# Resistance of Iranian *Lecanicillium fungicola* to Benzimidazole and Ergosterol Demethylation Inhibiting Fungicides

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#### **ABSTRACT**

Dry bubble disease is one of the most important serious diseases of the cultivated white button mushroom (*Agaricus bisporus* (Lange) Imbach). It is a cosmopolitan disease having a worldwide distribution. Ten isolates of *Lecanicillium fungicola* var. *fungicola* (Preuss) Zare and Gams were collected from mushroom farms. Sensitivity of the isolates to benomyl, carbendazim, carbendazim+iprodione and prochloraz manganese were studied. All the isolates were resistant to benomyl(ED50= 415.45-748.12 mg L<sup>-1</sup>), carbendazim (ED<sub>50</sub>= 1123.87-1879.59 mg L<sup>-1</sup>) and iprodione+carbendazim (ED<sub>50</sub>= 415.45-748.12 mg L<sup>-1</sup>). However, most of the isolates were sensitive to prochloraz manganese (ED<sub>50</sub>=1.62–12.58 mg L<sup>-1</sup>). As the primary source of the pathogen inoculum is casing soil and insects, stringent environmental hygiene of the mushroom houses will play a very important role in preventing and controlling the disease.

Keywords: Agaricus bisporus, Dry bubble, Fungicide resistance, Mushroom, Sporgon.

#### INTRODUCTION

Dry bubble disease is one of the most important serious diseases of cultivated white button mushroom (*Agaricus bisporus* (Lange) Imbach). It is a cosmopolitan disease having a worldwide distribution. The causal agent of the disease is *Lecanicillium fungicola*. The pathogen causes several macroscopic symptoms on its host, including necrotic lesions, stipe blowout and undifferentiated masses called dry bubble (Largeteau and Savoie, 2008).

Although the disease control is maintained by the use of chemicals, there is a limitation in the use of fungicides because both the host and the pathogen are fungi. Currently, Iranian mushroom growing centers rely on the use of prochloraz manganese and benzimidazole groups such as benomyl and carbendazim for control of the disease. The use of fungicides from the group of benzimidazole was prevalent in the late 1960s and in the early

1970s. Also, they were applied for the control of diseases in mushroom farms worldwide. The mechanism of action of benzimidazole is the prohibition of cell division by destruction of beta tubulin of microtubules. A simple shift in amino acids sequence of beta tubulin can prevent adjoining of benzimidazole fungicides to their site of action (Smith, 1988, Chaten, 1996). Benomyl was the first benzimidazole used for mushroom diseases control (Holmes et al., 1971, Snel and Fletcher, 1971). At the first introduction of the fungicide, good control of the disease was observed, but by 1974, demonstrating fungicide several reports resistance were published (Fletcher and Yarham, 1976). In Switzerland, about one year benomyl and carbendazim registration for mushroom diseases control, these fungicides lost their ability to control dry bubble disease due to the development of resistance (Gandy and Spencer, 1976). In Australia, benomyl and thiophanate methyl

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resistant isolates were first reported in 1987 (Nair and Macauley, 1987).

In the early 1980s, prochloraz manganese was introduced to mushroom industry for the control of wet bubble disease (Mycogone perniciosa), cobweb disease (Cladobotrium denderoides) and dry bubble disease (Lecanicillium fungicola) (Van Zaayen and Van Adrichem, 1982, Fletcher et al., 1983). Prochloraz manganese belongs to imidazole group and inhibits biosynthesis of ergosterol in cytoplasmic membranes (Buchenauer, 1987). Several reports of the disease resistance to the fungicide already exist. In the study of Geels (1996), the ED<sub>50</sub> of prochloraz manganese against L. fungicola isolates was found to be 40-80 mg L<sup>-1</sup>. In England, mushroom industry also depends on fungicide application for control of the disease. Grogan et al. (2000) reported that values of ED<sub>50</sub> of this fungicide against L. fungicola isolates were in the range of 5-8 mg L<sup>-1</sup>. The ED<sub>50</sub> value of the fungicide for one isolate was 5.9 mg L<sup>-1</sup> and it was able to grow on media containing 50 mg L<sup>-1</sup>of this fungicide. The ED<sub>50</sub> value of prochloraz manganese against L. fungicola isolates was determined at 0.8-8.8 mg L<sup>-1</sup>by Gea et al. (2005). Formerly, Gea et al. (1995) reported that the ED<sub>50</sub> values of the fungicide against L. fungicola isolates were less than 5 mg L<sup>-1</sup>. However, the ED<sub>50</sub> determined with the isolates that were collected prior to the fungicide introduction in mushroom farms was 0.7 mg L<sup>-1</sup> (Gea et al., 2005). In their research, the sensitivity rates of the pathogen to prochloraz manganese decreased gradually from 1996 to 1999.

In the study of Potočnik (2006), all isolates of L. fungicola were resistant to benomyl with ED<sub>50</sub> rates higher than 200 mg L<sup>-1</sup>, the isolates were of intermediate sensitivity to iprodione, with ED<sub>50</sub> rates between 11.93–22.80 mg  $L^{-1}$ , and were more sensitive to prochloraz manganese with ED<sub>50</sub> rates lower than 3 mg L<sup>-</sup> <sup>1</sup>. The aim of this study was to investigate the sensitivity of ten Iranian isolates Lecanicillium fungicola benomyl, carbendazim,iprodione+carbendazi m and prochloraz manganese.

#### MATERIALS AND METHODS

#### **Isolation and Identification**

The isolates of *Lecanicillium fungicola*, collected from Tehran mushroom growing centers, are presented in Table 1. Isolation was done by cutting small pieces of the diseased mushroom fruit bodies, which were then sterilized by immersion in 1% sodium hypochlorite for one minute. The pieces were placed on potato dextrose agar (PDA) (Potočnik *et al.*, 2008). After isolation of single spores, the isolates were maintained on potato carrot agar (PCA) at 4°C. Isolates were identified based on taxonomic criteria determined by Zare and Gams (2008).

#### **Fungicide Bioassay**

Lecanicillium isolates were cultured on potato dextrose agar (PDA) containing the following fungicide: prochloraz manganese (Sporgon WP50 Bayer Crops Co. Germany), carbendazim (Carbendazim WP50 Ghazal Chimi Co. Iran), benomyl (Benomyl WP50 Keshavarz Chimi Iran) and iprodion+carbendazim(Iprodione+Carbendazi m WP50 Ghazal Chimi Co. Iran). Suspensions of fungicides in ethanol were added to sterile PDA at 50°C. Fungicide concentrations were expressed based on active ingredient. The concentrations of all selected fungicides were

Table 1. Lecanicillium fungicola isolates.

Isolate	Year	of	Origin	of
code	isolation		isolate	
B1	2010		Shahriar	
B2	2009		Shahriar	
R3	2009	Hashtgerd		
G5	2009	Shahriar		
G6	2009	Shahriar		
G8	2010	Shahriar		
F3	2009	Shahriar		
F6	2009	Shahriar		
P4	2009	Shahriar		
P16	2009		Mohammadsh	nahr

as follow:

Prochloraz manganese: 0.1, 0.5, 1.0, 2.5, 5.0 10.0, 20.0, 50, 100 mg L<sup>-1</sup>; carbendazim, benomyl and iprodion +carbendazi: 1, 10, 20, 50, 100, 200, 500, 1,000, 1,500, 2,000, 2,500 and 3,000 mg L<sup>-1</sup>. Five mm mycelial agar disks were used as inoculum and centrally placed on both fungicide amended media and the control media and then incubated at 20°C in total darkness. For each treatment, three replicates were used. Colony diameter minus plug diameter was calculated after 14 days of inoculation and growth of colonies was used as a percentage of the control for ED<sub>50</sub> calculation i.e. fungicide concentration inhibiting radial mycelia growth by 50%.

The median effective dose (ED<sub>50</sub>) for each fungicide and isolate was calculated by interpolation from computer-generated log-probit plots (Probit SPSS version 16) of fungicide concentrations and relative growth inhibition: 100 (Colony diameter on amended medium/Colony diameter on control)×100. The means were separated by the lower and upper bound of each isolate, which were considered as factors in the analysis.

#### **RESULTS**

#### **Isolation and Identification**

Conidiophores were differentiated from by subtending hyphae being without secondary branches, typically erect, and procumbent with aging. Conidia were produced in globose heads. Octahedral crystals were present, hence all the isolates were identified as Lecanicillium fungicola. All the isolates grew at 24°C and no growth was observed at 30°C and, therefore, they belonged to variety fungicola (L. fungicola var fungicola).

#### **Fungicide Bioassay**

The  $ED_{50}$  value of benomyl and carbendazim fungicide against *L. fungicola* isolates was varied between 415.45-748.12

mg L<sup>-1</sup> and 1123.87-1879.59 mg L<sup>-1</sup>. respectively (Table 2). The ED<sub>50</sub> values of iprodione+carbendazim prochloraz and manganese against isolates were between 70.88-486.01 mg L<sup>-1</sup> and 1.62–12.58 mg L<sup>-1</sup>, respectively (Table 3). For benomyl, iprodione+carbendazim and prochloraz manganese, statistically significant differences were found between the ED<sub>50</sub> values of different isolates based on the lower and upper bound. None of the isolates were significantly different in their response to carbendazim and none grew on media containing 50 mg L<sup>-1</sup> prochloraz manganese, except isolate P12. All of the isolates were able to grow on media containing benomyl and carbendazim at concentration of 3,000 mg L<sup>-1</sup>.

#### **DISCUSSION**

The ED<sub>50</sub> range of benomyl against isolates of *L. fungicola* was between 415.45-748.12 mg L<sup>-1</sup>. All the isolates were resistant to benomyl. The isolate most resistant to this fungicide was P4 with the ED<sub>50</sub> of 747.12 mg L<sup>-1</sup>. Nevertheless, P4 isolate was the most sensitive to carbendazim in comparison to the other isolates.

The  $ED_{50}$  ranges of iprodione+carbendazim and prochloraz manganese against the tested isolates were between 70.88-486.01 and 1.62–12.58 mg  $L^{-1}$ , respectively. The isolate most resistant to prochloraz manganese was P4, with the  $ED_{50}$  of 12.58 mg  $L^{-1}$ .

Isolate P4 was different from other isolates in sensitivity to prochloraz manganese, benomyl and carbendazim. This isolate was recognized as one of the most resistant to both benomyl and prochloraz manganese and the most sensitive isolate to carbendazim.

Benzimidazole fungicides were used for the control of mushroom diseases in 1970s for the first time and they controlled diseases effectively early in their introduction (Holmes *et al.*, 1971, Snel and Fletcher, 1971). But, there is a long time since the resistance of *L. fungicola* to carbendazim



**Table 2.** ED<sub>50</sub> values of benomyl and carbendazim for isolates of *Lecanicillium fungicola* var. *fungicola*.

Isolate code	ED <sub>50</sub>	Lower bound	Upper bound	Slope	<i>P</i> -value
		ber	nomyl		
B1	569.71 abc	423.70	801.93	0.72±0.07	0.24
B2	401.32 abc	267.47	699.72	$0.73 \pm 0.09$	0.19
R3	451.68 ab	358.81	577.73	1.04±0.08	0.38
G5	482.61 ab	390.42	607.08	1.03±0.07	0.70
G6	497.89 abc	390.99	650.70	$0.89 \pm 0.06$	0.11
G8	633.88 ac	528.64	769.23	1.31±0.10	0.39
F3	415.45 a	329.31	534.96	1.05±0.11	0.33
F6	682.64 bc	570.51	828.19	1.34±0.10	0.20
P4	748.12 c	617.77	898.78	1.46±0.17	0.19
P16	513.21 abc	409.41	656.36	$0.97 \pm 0.08$	0.23
		carbo	endazim		
B1	1334.32 a	1180.22	1534.41	2.44±0.31	0.30
B2	1330.13 a	1099.16	1666.21	1.46±0.12	0.36
R3	1818.18 a	1481.17	2418.29	1.58±0.20	0.35
G5	1879.59 a	1353.93	2949.89	0.91±0.10	0.83
G6	1423.84 a	977.36	2358.56	$0.69 \pm 0.07$	0.72
G8	1395.80 a	1126.43	1801.33	1.38±0.12	0.79
F3	1245.70 a	1021.86	1577.10	1.34±0.12	0.92
F6	1285.18 a	955.14	1851.41	$0.99 \pm 0.08$	0.82
P4	1123.87 a	823.92	1661.83	$0.79 \pm 0.08$	0.88
B1	1334.32 a	1180.22	1534.41	$2.44 \pm 0.31$	0.30

 $ED_{50}$ = Fungicide concentration which inhibits mycelial growth by 50%, Slope= Regression coefficient at the 95% confidence level.

The ED<sub>50</sub> followed by different letters are significantly different based on lower and upper bound.

**Table 3.**  $ED_{50}$  values of iprodion+carbendazim and prochloraz manganese for isolates of *Lecanicillium fungicola* var. *fungicola*.

Isolate code	$ED_{50}$	Lower bound	Upper bound	Slope	<i>P</i> -value
		iprodion+	carbendazim		
B1	393.99 de	275.79	626.89	0.73±0.08	0.23
B2	270.22 cde	185.01	436.88	$0.63\pm0.09$	0.39
R3	486.01 e	354.18	728.35	$0.89 \pm 0.09$	0.36
G5	191.16 bcd	130.33	303.20	$0.58 \pm 0.07$	0.81
G6	70.88 a	44.30	107.59	$0.52\pm0.07$	0.64
G8	366.60 de	269.12	539.31	$0.84 \pm 0.08$	0.81
F3	396.48 de	278.31	637.73	$0.74 \pm 0.10$	0.16
F6	109.58 ab	74.76	166.95	$0.55 \pm 0.05$	0.97
P4	202.31 bc	158.80	264.77	$0.97 \pm 0.07$	0.28
P16	130.34 abc	90.93	195.06	$0.59\pm0.06$	0.37
		prochlora	z manganese		
B1	2.62 abc	2.01	3.75	1.13±0.17	0.69
B2	1.62 a	1.27	2.08	1.25±0.17	0.55
R3	2.77 bc	2.21	3.72	1.37±0.18	0.79
G5	1.98 ab	1.62	2.49	1.47±0.18	0.32
G6	4.75 c	3.46	6.53	$0.64 \pm 0.05$	0.37
G8	2.43 ab	2.00	3.05	1.59±0.19	0.43
F3	2.06 ab	1.62	2.74	1.21±0.17	0.43
F6	2.40 ab	2.02	2.92	1.82±0.19	0.10
P4	12.58 d	9.97	16.17	0.94±0.06	0.93
P16	1.86 ab	1.52	2.32	1.47±0.18	0.667

 $ED_{50}$ = Fungicide concentration which inhibits mycelial growth by 50%, Slope= Regression coefficient at the 95% confidence level.

The ED<sub>50</sub> followed by different letters are significantly different based on lower and upper bound.

and benomyl was reported (Fletcher and Yarham, 1976; Gandy and Spencer, 1976; Nair and Macauley, 1987).

Gea et al, (1996) reported that the values of the ED<sub>50</sub> of benomyl, chlorotalonil and iprodione against isolates of L. fungicola were higher than 50 mg L<sup>-1</sup>. In a study by Potočnik (2006), all of the isolates were highly resistant to benomyl, with the EC<sub>50</sub> values exceeding 200 mg L<sup>-1</sup>, moderately sensitive to iprodione, having values of EC<sub>50</sub> between  $11.93-22.80 \text{ mg L}^{-1}$ . In comparison with these investigations, our bioassay results showed that the Iranian L. fungicola isolates were very resistant to benomyl, carbendazim, and iprodione+carbendazim.

Isolates collected from Iranian mushroom growing industry had background of benomyl and carbendazim use for disease control in the cropping system. Thus, cross resistance studies were impossible. Most likely reasons for the appearance of high ED<sub>50</sub> values of selected fungicides for Iranian L. fungicola isolates were attributed to rambunctious use of these fungicides in mushroom industry that caused population of L. fungicola to adapt to these fungicides. All isolates were able to grow on media containing benomyl and carbendazim at 2,000, 2,500 and 3,000 mg L<sup>-1</sup>, which is in accordance with Gams and Van Zaayen (1982) reports.

Prochloraz manganese was introduced to mushroom industry for dry bubble disease control in late 1980s. In the beginning, it effectively controlled *Agaricus bisporus* diseases. But over time, its effectiveness on the fungus diminished. Lately, decreased sensitivity of the pathogen to this fungicide has been reported worldwide (Geels, 1996; Grogan *et al.*, 2000; Bernardo *et al.*, 2002; Gea *et al.*, 2005; Allan *et al.*, 2008).

In the present investigation, fungicide bioassay results of prochloraz manganese and *Lecanicillium fungicola* isolates showed that most of the isolates were highly sensitive to this fungicide. Among these isolates, isolate P4 with the ED<sub>50</sub> of 12.58 mg L<sup>-1</sup>was weakly resistant to this fungicide.

Nonetheless, benomyl, carbendazim, and iprodione+carbendazim are still being used for the control of mushroom diseases, especially dry bubble disease in Iran, without any good results. Hence, it is necessary that mushroom growers do not use these fungicides for the control of dry bubble disease. Prochloraz manganese is the only effective fungicide for dry bubble disease control. It is important to decrease the progress of pathogen resistance to this fungicide using integrated disease management items, such disease monitoring and steam pasteurizing the spent mushrooms substrate after crop termination.

The pathogen shift to a fungicide tolerant population has been proved in several research (Bonnen and Hopkins, 1997; Gea et al., 2005). In the present investigation, fungicides bioassay tests were done on isolates collected recently, but isolates from before introduction of the studied fungicides to Iranian mushroom industry were not available. Therefore, we were not able to confirm population shift to a higher tolerance level to this fungicides.

These information show that the fungicide has gradual selective pressure on the pathogen population and, within some years, the sensitivity rate has been shifted from low to intermediate. In the situation of fungicide use for a given disease control, resistant isolates can better compete with the sensitive ones and the population of isolates resistant to the fungicide increases. Thus, the use of fungicide induces a special selection so that, after some years, there will not be sensitive population of the pathogen and its genetic constitution will be changed to a more clonal population (Largeteau *et al.*, 2006; Largeteau *et al.*, 2008).

The observation of possible failure of Sporgon to control *Agaricus bisporus* dry bubble disease could be a terrible hazardous for Iranian mushroom industry. With the continuation use of this fungicide for the control of dry bubble disease as the single disease control strategy, the pathogen population will be shifted to a more tolerant one to prochloraz-manganese than previous.



Hence, mushroom growers will not be able to rely on fungicides for dry bubble disease control as the sole tactic in control procedure and should use integrated disease management for further controlling actions.

#### REFERENCES

- Allan, J., Shah, F. A. and Khan, I. 2008 Establishing a Baseline for Fungicide Sensitivity of Three Major Mushroom Pathogens in Australia. *Mushroom Sci*, XVII(I): 565-569.
- Bernardo, D., Novaes-Ledieu, M., Perez Cabo, A., Gea Alegría, F. J. and García Mendoza, C. 2002. Effect of the Fungicide Prochloraz-Mn on the Cell Wall Structure of Verticillium fungicola. Int. Microbiol., 5: 121-125.
- 3. Bonnen, A. M. and Hopkins, C. 1997. Fungicide Resistance and Population Variation in *Verticillium fungicola*, a Pathogen of the Button Mushroom, *Agaricus bisporus*. *Mycol. Res.*, **101**: 89-96.
- Buchenauer, H. 1987. Mechanism of Action of Triazolyl Fungicides and Related Compounds In: "Modern Selective Fungicides: Properties, Applications, Mechanisms of Action", (Ed.): Lyr, H.. John Wiley and Sons, New York, PP. 205-232.
- 5. Chaten, C. E. 1996. The Mutable and Treacherous Tribe Revisited. *Plant Pathol.*, **45**: 1-12.
- Fletcher, J. T., Hims, M. J. and Hall, R. J. 1983. The Control of Bubble Diseases and Cobweb Disease of Mushrooms with Prochloraz. *Plant Pathol.*, 32: 123-131.
- 7. Fletcher, J. T. and Yarham, D. J. 1976. The Incidence of Benomyl Tolerance in *Verticillium fungicola, Mycogone perniciosa* and *Hypomyces rosellus* in Mushroom Crops. *Ann. Appl. Biol.*, **84**: 343-353.
- 8. Gams, W. and Van Zaayen, A. 1982. Contribution to the Taxonomy and Pathogenicity of Fungicolous *Verticillium* species. I. Taxonomy. *Neth J. Plant Pathol.*, 88: 57-78.
- 9. Gandy, D. G. and Spencer, D. M. 1976. The Use of Chlorothalonil for the Control of Benzimidazole Tolerant Strains of *Verticillium fungicola* (Preuss) Hassebr. on the Cultivated Mushroom. *Sci* . *Hortic.*, 5: 13-21.

- Gea, F. J., Navarro, M. J. and Tello, J. C. 2005. Reduced Sensitivity of the Mushroom Pathogen *Verticillium fungicola* to Prochloraz-manganese *In vitro*. *Mycol. Res.*, 109: 741-745.
- Gea, F. J., Pardo, A., Navarro, M. J. and Pardo, J. 1995. Fungal Diseases of Mushroom Culture from Castilla-La Mancha (Spain): Incidence of *Verticillium fungicola*. *Mushroom Sci.*, **15(1 and 2)**: 643-651.
- 12. Gea, F. J., Tello, J. C. and Honrubia, M. 1996. *In vitro* Sensitivity of *Verticillium fungicola* to Selected Fungicides. *Mycopathol.*, **136**: 133-137.
- Geels, F. 1996. Resistance to Sporgon Tested Among Recently Isolated Strains of Verticillium fungicola var. fungicola. De Champignoncultuur (Netherlands), 40: 401-406.
- 14. Grogan, H., Keeling, C. and Jukes, A. 2000. In Vivo Response of the Mushroom Pathogen Verticillium fungicola (dry bubble) to Prochloraz-manganese. The BCPC Conference: Pests and Diseases. 13-16 November 2000. Proceedings of an International Conference Held at the Brighton Hilton Metropole Hotel, Brighton, British Crop Protection Council. , UK. 1: 273-278.
- 15. Holmes, J. H., Cole, H. and Wuest, P. J. 1971. Control of the *Verticillium* Disease of the Cultivated Mushroom, *Agaricus bisporus* with Benomylspray Applications. *Plant Dis. Rep.*, **55**: 684-7.
- Largeteau, M. L., Baars, J. P. P., Regnault-Roger, C. and Savoie, J. M. 2006. Molecular and Physiological Diversity Among Verticillium fungicola var. fungicola. Mycol. Res., 110: 431-440.
- Largeteau, M. L., Mata, G. and Savoie, J.-M. 2008. Verticillium fungicola var. fungicola: Comparison of Some Mexican and French Isolates. Revista Mexicana de Micologia, 26: 35-40.
- Largeteau, M. L. and Savoie, J. M. 2008. Effect of the Fungal Pathogen Verticillium fungicola on Fruiting Initiation of Its Host, Agaricus bisporus. Mycol Res., 112: 825-828.
- 19. Nair, N. and Macauley, B. 1987. Dry Bubble Disease of *Agaricus bisporus* and *A. bitorquis*, and Its Control by Prochloraz-Manganese Complex. *Newzeal. J. Agr. Res.*, **30:** 107-116.

- 20. Potočnik, I. 2006. Causal Agents of Bubble Diseases of White Button Mushrom (Agaricus bisporus (Lange) Imbach) and Their Sensitivity to Fungicides. Faculty of Biology, University of Belgrade, Serbia and Montenegro.
- Potočnik, I., Vukojević, J., Stajić, M., Tanović, B. and Todorović, B. 2008. Fungicide Sensitivity of Selected Verticillium fungicola Isolates from Agaricus bisporus Farms. Arch. Biol. Sci., 60: 151-157.
- 22. Smith, C. M. 1988. History of Benzimidazole Use. In Fungicide Resistance

- in Northern America. *The Am. Phytopathol. Soc.*, **43**: 23-24.
- 23. Snel, M. and Fletcher, J. T. 1971. Benomyland Thiabendazole for the Control of Mushroom Diseases. *Plant Dis.Rep.*, **55**: 120-121.
- 24. Van Zaayen, A. and Van Adrichem, J. C. J. 1982. Prochloraz for Control of Fungal Pathogens of Cultivated Mushrooms. *Neth. J. Plant Pathol.*, **88**: 203-213.
- 25. Zare, R. and Gams, W. 2008. A Revision of the *Verticillium fungicola* Species Complex and its Affinity with the Genus *Lecanicillium. Mycol. Res.*, **112**: 811-824.

## مقاومت Lecanicillium fungicola ایران به قار چکشهای گروه بنزیمیدازول و بازدارنده دمتیلاسیون ارگوسترول

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#### چكىدە