Diversity and Population Structure Analysis of Faba Bean (Vicia faba L.) Accessions Using SSR Markers

N. A. Tahir1*, D. A. Omer2, D. D. Lateef3, D. A. Ahmad3, S. H. Salih3, and L. Hiwa Khal1

ABSTRACT

The awareness and conception of the genetic diversity in faba bean (Vicia faba) accessions are important for the enforcement of degree addressed to their usages and conservations. The aim of this work was to estimate the genetic diversity and population structure in Iraqi faba bean using SSR markers for utilization in crossing and variety development. To assess genetic variation and population structure among faba bean accessions, 25 microsatellite loci were exerted. The analysis of diversity indices in the set of faba bean accessions examined here showed that the microsatellites were informative for genotype characterization. In total, 72 polymorphic alleles were exposed to an average of 2.88 per locus and three unique alleles were detected. The average of PIC, gene diversity, marker index, resolving power and Shannon diversity was 0.513, 0.569, 1.671, 2.173 and 0.830, respectively. The patterns detected in the dendrogram and PCA divided 19 accessions into five distinct clusters with different levels of sub-grouping within the cluster. High-level genetic differentiation within a population or group (83%) was significantly greater than that among groups or populations (17%), as planned by Analysis Of Molecular Variance (AMOVA). The model of clustering, based on the analysis of STRUCTURE software, identified four groups genetically dispersed. These findings have additional importance in faba bean breeding as well as maintenance programs.

Keywords: Genetic variation indices, Genetic structure, Legume, Microsatellites.

INTRODUCTION

Faba bean (Vicia faba L.), also called broad bean, and horse bean originated in the Mediterranean-West Asia region (Tanno and Willcox, 2006). The today faba bean has an important role in human food and animals feed worldwide because of its contents of protein (27-34%), starch (45%) and essential vitamins (Mejri et al., 2012). Faba bean is a diploid species (2n = 2x = 12 chromosomes). The size of genomic DNA approximately is 13,000 Mb (Ellwood et al., 2008). In Iraq, faba bean is one of the fundamental legume plants grown mostly under rain-fed conditions. The unreliability of the yield is largely due to the lack of adequate genetic tool and feasible resistance/tolerance to the prevailing diseases (Ascochyta), pests, and drought. Hence, it is crucial to evolve new accessions that are able to tolerate the biotic and abiotic stresses accountable for low productivity in Iraq. The display of modern genetic tools with improved agronomic traits and increased genetic variation leads to the expansion of high-yielding accessions (Hwang et al., 2008). In this circumstance, a convenient description of the genetic divergence at the morphology and molecular levels in the accessible local faba bean accessions resource is indispensable for additional improvement of yield and stress.

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tolerance. The agro-morphological traits are unsuitable to the development of gene pools because these traits are affected by environmental factors and stage of plant development, or they demonstrate merely limited diversity (Terzopoulos and Bebeli, 2008). In faba bean, several molecular techniques have been profitably employed to appraise the genetic relationship, such as isozyme (Mancini et al., 1989; Tahir, 2015), Restriction Fragment Length Polymorphism (RFLP) (Torres et al., 1993), Inter-Simple Sequence Repeat (ISSR) (Salazar-Laureles, 2015), Amplified Fragment Length Polymorphism (AFLP) (Zong et al., 2009), Randomly Amplified Polymorphic DNA (RAPD) methods (Tair, 2015), target region amplification polymorphism (TRAP) (Kwon et al., 2010), Single Nucleotide Polymorphism (SNP) (Kaur et al., 2014) and Simple Sequence Repeats (SSRs) (Kaur et al., 2012). SSR markers are often applied in plant diversity and population structure because they are easy, PCR-based, highly polymorphic, and can be used to differ between closely related accessions. To develop V. faba accessions, an introductory description, estimation, and comparison of the genetic variation among regional and ICARDA accessions could be helpful for the imposition of this genetic variability in breeding programs. The aim of this study was to evaluate the genotypic diversity, relatedness and population structure among local V. faba resources used in Iraq and the cultivated ICARDA accessions based on SSR markers.

MATERIALS AND METHODS

Genotype Collection

A total of 19 faba bean (Vicia faba L.) accessions, including eight local and eleven ICARDA genotypes, was investigated in the present work (Table 1), these accessions were gathered from the Agriculture Research, Ministry of Agriculture, Sulaimani, Kurdistan region, Iraq.

DNA Extraction and Electrophoresis

Genomic DNA was isolated from faba bean fresh leaves by the CetylTrimethylAmmonium Bromide (CTAB) procedure (Doyle, 1990). The

<table>
<thead>
<tr>
<th>No.</th>
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</tr>
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<tbody>
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<td>Zaffaron</td>
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</tr>
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<td>Local check</td>
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<td>G9</td>
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<td>ICARDA-Lebanon</td>
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<td>19</td>
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<td>Giza-3</td>
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</table>
quality and concentration of the extracted DNA were determined by 1% (w/v) agarose gels quantified using a Bio-Rad gel imaging system. The extracted genomic DNA was diluted to 70 ng μL\(^{-1}\) and stored at −20°C.

**Molecular Marker (SSR) Assay**

Out of 30 markers (Zeid et al., 2009, Yang et al., 2012, Suresh et al., 2013), 25 primers were successfully amplified and utilized for diversity and structure studies (Table 2). PCR reaction was conducted in a 20 μL reaction containing 1X PCR buffer, 250 mM dNTPs, 0.30 μL of primer, 3 mM MgCl\(_2\), 1 μL Taq polymerase and 70 ng template DNA. PCR amplification was conducted using Applied Biosystems Thermocycler following the PCR protocol: Initial denaturation at 94°C for 8 minutes, 38 cycles at 94°C for 1 minute, T°C (T°C depending upon the primer pair) for 1 minute, 72°C for 2 minutes and a final extension step at 72°C for 7 minutes. Amplified products were resolved on 2.5% agarose gels at 80 V in 1X TBE buffer, and fragment sizes were determined by 100-bp DNA ladder (Invitrogen, USA). The size of bands was scored using the GelAnalyzer software.

**Scoring and Statistical Data Analysis**

The scorable bands were coded manually as either present (1) or absent (0). Scored data were applied to the calculation of Jaccard’s similarity coefficient using XLSTAT 11 software. The Jaccard’s coefficient was converted to a dissimilarity matrix to create dendrogram using the Unweighted Pair-Group Method with Arithmetic averages (UPGMA) by using XLSTAT 11 software. Polymorphism Information Content (PIC), gene diversity, frequency of the major allele, marker index, resolving power and Shannon's diversity index were measured (Table 3). The SSR markers proved to be reproducible and informative polymorphisms. The results obtained are presented in Tables 3. The total numbers of amplified and polymorphic bands were recorded according to clarity and their

**RESULTS AND DISCUSSION**

**Allelic Variation in Faba Bean Accessions Using SSR Markers**

Out of 30 primers, 25 primers revealed amplified fragments. The lack of amplification of some microsatellites primer was due to an annealing failure of primer. This annealing defeat can be due to differences in the base sequences of the loci between faba bean accessions. For each SSR primer, total number of amplified bands, number of polymorphic bands, percentage of polymorphic bands, number of monomorphic bands, size of amplified bands, number of unique bands, Polymorphism Information Content (PIC), gene diversity, frequency of the major allele, marker index, resolving power and Shannon's diversity index were measured (Table 3). The SSR markers proved to be reproducible and informative polymorphisms. The results obtained are presented in Tables 3.
Table 2. Primer names, sequences, motif and annealing temperature of SSR markers used in this work.

<table>
<thead>
<tr>
<th>No</th>
<th>Primer name</th>
<th>Sequence: 5’-3’</th>
<th>Motif</th>
<th>Annealing temperature (°C)</th>
</tr>
</thead>
</table>
| 1  | CAAS1       | F: AGTCAGGGGGTGCATTTTTC  
R: TCITGCGCAATTTTGGACATC | (AAAGGG)7 | 55 |
| 2  | CAAS2       | F: TACAAAGCTCTGGGGCCTA  
R: ATATGCGCCCAAAAGATGT | (GAA)9 | 56 |
| 3  | CAAS4       | F: ATGGCAAGTGCTAGGCCAAG  
R: ATATGCGCCCAAAAGATGT | (CA)11 | 55 |
| 4  | CAAS5       | F: TACATGCTCCGGCAATATT  
R: CCAGCCAGCCATTTCCACCT | (ACA)15 | 55 |
| 5  | CAAS6       | F: TGGAAATTAATCCGAAACA  
R: CACACATGAAATGGGTAAT | (A)10 | 56 |
| 6  | CAAS7       | F: GACCCAGGTCTTCACACAA  
R: TGGTGGGATCCATTTTGAA | (A)10 | 60 |
| 7  | CAAS8       | F: ATTGTCAGAGGACTTGCGG  
R: CTAAGGGTACGCTGGTAGT | (AAG)6 | 56 |
| 8  | CAAS9       | F: AGTGACGGTGTAGGTTGGGA | (AAG)8 | 56 |
| 9  | CAAS11      | F: TCCCGCTATTCTTGCTCTGT  
R: GCCTAACCAAGTGCTTTTCA | (ACA)10 | 55 |
| 10 | CAAS12      | F: GAGAGGATCCACCACATGA  
R: GGCAAAGAGCCATGGTACA | (TGT)9 | 56 |
| 11 | CAAS14      | F: CCCTAGATCTCAAAAACATGA | (ACA)6 | 60 |
| 12 | CAAS15      | F: AACCAACATCAATGGGACT  
R: GCTATTTCTTTTCTTCCTCA | (AAG)8 | 56 |
| 13 | CAAS16      | F: TCAATTTTCCCCCCTTGCAAATA  
R: GCCAAGGTCACACCCACCTT | (CA)7 | 55 |
| 14 | GBSSR-VF-38 | F: ACCATTGGGCTGTTTCT  
R: CCCTACCAAAATGGTGGA | (GTG)6 | 58 |
| 15 | GBSSR-VF-164 | F: ACCATTGGGCTGTTTCT  
R: CAGAGGGAAGGTTGTTACGA | (CAG)6 | 60 |
| 16 | GBSSR-VF-311 | F: GGCCTTTCCAAAAGAGGG  
R: ACCATTGGGCTGTTTCT | (GTG)6 | 58 |
| 17 | VfG1        | F: TTTGCAAAAATAGAACAATATC  
R: GCCATTCAGGTTCACCTTGTA | (AG)15 | 50 |
| 18 | VfG3        | F: TTTTGGTGCTCTCTCTCCTCTAC  
R: GCACGTGTGTGCTGATACAA | (AG)7 | 50 |
| 19 | VfG4        | F: AAGGGGAGGGGCAATAACAGAA  
R: AATCCGCAAGGGTCCTTTT | (AG)5 | 50 |
| 20 | VfG9        | F: GGTGTTAGAATAGAATAATGG  
R: AAGATGTTGCAATATTTGTTT | (AG)16 | 50 |
| 21 | VfG10       | F: ACCAAAAAGGCTCTTTATCTCA  
R: AAGAGAGAGAAGAGAGCTTTC | (AG)5 | 50 |
| 22 | VfG11       | F: GCAAAAGGGGACATTTTTGT  
R: CAAAGGAGGGGCAACCTTTT | (AG)8 | 50 |
| 23 | VfG13       | F: GTGGGAGATTTATTGAGGGA  
R: TGGCTTATATCCGCTACAAT | (AG)10 | 50 |
| 24 | VfG19       | F: AGCGATGGTGTGCTATGCTTA  
R: TCTCTCAAGGAAATCCACCTT | (AG)9 | 50 |
| 25 | VfG27       | F: CCCAAAAAGAGAGCCACTGAT  
R: AGGGGTTTACACGTGGCTT | (AG)9 | 50 |
Table 3. Genetic diversity indices from SSR primer for screening 19 accessions of faba bean. 

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<th>Primer name</th>
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<th>NPB</th>
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<th>NMB</th>
<th>Size (bp)</th>
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<th>PIC</th>
<th>GD</th>
<th>MAF</th>
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<td>0.474</td>
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<td>0.714</td>
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<td>2</td>
<td>0</td>
<td>2</td>
<td>0.823</td>
<td>0.842</td>
<td>0.889</td>
<td>4.939</td>
<td>4.737</td>
<td>1.822</td>
</tr>
<tr>
<td>Total Average</td>
<td>3.280</td>
<td>2.880</td>
<td>87.000</td>
<td>0.400</td>
<td>0.120</td>
<td>0.513</td>
<td>0.569</td>
<td>0.543</td>
<td>1.671</td>
<td>2.173</td>
<td>0.830</td>
<td></td>
</tr>
</tbody>
</table>

*Total Number of Amplified Bands, **Number of Polymorphic Bands, ***Percentage of Polymorphic Bands, ****Number of Monomorphic Bands, *****Size of Amplified Bands, ******Number of Unique Bands, *******Polymorphism Information Content, ********Gene Diversity, *******Frequency of the Major Allele, ********Marker Index, **********Resolution Power and *******Shannon diversity index.
molecular weight concerning the DNA Ladder. A total of 82 reproducible alleles was detected with a molecular weight ranging from 50 to 800 bp along the different faba bean accessions. Twenty-five SSR primers amplified 72 polymorphic bands. The number of alleles varied from 1 (GBSSR-VF-311 and VfG1 and VfG10) to 6 (CASS2 and VfG19) with a mean of 2.880 by locus. Among 25 primers used, only VfG1 (1 band) and VfG11 (2 bands) showed unique band. A singular band is a band amplified in an accession, but not in the rest of accessions. It is useful in the marker-assisted selection. The considerable number of mean detectable and polymorphic fragments might be due to the sequence length variations of the primers used in this study. The difference in the number of alleles amplified by various primers is affected by primer sequence and fewer number of annealing locations in the genome. Comparing our results with previous studies, we found that the number of alleles per locus in our results (3.28) was inferior to that obtained by Rebaa et al. (2017), which showed 6.62 alleles per locus. Abid et al. (2015) obtained 101 polymorphic bands, and a range from 2 to 10 alleles per primer, with a mean of 5.94 alleles per marker on the screening of forty-six faba bean accessions through the 17 SSR loci.

According to Zeid et al. (2009), 53 polymorphic bands were detected during the discriminating of 10 faba bean accessions by using 73 SSR markers. The number of amplified fragments is comparable to those described for faba bean cultivars in other countries such as Korea. Certainly, the number of amplified bands for the SSR loci publicized by Suresh et al. (2013) ranged from 2 to 15. In addition, the mean number of alleles (3.28) was similar compared with the mean number of alleles per locus (3.37) for 53 faba bean accessions from Portugal, Spain, and Morocco with 28 SSR primers. (Oliveira et al., 2016). The variation in the number of alleles per locus between our results and previous researches, can be explained by the difference in the used genotypes and primers.

The markers with many alleles are considered as highly informative and powerful markers. The power of marker can be quantitatively measured by several methods of statistics called the Polymorphism Information Content (PIC), Marker Index (MI), Resolving Power (RP), and Shannon's diversity index (H), which were used to assess the information of the SSR primers in this work. Primers with a diversity of statistics index values of zero or less than zero should not be used for analysis, due to lesser degree of information for gene diversity. The greatest value of the diversity index intend to a higher degree of polymorphism of the SSR markers and, therefore, helped to select the best SSR loci in the genetic divergence analysis. PIC value varied from 0.178 (VfG10) to 0.823 (VfG19) with a mean of 0.513, gene diversity ranged from 0.198 (VfG10) to 0.842 (VfG19) with an average of 0.569, the frequency of the major allele aligned from 0.211 (VfG19) to 0.889 (VfG10) with a mean 0.543 across all accessions (Table 3). The average values of marker index, resolving power, and Shannon index (H) was 1.671, 2.173 and 0.830, respectively. Primer VfG19 was very informative due to its high ability to detect divergence among faba bean accessions. The values of diversity indices designate the discriminatory power of the used SSR primer to distinguish the accessions. The value of genetic diversity of this work is superior to that reported by Gol et al. (2016), who studied the genetic diversity in faba bean accessions from Turkey.

Several previous studies demonstrated the power of SSR in the discrimination of faba bean. The PIC values discovered are similar to those reported for faba bean cultivars in other countries. Indeed, the PIC values for the SSR loci exhibited by Suresh et al. (2013) ranged from 0.16 to 0.88, by Zeid et al. (2009) varied from 0.16 to 0.72, by Gong et al., 2011 reached from 0.064 to 0.428, by Abid et al. (2015) varied from 0.38 to 0.84,
and by Oliveira et al. (2016) extended from 0.071 to 0.662.

**Clustering and Principal Component Analyses (PCA)**

Clustering and main component analyses were applied for assessing the linkage between faba bean accessions. Jaccard similarity coefficients were between 0.314 (G9 vs. G11) to 0.891 (G6 vs. G15), signifying an agreeable representation of the genetic distance among the nineteen faba bean accessions. Our result is in accordance with the reporting of Kumari et al. (2013) that indicated the existence of a high degree of genetic variation, which ranged from 0.11 to 0.73 among the twenty-eight accessions. In the dendrogram constructed by Jaccard similarity coefficients using unweighted pair-group method (UPGMA), all 19 accessions clustered into five clusters (A, B, C, D, and E) with a mean similarity of 0.410 (Figure 1). Cluster A included G6, G7, and G8. Cluster B comprised G17. Cluster C included G1, G2, G3, G4, and G5. Cluster D consisted of G18, and the remaining seven accessions belonged to cluster E. Within the group, two, four, and five subgroups were detected in groups A, C, and E, respectively. Interestingly, all ICARDA faba bean accessions were observed in group A, B, D, and E. Indeed, accessions 17 and 18 from ICARDA were quite distant from the other accessions, suggesting a high level of genetic diversity.

To better clarify the association between the accessions, PCA was carried out using the similarity data set (Figure 2). The biplot graph of the two first Components (PC1 and PC2) displayed 19.29 and 13.04% of the total distinction, showing a cumulative variance of 32.33%. A high degree of overlapping appeared in the scatter plot that arranged accessions into distinctive clusters (Figure 2). PCA classified nineteen faba bean accessions into five dissimilar clusters. PCA output was congruent with the results generated by UPGMA clustering (Figure 1). In clustering and PCA, G17 and G18 were farthest from the remaining accessions, and...
as such, they showed the lowest similarity coefficient values. Moreover, PCA analysis showed that all local faba bean accessions were clustered separately from the accessions of another origin. The regional accessions (red color) belonging to the A and C groups (as inferred by UPGMA clustering) were distributed to the right of the plot. The ICARDA accessions (green color) of four groups (B, C, D, and E) were placed in the left section of the plot.

The analysis of AMOVA revealed significant levels of genetic variation (P<0.01) among populations (local and ICARDA) and individuals within populations. AMOVA of the two sub-populations demonstrated that most of the variation occurred in the intra-sub-populations (83% of the total variation), and only 17% could be ascribed to differences between sub-populations (Table 4). This result also confirmed the existing of great variation within populations. Oliveira et al. (2016) analyzed the variance among cultivated types of faba bean through SSR loci showing the highest proportion of genetic distance (36%) within accessions and not among different accessions (10%). An AMOVA analysis through SSR marker was utilized for 35 Northern Africa, Eastern Africa, and Near East, faba bean germplasms detecting a low proportion of genetic difference within populations (75.4%) compared to our results (El-Esawi, 2017).

**Genetic Structure in Faba Bean Accessions**

In order to collect information about population structure in faba bean accessions

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**Table 4.** Analysis of molecular variance (AMOVA) of the two sub-populations of faba bean accessions.

<table>
<thead>
<tr>
<th>Source</th>
<th>df</th>
<th>SS</th>
<th>MS</th>
<th>Est var</th>
<th>%</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Among Pops</td>
<td>1</td>
<td>40.626</td>
<td>40.626</td>
<td>2.905**</td>
<td>17%</td>
<td>0.001</td>
</tr>
<tr>
<td>Within Pops</td>
<td>17</td>
<td>233.216</td>
<td>13.719</td>
<td>13.719**</td>
<td>83%</td>
<td>0.001</td>
</tr>
<tr>
<td>Total</td>
<td>18</td>
<td>273.842</td>
<td>16.623</td>
<td></td>
<td>100%</td>
<td></td>
</tr>
</tbody>
</table>

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**Figure 2.** PCA plot of the first and second components among 19 accessions based on genetic variation in 25 SSR marker. Different clusters are denoted by different shapes. Accessions symbols are defined under Figure 1.
depending on allele frequencies and not on provenance or taxonomy; we used the method, STRUCTURE analysis. Analysis of population structure was carried out to detect the number of populations that may be created from 19 accessions using 25 SSR markers. In this analysis, the real K value with the highest value of K for the 19 individuals was K= 6 followed by peaks at K= 2 (Figure 3). By the method of Evanno et al (2005), all the analyzed accessions were split into four groups or sub-populations (Figure 4). Here, four sub-populations with the strong mixing of the accessions were acquired as shown in Figure 4. Each individual was characterized by a single color line, and each cluster was denoted by a color. The greater area of a color that an individual expected, the greater the opportunity that the individual belonged to the matching group. Groups 1 (green line) and 2 (yellow line) included all local accessions, while groups 3 (violet line) and 4 (blue line) consisted of ICARDA accessions. Accessions in the admixture class may have partial ancestry in more than one background. They possibly had an intricate history involving intercrossing or

Figure 3. Determining the optimal value of K by the (ΔK) procedure described by Evanno et al. (2005).

Figure 4. Nineteen accessions of faba bean clustered into different sub-populations by STRUCTURE software. Accessions are coordinated as per estimated membership coefficients (q) in K= 6 clusters. Accessions symbols are defined under Figure 1.
feasibly resulting from the gene flow between taxa. Relative low percentages of pure accessions in both clusters indicate high admixture and natural interbreeding of accessions. Cross-pollination of faba bean is expedited by insect pollinators like honeybees, bumblebees and diverse solitary bees. This part of the result demonstrated a lofty gene flow between local and ICARDA groups. Since the places of a collection of samples are linked to each other, it is possibly a reason for having a blend between two populations.

CONCLUSIONS

SSR marker methods have provided beneficial information in terms of detection of the level of polymorphism and characterization in faba bean accessions. In this work, there was manifest variation between the local and ICARDA accessions, as evident in the dendrogram and PCA plot. Furthermore, we detected the level of variation in genetic structure present in the accessions distributed across different regions of Iraq. The results of the current study can be supportive in scheduling the exploitation patterns and management of faba bean existing in north Iraq. Moreover, the results can be useful to the breeders to select effectively the parents leading to progenies with high differentiation among them. It similarly allows the finest arrangement of faba bean gene pool management and more effective sampling of the obtainable accessions resources. Hence, further investigations in these accessions can possibly add novel alleles to biotic or abiotic stress tolerances, which are important for faba bean breeding.

ACKNOWLEDGEMENTS

The authors are grateful to the Faculty of Agricultural Sciences for technical support of this work.

REFERENCES

Diversity and Population Structure Analysis


تجزیه تحلیل تنوع و ساختار جمعیتی لوبیای فابا (Vicia faba) (با استفاده از نشانگر SSR) های ن. ا. طاهر، د. ا. عمر، د. لطیف، د. 1. احمد، س. ه. صالح، و ل. هیوا خال

چکیده

آگاهی و درک مفهوم تنوع زننیکی نمونه‌های لوبیای فابا (Vicia faba) برای اعمال درجه‌بندی در مصرف و حفاظت آنها مهم است. هدف این پژوهش تخمین تنوع زننیکی و ساختار جمعیتی لوبیای فابا در عراق با استفاده از نشانگر‌های SSR برای کاربرد در تحقیقات تولید و تولید ارقام بود. به منظور ارزیابی تغییرات زننیکی و ساختار جمعیتی در میان نمونه‌های لوبیای فابا، 25 مکان زننی ریزمادری اعمال شد. تجزیه شاخصی های تنوع در مجموعه لوبیای فابا های بررسی شده در آن مطالعه نشان داد که بر ماهوره ها برای مشخص کردن زننیکی ها مفید و آموزنده بودند. در کل، 27٪ پلیمورفیکی به طور ماینگین در معرض 2/88 مکان زننی قرار داده شد و 3 آل کمک‌کننده و فرد تشخیص داده شد. ماینگین resolwing power تجزیه زننیکی، شاخص نشانگر، به ترتیب برای برد 1/5120 به بهترین مولفه های اصلی (PCA)، 19 نمونه لیت شده را به 5 خوشه مناسب دسته بندی کرد که هر خوشه زیر- گروه به دنبالی منفیتی داشت. همان گونه که طبق تجزیه واریانس ملکولی (AMOVA) بر نمایه ریزی شده بود، تغییر زننیکی در سطح بالا در درون یک جامعه یا گروه (86/7) به طور معادل دیپتری بیشتر از تغییر این گروه ها یا جامعه ها (7/17) بود. مدل خوشه به دنبال بر مبنای تجزیه نرم افزار STRUCTURE بین گروه ها یا جامعه ها (7/17) بود. مدل خوشه به دنبال بر مبنای تجزیه نرم افزار چهار گروه به راه که از نظر زننیکی پراکندگی بودند. شناسایی کرده‌این یافته‌ها در برنامه‌های بهبودی و تهیه‌ها داده استفاده مورد استفاده دادند.