computed to test significant differences in mtDNA diversity between the breeds using ARLEQUIN 3.01 (Excoffier *et al.*, 2005). The Median Joining (MJ) networks were plotted using the Network 4.6.1.2 program (<http://www.fluxus-engineering.com>) to reveal the possible relationships between haplotypes.

## RESULTS AND DISCUSSION

In this study, 24 widely-used models were investigated for phylogenetic analyses. Models with least Bayesian Information Criterion (BIC), Akaike Information Criterion (AIC) and Maximum Likelihood (ML) information indices were used for description of the best substitution pattern. In the end, Kimura 2 (K2) model was selected as the best model for phylogenetic studies.

From the 257 analysed animals, nine haplotypes were found in Zel and Lori-Bakhtiari breed samples which were designated H<sub>1</sub> to H<sub>9</sub> (Table 2). In contrast to the Lori-Bakhtiari breed which has only four haplotypes (H<sub>1</sub>, H<sub>2</sub>, H<sub>3</sub> and H<sub>4</sub>), Zel breed has higher mtDNA diversity with eight

haplotypes (all haplotypes except H<sub>2</sub>). Although, most of the selection pressures do not act on the mtDNA, unless the selection occurs on energy metabolism, it is an observation that the low diversity of haplotypes in Lori-Bakhtiari breed may be due to the higher selection of pressure imposed on this breed (Moradi *et al.*, 2012). The higher haplotypes number seen in Zel breed is also likely to be due to the higher number of animals analysed: 150 Zel versus 107 Lori-Bakhtiari. Furthermore, the number of haplotypes present in the females was greater than that in the males (Table 3). Though most of the breeding programs in sheep are focused on rams, the latters do not pass on their mtDNA to the progeny. This again is possibly due to the higher number of female animals analysed (223 females versus 34 males).

The results of this study revealed that the most common haplotypes in Lori-Bakhtiari and Zel populations were H<sub>3</sub> and H<sub>1</sub>, respectively. These haplotypes were present in high frequencies in both breeds (Figure 2). Haplotype H<sub>2</sub> appeared to be population-specific in the Lori-Bakhtiari breed, while H<sub>5</sub>, H<sub>6</sub>, H<sub>7</sub>, H<sub>8</sub> and H<sub>9</sub> haplotypes are found only in the Zel population but at low frequencies (Table 2). The network analysis of haplotypes shows big differences between low-frequency haplotypes in Zel and Lori-Bakhtiari

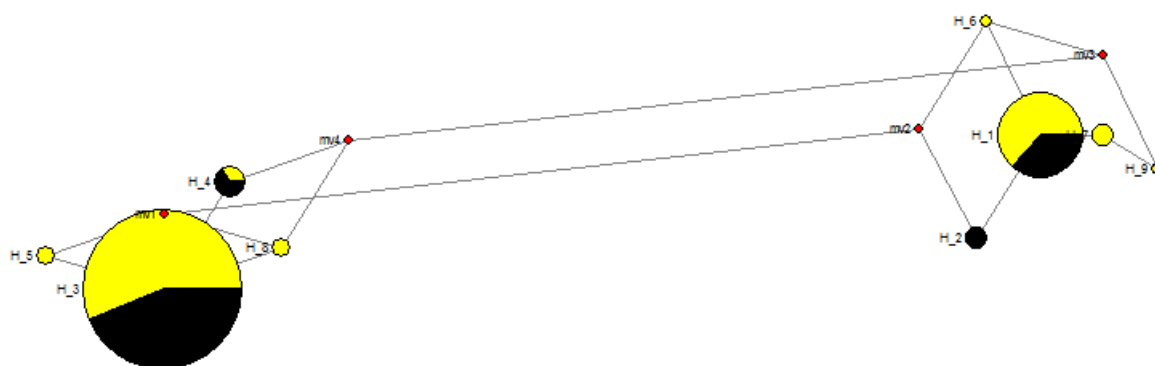
**Table 2.** The two mtDNA haplogroups A and B, and the nine haplotypes present in the 257 Zel and Lori-Bakhtiari sheep. Haplogroups A and B are represented by H<sub>1</sub> and H<sub>3</sub> haplotypes, respectively. The other haplotypes are variants of the two ancestral haplogroups.

| Haplogroup | Haplotype      | Breed                  | SNP variants <sup>a</sup> |
|------------|----------------|------------------------|---------------------------|
|            |                |                        | TCCGAGGTTG                |
| A          | H <sub>1</sub> | Lori-Bakhtiari and Zel | TCCGAGGTTG                |
| A          | H <sub>2</sub> | Lori-Bakhtiari         | TCCGAGGTTA                |
| B          | H <sub>3</sub> | Lori-Bakhtiari and Zel | <b>CTTAGAACCA</b>         |
| B          | H <sub>4</sub> | Lori-Bakhtiari and Zel | <b>CTTAGAACCG</b>         |
| B          | H <sub>5</sub> | Zel                    | <b>CTCAGAACCA</b>         |
| A          | H <sub>6</sub> | Zel                    | TTCGAGGTTG                |
| A          | H <sub>7</sub> | Zel                    | <b>CCTGAGGTTG</b>         |
| B          | H <sub>8</sub> | Zel                    | <b>TTTAGAACCA</b>         |
| A          | H <sub>9</sub> | Zel                    | <b>TCCTGAGTTG</b>         |

<sup>a</sup> Haplotype with the highest frequency is haplotype H<sub>1</sub> which is used here as the reference sequence. The nucleotide variants which are different to the reference bases are bold.

**Table 3.** Population statistics and haplotype diversity in Lori-Bakhtiari and Zel breeds based on mtDNA SNP markers.

| Population | No. haplotypes | Haplotype diversity | No. different nucleotides (K) |
|------------|----------------|---------------------|-------------------------------|
| All        | 9              | 0.454               | 7.473                         |
| SubLori1   | 4              | 0.428               | 7.011                         |
| SubLori2   | 2              | 0.303               | 6.061                         |
| SubLori3   | 2              | 0.476               | 9.542                         |
| SubZel1    | 5              | 0.708               | 10.401                        |
| SubZel2    | 4              | 0.692               | 9.077                         |
| SubZel3    | 2              | 0.533               | 10.660                        |
| SubZel4    | 2              | 0.556               | 11.112                        |
| SubZel5    | 4              | 0.600               | 10.152                        |
| SubZel6    | 3              | 0.522               | 8.206                         |
| SubZel7    | 2              | 0.209               | 4.183                         |
| SubZel8    | 2              | 0.143               | 2.857                         |
| SubZel9    | 2              | 0.286               | 5.714                         |
| Lori       | 4              | 0.421               | 6.904                         |
| Zel        | 8              | 0.478               | 7.898                         |
| Female     | 9              | 0.452               | 7.512                         |
| Male       | 4              | 0.480               | 7.471                         |

**Figure 2.** Network analysis of observed haplotypes in Zel and Lori-Bakhtiari sheep. Black is Lori-Bakhtiari breed and yellow is Zel breed.

population (Figure 2). The existence of two common haplogroups (i.e. haplotypes H<sub>1</sub> and H<sub>3</sub>) in the indigenous Iranian sheep breeds, which came from strikingly different geographical regions, suggests that these haplogroups would constitute the early haplotypes of the two breeds, while the population specific haplotypes evolved later. Since most of the genes on the

mitochondrial genome are associated with energy metabolism, and are related with adaptation to different climatic conditions (Upholt *et al.*, 1977), it seems feasible that the population specific haplotypes can be developed later due to the evolution and adaptation to different environments. As shown in Figure 2, there are four more potential haplotypes (designated mv1 to

mv4); these theoretical potential haplotypes were not observed in the studied samples, but they may exist in a wider population.

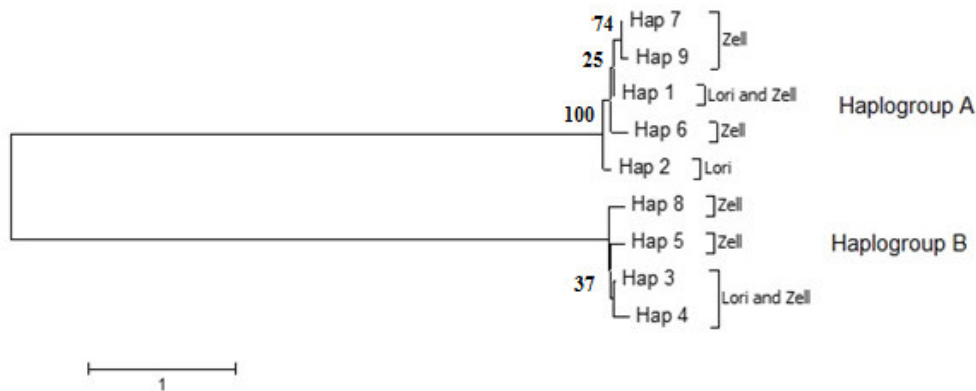
Based on the statistical results, the Lori-Bakhtiari and Zel populations could be grouped into three and nine subpopulations, respectively (Table 3). The haplotype diversity and the number of different nucleotides for all subpopulations, breeds and sexes have been shown in Table 3, however, these statistics for all animal samples were 0.45 and 7.47, respectively. Mohammadhashemi *et al.* (2012) analysed ten mitochondrial regions in Moghani sheep breed and reported a haplotype diversity of 0.3, whereas Wang *et al.* (2006) in a study on Helan Mountain sheep population in China reported that the haplotype diversity was 0.79. The results obtained in our present study are within the reported ranges. Oner *et al.* (2013) reported that the average number of different nucleotides was 7.456 in nine Turkish native breeds, which is in agreement with the results of Iranian indigenous breeds. However, the haplotype diversity in Iranian breeds is low in comparison with sheep in China (Zhao *et al.*, 2011), Asia, India, Europe (Pardeshi *et al.*, 2007) and Africa (Agaviezor *et al.*, 2012). This difference could be due to breed differences or the sampling method used in the studies.

Estimation of the genetic differentiation Fixation index ( $F_{ST}$ ) and gene flow in all ten SNP sites between Zel and Lori-Bakhtiari breeds were  $0.052 \pm 0.003$  and  $4.715 \pm 0.249$ , respectively. Lori-Bakhtiari is one of the most important Iranian fat-tailed breeds which have adapted to mountainous and valley regions in the southern part of Iran, along the Zagros Mountains. Zel is the only thin-tailed breed that is present largely on the northern part of Iran, near the Caspian Sea (Moradi *et al.*, 2012). The findings of low  $F_{ST}$  and high gene flow indicate that the different geographical areas have no barrier on the genetic structure of the populations. Different studies have been carried out to examine population differentiation and gene flow

among different breeds of sheep all over the world. Agaviezor *et al.* (2012), in a study of genetic diversity in Nigerian sheep breeds using mtDNA analysis, reported a low  $F_{ST}$  (between 0.0013- 0.0033) which is consistent with our results. Agaviezor *et al.* (2013), reported a presence of gene flow within four Nigerian sheep breeds. Similar findings were obtained by Moradi *et al.* (2012) based on SNP markers in the nuclear DNA. They reported low  $F_{ST}$  with a mean of 0.024 (SD= 0.035) using the same Zel and Lori-Bakhtiari animal samples. Kijas *et al.* (2009) studied 23 domestic and two wild sheep breeds and showed that sheep in general exhibit low differentiation, consistent with the short evolutionary history of the species. The results of our study using mitochondrial genome are consistent with the reported findings.

Analysis of Molecular Variance (AMOVA) within and between populations was conducted for all ten mtDNA SNPs. The results revealed that for one marker, about 2% of the genetic variation is found between breeds, compared to 98% variation within breeds. For the remaining nine SNP markers, the variation within and between breeds were 97 and 3%, respectively. AMOVA revealed that mtDNA diversity is mainly distributed within breeds, mainly due to the maternal inheritance of the organelles. This is in agreement with the result of low  $F_{ST}$  and high level of gene flow between populations. As it was shown earlier, both breeds are formed from two main haplogroups (haplotypes  $H_1$  and  $H_3$ ) with a relatively high frequency and these results correspond to low genetic diversity among breeds. Meadows *et al.* (2005) studied the mtDNA diversity in different Asian and European sheep breeds, and reported that sheep has the largest gene flow and weakest population structure in domestic animals, so that only 2.7 percent of the total genetic diversity is intercontinental diversity, whereas it is 10 and 50 percent for goat and cow, respectively.

Phylogenetic analysis of the Iranian animals based on mtDNA haplotypes is



**Figure 3.** Phylogenetic tree of the nine different haplotypes in Zeland Lori-Bakhtiari breeds based on the maximum likelihood method.

shown in Figure 3. The phylogenetic tree was obtained using the maximum likelihood and Bootstrap method as the best model. The numbers on the phylogenetic tree are Bootstrap percentages in the formation of branches. The phylogenetic analysis classified all animals into two mtDNA haplogroups A and B. The two haplogroups were found in both breeds. Different variants of haplogroup A gave rise to haplotypes H<sub>1</sub>, H<sub>2</sub>, H<sub>6</sub>, H<sub>7</sub> and H<sub>9</sub>; similarly, variants of haplogroup B generate H<sub>3</sub>, H<sub>4</sub>, H<sub>5</sub> and H<sub>8</sub> haplotypes. H<sub>1</sub> and H<sub>3</sub> haplotypes have the highest frequencies in both breeds, indicating that they are the ancestral types. Haplogroup A in terms of number of haplotypes and haplogroup B in terms of number of individuals has more frequency.

At least four mtDNA haplogroups have been reported in the literature (Wood and Phua, 1996; Tapio *et al.*, 2006; Pariset *et al.*, 2011). Wood and Phua (1996), in a study on the control region of mtDNA of 50 animals from five New Zealand sheep breeds, reported that all samples can be categorized into either of two haplogroups, A or B. Hiendleder *et al.* (1999) using RFLP markers analysed 239 samples from five European, three Central Asian and two African sheep breeds, and reported the same results. Lasagna *et al.* (2013) studied genetic diversity among three Italian breeds based on

mtDNA, and detected two haplogroups A and B; there was one haplotype in haplogroup A and 82 haplotypes in haplogroup B. Mariotti *et al.* (2013) analysed 138 sheep samples belonging to seven Italian breeds; based on 68 SNPs in the mtDNA control region, they detected three haplogroups in their animals. In a similar study on 406 unrelated animals of 48 sheep breeds from European, Caucasian and Central Asian regions, Tapio *et al.* (2006) identified a total of four haplogroups, indicating four maternal lineages in the modern sheep breeds. Based on the different haplogroups reported in sheep (Wang *et al.*, 2007; Zhao *et al.*, 2011), it is likely that domestic sheep are formed from at least four different lineages of wild type Mouflon (Tapio *et al.*, 2006). The presence of two mtDNA haplogroups in Iranian sheep breeds, may suggest that Iranian sheep originated from two sources of Mouflon, or from a large population containing both derivative haplogroups. However, further researches and complete mitochondrial sequencing are needed to establish the origin of the breeds.

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## تنوع هاپلوتیپی و ژنتیکی ژنوم میتوکندریایی در گوسفندان بومی ایران و کاوش تاریخچه اهلی سازی گوسفند

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### چکیده

یافته‌های باستان‌شناسی نشان می‌دهد که ایران یکی از مهدهای اصلی اهلی سازی گوسفند در جهان محسوب می‌شود. هدف از اجرای این تحقیق بررسی تنوع ژنتیکی نژادهای گوسفند بومی ایرانی با استفاده از اطلاعات ژنوم میتوکندریایی و کاوش تاریخچه تکاملی در این نژادها بود. به این منظور نشانگرهای SNP در ناحیه کنترل DNA میتوکندریایی در نمونه‌های غیرخویشاوند دو نژاد زل و لری-بختیاری (که در داخل یا نزدیکی مراکز اهلی سازی گوسفند قرار دارند) با استفاده از دستگاه Sequenom MassARRAY platform تعیین ژنوتیپ شدند. آنالیزهای فیلوژنتیکی نشان داد که جمعیت‌های گوسفند ایرانی در دو هاپلوگروپ A و B قرار می‌گیرند. آماره‌های  $F_{ST}$  و  $G_{ST}$  به ترتیب ۰/۰۵۴ و ۴/۷۱۵ بدست آمد که نشان دهنده تمایز ژنتیکی پایین و جریان ژنی بالا بین این دو نژاد است. نتایج آنالیز واریانس تنوع ژنتیکی (AMOVA) نیز نشان داد که حدود ۹۷٪ از کل تنوع ژنتیکی مشاهده شده در داخل نژادها پراکنده شده است. نتایج حاصل از آنالیزهای هاپلوتیپی مجموعاً باعث شناسایی ۹ هاپلوتیپ مختلف در بین جمعیت‌های گوسفند ایرانی شد که ۸ مورد از آنها در نژاد زل و تنها ۴ هاپلوتیپ در نژاد لری بختیاری مشاهده شد. دو هاپلوتیپ  $H_1$  و  $H_3$  دارای بیشترین فراوانی در نژادهای ایرانی بودند که دارای بالاترین فراوانی در هر دو نژاد بودند. در این تحقیق هاپلوتیپ  $H_2$  مختص نژاد لری بختیاری و هاپلوتیپ‌های  $H_5$ ،  $H_6$ ،  $H_7$ ،  $H_8$  و  $H_9$  مختص نژاد زل شناسایی شدند. وجود دو هاپلوتیپ عمده و مشترک  $H_1$  و  $H_3$  در هر دو نژاد نشان می‌دهد که این نژادها (که در دو منطقه ژئوگرافیکی کاملاً متفاوت قرار دارند) دارای جد مشترکی بوده و این هاپلوتیپ‌ها می‌توانند به عنوان هاپلوتیپ‌های مبدأ در نژادهای ایرانی مطرح باشند، درحالی‌که هاپلوتیپ‌های اختصاصی بعداً در جهت انطباق با شرایط محیطی به وجود آمده‌اند.