Biocontrol Potential of Root Endophytic Fungi and *Trichoderma* Species Against Fusarium Wilt of Lentil Under *In vitro* and Greenhouse Conditions

H. Kari Dolatabadi¹, E. Mohammadi Goltapeh¹ *, N. Mohammadi¹, M. Rabiey¹, N. Rohani², and A. Varma³

**ABSTRACT**

Lentil wilt, caused by *Fusarium oxysporum* f. sp. *lentis* is one of the main limiting factors to successful lentil cultivation. Effect of four antagonistic fungi, namely: *Piriformospora indica*, *Sebacina vermifera*, *Trichoderma viride* and *Trichoderma harzianum* as well as combinations among them were evaluated against Fusarium wilt of lentil. In *in vitro* experiments, the effect of the antagonistic fungi against the pathogen were evaluated in dual cultures of volatile metabolite and colonization. Results revealed that antagonistic fungi could produce a proper zone of inhibition. In pot culture experiments, two isolates of pathogen were inoculated within three time schedules, namely: 10 days prior to sowing, concomitant with sowing and 10 days past sowing. Seventeen treatments, including two controls and fifteen combinations of antagonistic fungi were employed to inoculate seeds concordant with sowing. Plant height, plant dry weight, root length, root dry weight along with disease severity were assessed at the flowering stage. Maximum plant height and minimum disease severity with regard to the two isolates of pathogen were observed in pots treated with *S. vermifera*+*T. harzianum.*

**Keywords:** Biological control, *Fusarium oxysporum* f. sp. *Lentis*, *Piriformospora indica*, *Sebacina vermifera*, *Trichoderma harzianum*, *Trichoderma viride*.

**INTRODUCTION**

Lentil (*Lens culinaris* Medic.) is one of the oldest known high protein containing food legumes. Lentil seeds are rich in protein, the mean level being at about 28.5% (Stoilova and Pereira, 1999). Among the biotic factors, diseases are serious threats to lentil production in many parts of the world. Lentil suffers from a number of diseases that are caused by fungi, bacteria, viruses, nematodes and plant parasites (Khare *et al.*, 1979). Such diseases as *Ascochyta* blight and Lentil wilt play major roles in reducing lentil yield (Hamdi and Hassanein, 1996). Lentil wilt, caused by *Fusarium oxysporum* f. sp. *lentis* (FOL) is one of the main limiting factors to successful lentil cultivation. This pathogen is responsible for severe grain losses (Stoilova and Chavdarov, 2006; Bayaa *et al.*, 1986). The disease appears in either the early stages of crop growth (seedling) or during the reproductive stage (adult stage) (Stoilova and Chavdarov, 2006; Khare, 1981). A majority of terrestrial plants live in mycorrhizal symbiosis with fungi.
MATERIALS AND METHODS

F. oxysporum f. sp. lentis and Trichoderma spp. Cultures

Two isolates of FOL were obtained from wilted lentil plants collected from lentil growing areas in Mashhad (F1) and Ilam (F2). The fungus was isolated from the roots of the wilted plants. Cultures were maintained on Potato Dextrose Agar (PDA) medium and stored at 4°C for later use, and while the Trichoderma species as T. harzianum and T. viride being employed.

Fungal Solid Culture of P. indica and S. vermifera

Endophytic fungi (P. indica, S. vermifera) and Trichoderma species were collected from Plant Pathology Type Culture Collection (Mohammadi Goltapeh), Tarbiat Modares University, Tehran, Iran. P. indica and S. vermifera were placed on Kafer’s medium (Kafer, 1977). The plates were incubated at 25°C for a week.

In vitro Evaluation of Antagonistic Fungi against F. oxysporum f. sp. Lentis, Dual Culture Assay

Piriformospora indica, S. vermifera, T. harzianum and T. viride were evaluated against two isolates of FOL through dual culture technique as described by Morton and Stroube (1955) and as well by Kucuk and Kivanc (2003). Petri dishes (90 mm) containing PDA were inoculated with a 5 mm plug of 7 day old pure culture of antagonistic fungi and pathogen. One mycelial disc of each fungus was placed at opposite sides on PDA plates and incubated at 25±1°C with the radial growth of FOL being measured 2, 4 and 6 days after incubation. Control Petri dishes were inoculated with FOL (F1 and F2) and a sterile agar plug. A number of three replications were assigned to each treatment. Percent inhibition of FOL radial growth was determined. For each interaction, a clean and sterile glass microscope slide was placed in the middle of plates and sterilized. A thin layer of autoclaved melted potato dextrose agar was then spread over the slide. Five mm discs of seven days old culture cut from the edge of each pathogen and antagonistic fungi were placed at opposite sides on PDA plates and incubated at 25±1°C in an
Biocontrol Potential of Root Endophytic Fungi __________________________________

incubator. After one week past, the slides were observed microscopically for hyphal interactions.

**Effect of Volatile Metabolites**

The effect of volatile metabolites produced by the antagonistic microorganisms on FOL mycelial growth was determined by the method described by Dennis and Webster (1971) and as well by Goyal et al. (1994). The antagonistic fungi were centrally inoculated by placing 5 mm diameter mycelia disc taken from 3 day old culture on the PDA plate and then incubated at 25±1°C for 2 days. The top of each Petri dish was replaced with bottom of the PDA plate centrally inoculated with the pathogen. Two plates were sealed together with paraffin tape and further incubated at 25°C. As for control, instead of *Trichoderma* spp. a 5 mm diameter of sterile PDA medium was used, being placed in plate. Three replications were employed for each treatment. Colony diameter of the pathogen was measured at 4 and 6 days after incubation and the inhibition of mycelial growth determined. The percent growth inhibition in all the above experiments was calculated by using the following equation (Vincent, 1947):

\[ I = \frac{(C-T)}{C} \times 100 \]

where, \( I \) = Percent growth inhibition; \( C \) = Colony growth rate in checked plates, \( T \) = Colony growth rate in each treatment.

**Comparison of Antagonistic Fungi in Colonization of FOL Mycelium**

Colonization of antagonistic effect of the fungi on FOL mycelium was carried out by some modified method described by Mohammadi and Danesh (2006), the study being carried out in two phases as follows:

In the first phase, 5 mm discs of FOL were placed on PDA plates and incubated at 25±1°C for 4 days before 5 mm discs of *P. indica*, *S. vermifera*, *T. harzianum* and *T. viride* mycelia being placed of the center of the Petri dish. In the second phase, 5 mm discs of FOL were placed on PDA plates and incubated at 25±1°C for 12 days before 5 mm discs of *P. indica*, *S. vermifera*, *T. harzianum* and *T. viride* mycelia being placed of the center of the Petri dish. Each treatment was replicated thrice.

**Pot Culture Experiments**

**Preparation of Inoculums**

Liquid culture of *P. indica* and *S. vermifera* were prepared in 500 ml flasks containing 200 ml of autoclaved KM liquid medium through inoculation with four mycelial discs from 10 day old agar culture of *P. indica* and *S. vermifera*. Flasks were kept on a shaker (140 rpm) at 25±1°C for 15 days until dense mycelial suspensions were generated. Then, the broth cultures were stored at 4°C for later pot culture experiments.

To produce inoculums for pathogen and *Trichoderma* species, 90 g sand, 10 g straw, and 20 ml distilled water for FOL and 100 g wheat seeds and 40 ml distilled water for *Trichoderma* species were separately added to bottle glasses and sterilized at 121°C for 120 minutes. Bottle glasses were separately inoculated with five 10 mm discs of pathogen and *Trichoderma* species cut from the margin of a 4 day old culture. The flasks were placed in incubator at 23 to 26°C until completely colonized.

**Put Cultures**

Seeds of *Lens culinaris* were obtained from the Dryland Agricultural Research Institute, Maragheh, Iran. Pot culture experiments were conducted in greenhouse during 2009 while; using a completely randomized design with a factorial combination of three time schedules, two isolate pathogens, and fifteen combinations of antagonistic fungi as well as two controls (non-pathogen vs. pathogen) in 3 replicates to evaluate the performances of antagonistic fungi as biocontrol agents against wilt. Seeds of *Lens culinaris* were surface-
sterilized by being soaked in 1% sodium hypochlorite for 1 minute, then bring rinsed three times in sterile distilled water and placed in sterilized perlite for germination. After 3-4 days past when the plumule and radicle appeared, three germinated seeds were transferred to pots and grown in a 2:1:1 mixture of sand: peat: perlite in greenhouse at 24:18°C day⁻¹: night cycle, with a photoperiodicity of 16:8 light/dark. Soil had been disinfected with a 7% formaldehyde solution. Pots were inoculated with pathogen in three time schedules (time 1; 10 days prior to sowing, time 2; concomitant with sowing, and time 3; 10 days past sowing). Antagonistic fungi were inoculated concordant with sowing.

Treatments:

T1= Control (Non pathogen)
T2= Control (Pathogen)
T3= Pathogen+P. indica
T4= Pathogen+S. vermifera
T5= Pathogen+T. viride
T6= Pathogen+T. harzianum
T7= Pathogen+P. indica+S. vermifera
T8= Pathogen+P. indica+T. viride
T9= Pathogen+P. indica+T. harzianum
T10= Pathogen+S. vermifera+T. viride
T11= Pathogen+S. vermifera+T. harzianum
T12= Pathogen+T. viride+T. harzianum
T13= Pathogen+P. indica+S. vermifera+T. viride
T14= Pathogen+P. indica+S. vermifera+T. harzianum
T15= Pathogen+P. indica+T. viride+T. harzianum
T16= Pathogen+S. vermifera+T. viride+T. harzianum
T17= Pathogen+P. indica+S. vermifera+T. viride+T. harzianum

Pots were inoculated at 30 g kg⁻¹ with mass multiplied pathogen and 10 g kg⁻¹ with mass multiplied Trichoderma species (10⁶ CFU g⁻¹). For inoculation with P. indica or S. vermifera, 1 g of crushed mycelia was added to radicle seeds. The controls were kept devoid of inoculation of antagonistic fungi.

Plant height, root length, as well as plant and root dry weights were assessed at flowering stage after nine weeks past. The disease severity was recorded starting from the 5th day after inoculation and continued for nine weeks using a 1-9 scale (Bayaa et al., 1995). 1: no symptoms, 3: yellowing of the basal leaves only, 5: yellowing of 50% of the foliage, 7: complete yellowing of the foliage, flaccidity of the top leaves along with partial drying, 9: whole plant or part of the plant wilted and/or dried.

Statistical analysis

The collected data were statistically analyzed using software SAS 6.12. Data were subjected to analyses of variance and treatment means were compared through Tukey’s test (P= 0.05)

RESULTS

In vitro Experiment

Dual culture studies on the antagonistic fungi against F1 isolate of FOL indicated that at 2, 4 and 6 days after incubation, T. harzianum caused maximum growth inhibition on mycelium FOL, then T. viride had a marked significant inhibitory effect on the growth of the pathogens (Figure 1-a). The growth inhibition of F2 isolate of FOL by antagonistic fungi after 2 and 4 days past incubation revealed that T. viride and T. harzianum resulted in maximum growth inhibition (Figure 1-b). P. indica and S. vermifera were ineffective in reducing radial growth of FOL after 2 and 4 days of incubation. However, P. indica and S. vermifera were effective in radial growth inhibition of FOL after 6 days of incubation (Figures 1a-b).

Trichoderma species differentially limited the colonical growth of the pathogen, overgrew the pathogen colony and produced yellow pigment. Microscopically, the hyphal interaction indicated that antagonistic hyphae coiled around the hyphae of pathogen, denatured the mycelia and finally killed them. P. indica, S. vermifera and Trichoderma species either formed hook or
bunch-like structures around the hyphae of FOL before penetration, or sometimes entered them directly (Figure 2).

The observances on days 4 and 6 after incubation, indicated that the antifungal activity of volatile metabolite was varied. Percentage reduction in mycelial growth of F1 isolate of FOL with T. viride was greater than that of the others (Figure 3a). The observations on days 4 and 6 after incubation revealed that volatile metabolites produced by T. harzianum caused maximum growth inhibition of F2 isolate of FOL, followed by T. viride (Figure 3-b). Antagonists P. indica and S. vermifera were ineffective in reducing radial growth of FOL, on days 4 and 6 past incubation (Figures 3a-b).

Effect of colonization in the first phase revealed that T. harzianum rendered the highest colonization of FOL (isolates F1 and F2) myceli within 4 days, T. viride, S. vermifera and P. indica presented...
colonization rate of 5, 9 and 10 days, respectively. In the second phase *T. harzianum, T. viride, S. vermifera* and *P. indica* had colonization rates of 5, 5, 9 and 10 days, respectively.

**Pot Culture Experiments**

To assess the influence of *P. indica, S. vermifera* and *Trichoderma* species on plant morphology, several parameters including plant height, root length, dry weight of plant and root, as well as disease severity were scrutinized. Results revealed the significant effect of the treatments (P= 0.05) (Figure 4).

The effect of the three time variables of inoculated pathogen along with the treatments (without considering the controls T1 and T2) revealed that the tallest plant was observed for time 3 and soil treated with *S. vermifera+ T. harzianum* (T11) (Table 1). As for time 1 treatment, maximum shoot length with respect to F1 isolate was observed in pots inoculated with *S. vermifera+ T. harzianum* (T11) and the tallest plant with respect to F2 isolate was observed in pots inoculated with *P. indica+S. vermifera+ T. harzianum* (T14).
Table 1: Effect of antagonistic treatments on plant height of *Lens culinaris* in pots inoculated with two isolateS (F1 and F2) of *Fusarium oxysporum f.sp. lentis* in three times.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Time 1</th>
<th>Time 2</th>
<th>Time 3</th>
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<tbody>
<tr>
<td></td>
<td>F1</td>
<td>F2</td>
<td>F1</td>
</tr>
</tbody>
</table>
| T1= Control (Non-pathogen); T2= Control (Pathogen); T3= Pathogen+*P. indica*; T4= Pathogen+*S. vermifera*; T5= Pathogen+*T. viride*; T6= Pathogen+*T. harzianum*; T7= Pathogen+*P. indica*+*S. vermifera*; T8= Pathogen+*P. indica*+*T. viride*; T9= Pathogen+*P. indica*+*T. harzianum*; T10= Pathogen+*S. vermifera*+*T. harzianum*; T11= Pathogen+*S. vermifera*+*T. harzianum*; T12= Pathogen+*T. viride*+*T. harzianum*; T13= Pathogen+*P. indica*+*S. vermifera*+*T. viride*; T14= Pathogen+*P. indica*+*S. vermifera*+*T. harzianum*; T15= Pathogen+*P. indica*+*T. viride*+*T. harzianum*; T16= Pathogen+*S. vermifera*+*T. viride*+*T. harzianum*; T17= Pathogen+*P. indica*+*S. vermifera*+*T. viride*+*T. harzianum*; T1= Control (Non-pathogen); T2= Control (Pathogen); T3= Pathogen+*P. indica*; T4= Pathogen+*S. vermifera*; T5= Pathogen+*T. viride*; T6= Pathogen+*T. harzianum*; T7= Pathogen+*P. indica*+*S. vermifera*; T8= Pathogen+*P. indica*+*T. viride*; T9= Pathogen+*P. indica*+*T. harzianum*; T10= Pathogen+*S. vermifera*+*T. harzianum*; T11= Pathogen+*S. vermifera*+*T. harzianum*; T12= Pathogen+*T. viride*+*T. harzianum*; T13= Pathogen+*P. indica*+*S. vermifera*+*T. viride*; T14= Pathogen+*P. indica*+*S. vermifera*+*T. harzianum*; T15= Pathogen+*P. indica*+*T. viride*+*T. harzianum*; T16= Pathogen+*S. vermifera*+*T. viride*+*T. harzianum*; T17= Pathogen+*P. indica*+*S. vermifera*+*T. viride*+*T. harzianum*; T1= Control (Non-pathogen); T2= Control (Pathogen); T3= Pathogen+*P. indica*; T4= Pathogen+*S. vermifera*; T5= Pathogen+*T. viride*; T6= Pathogen+*T. harzianum*; T7= Pathogen+*P. indica*+*S. vermifera*; T8= Pathogen+*P. indica*+*T. viride*; T9= Pathogen+*P. indica*+*T. harzianum*; T10= Pathogen+*S. vermifera*+*T. harzianum*; T11= Pathogen+*S. vermifera*+*T. harzianum*; T12= Pathogen+*T. viride*+*T. harzianum*; T13= Pathogen+*P. indica*+*S. vermifera*+*T. viride*; T14= Pathogen+*P. indica*+*S. vermifera*+*T. harzianum*; T15= Pathogen+*P. indica*+*T. viride*+*T. harzianum*; T16= Pathogen+*S. vermifera*+*T. viride*+*T. harzianum*; T17= Pathogen+*P. indica*+*S. vermifera*+*T. viride*+*T. harzianum*; Values in the same column, followed by the same letters are not significantly different (Tukey’s test, P<0.05).

For time 2, shoot length with respect to F1 isolate was highest with *T. viride*+*T. harzianum* (T12), and as regards F2 isolate it was highest for *P. indica*+*S. vermifera*+*T. harzianum* (T14) (Table 1).

Results revealed that the longest root with respect to F1 isolate was observed in the pots inoculated with *P. indica*+*T. viride*+*T. harzianum* (T15), followed by *P. indica*+*S. vermifera* (T7) and *S. vermifera*+*T. harzianum* (T11) within time 3 treatment (Table 2). In time 2, the influence of treatment on root length with respect to FOL isolates revealed that in pots inoculated with F1 isolate, maximum root length was observed in the pots inoculated with *P. indica*+*S. vermifera* (T7), and in pots inoculated with F2 isolate, maximum root length was observed for *S. vermifera*+*T. viride* (T10). (Table 2) treatment. In time 1 treatment, minimum root length was observed for *P. indica*+*S. vermifera*+*T. viride* (T13) in pots inoculated with F1 isolate (Table 2). The effect of three time variables of inoculated pathogen and treatments (without considering the controls T1 and T2) revealed that the highest plant dry weight with respect to F1 isolate was observed for time 3 and soil treated with *S. vermifera*+*T. harzianum* (T11) (Table 2). In time 1, in pot cultures inoculated with F1 isolate, maximum plant dry weight was observed for the pots inoculated with *S. vermifera*+*T. harzianum* (T11), with a minimum plant dry weight being observed for the pots inoculated with *P. indica*+*S. vermifera*+*T. viride* (T13) (Table 2). As for time 2, influence of treatment on plant dry weight...
Table 2: Effect of antagonistic treatments on root length and plant dry weight of *Lens culinaris* in pots inoculated with two isolates (F1 and F2) of *Fusarium oxysporum f.sp. lentis* in three times.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Mean root length (cm)</th>
<th>Mean plant dry weight (g)</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Time 1</td>
<td>Time 2</td>
</tr>
<tr>
<td>T1= Control (Non-pathogen)</td>
<td>F1</td>
<td>F2</td>
</tr>
<tr>
<td>T2= Control (Pathogen)</td>
<td>14.67a</td>
<td>14.33a</td>
</tr>
<tr>
<td>T3= Pathogen+<em>P. indica</em></td>
<td>6.33cd</td>
<td>4.67bc</td>
</tr>
<tr>
<td>T4= Pathogen+<em>S. vermifera</em></td>
<td>10.33abcd</td>
<td>13.67a</td>
</tr>
<tr>
<td>T5= Pathogen+<em>T. viride</em></td>
<td>16.67a</td>
<td>8.00abc</td>
</tr>
<tr>
<td>T6= Pathogen+<em>T. harzianum</em></td>
<td>8.00bcd</td>
<td>9.33abc</td>
</tr>
<tr>
<td>T7= Pathogen+<em>P. indica</em></td>
<td>14.67ab</td>
<td>10.00abc</td>
</tr>
<tr>
<td>T8= Pathogen+<em>S. vermifera</em></td>
<td>7.67bcd</td>
<td>12.33abc</td>
</tr>
<tr>
<td>T9= Pathogen+<em>T. viride</em></td>
<td>7.67bcd</td>
<td>8.67abc</td>
</tr>
<tr>
<td>T10= Pathogen+<em>T. harzianum</em></td>
<td>5.00d</td>
<td>12.67ab</td>
</tr>
<tr>
<td>T11= Pathogen+<em>P. indica</em></td>
<td>12.67abcd</td>
<td>12.00abc</td>
</tr>
<tr>
<td>T12= Pathogen+<em>S. vermifera</em></td>
<td>7.67bcd</td>
<td>8.00abc</td>
</tr>
<tr>
<td>T13= Pathogen+<em>T. viride</em></td>
<td>8.33bcd</td>
<td>11.67abc</td>
</tr>
<tr>
<td>T14= Pathogen+<em>T. harzianum</em></td>
<td>13.67abc</td>
<td>13.00ab</td>
</tr>
<tr>
<td>T15= Pathogen+<em>P. indica</em></td>
<td>14.00abc</td>
<td>14.00ab</td>
</tr>
<tr>
<td>T16= Pathogen+<em>S. vermifera</em></td>
<td>14.00abcd</td>
<td>12.00abc</td>
</tr>
<tr>
<td>T17= Pathogen+<em>T. viride</em></td>
<td>11.33abc</td>
<td>11.00abc</td>
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</table>

Values in the same column, followed by the same letters are not significantly different (Tukey’s test, P< 0.05).
weight with respect to FOL isolates revealed that F2 isolate treated with \textit{P. indica}+\textit{S. vermifera}+\textit{T. harzianum} (T14) and \textit{S. vermifera}+\textit{T. harzianum} (T11) rendered maximum plant dry weights (Table 2).

The effect of three time treatments of inoculation of pathogen on root dry weight revealed that the highest root dry weight was observed for time 3, and minimum root dry weight observed for time 1 treatment (Table 3). As for time 1, the effect of 17 treatments (with an omission of T1 and T2) on root dry weight revealed that the highest root dry weight was observed in the pots inoculated with \textit{S. vermifera} (T4) (Table 3).

Minimum disease severity with respect to two isolates of FOL were observed for time 3 treatment and in soil treated with \textit{S. vermifera}+\textit{T. harzianum} (T11) (Table 3). Isolate F1 treated with \textit{P. indica}+\textit{T. harzianum} (T9), \textit{S. vermifera}+\textit{T. harzianum} (T11) and \textit{T. viride}+\textit{T. harzianum} (T12) as well as isolate F2 treated with \textit{T. harzianum} (T6), \textit{S. vermifera}+\textit{T. harzianum} (T11) and \textit{P. indica}+\textit{S. vermifera}+\textit{T. harzianum} (T14) demonstrated a minimum of wilt incidence in the case of time 3 treatment (Table 3).

**DISCUSSION**

Plants are often colonized by many fungi that do not cause any disease symptoms. Many of these endophytic fungi render beneficial effects on plant growth and development. Positive effects may come from providing essential nutrients as a result of their colonization (Harrison, 2005). The members of Basidiomycetes \textit{P. indica}, and \textit{S. vermifera} form mutuality symbiosis with a broad spectrum of terrestrial plants. Their presence causes such beneficial activities as an increase in vegetative biomass, grain yield, local and systemic disease resistance, as well as tolerance to abiotic stresses (Waller et al., 2005). Endophytic colonization of either seed surface or roots, and promotion of growth are desirable outcomes as regards biocontrol activity (Kleifeld and Chet, 1992).

Our studies in dual culture revealed that all antagonistic fungi inhibited mycelial growth of the pathogen. Antagonists \textit{P. indica} and \textit{S. vermifera} were ineffective in reducing radial growth in FOL at times 2 and 4 days after incubation. However, these fungi were able to reduce mycelial growth of the pathogen by day 6 after incubation in dual culture, suggesting that they do not act by producing volatile metabolites within 2 and 4 days after incubation but by days 6 after incubation they were effective in reducing radial growth through other such mechanisms as competition or parasitism.

\textit{Trichoderma} spp. inhibited the growth of FOL through a production of volatile metabolites. By days 4 and 6 after incubation, the percentage reduction in mycelial growth of F1 isolate in the presence of \textit{T. viride} was greater than those of the other antagonists. \textit{Trichoderma} species are known to produce a number of such antibiotics, as trichodermin, viridian and harzianolide (Simon and Sivasithamparam, 1988; Schirrmbock et al., 1994; Dennis and Webester, 1971).

The present work’s studies on colonization revealed that \textit{T. harzianum} had the highest colonization of FOL mycelium. Our results are in agreement with those of Ghaahfarokhi and Goltapeh (2010), they reported that \textit{P. indica}, \textit{S. vermifera} and \textit{Trichoderma} species were the most potent agents in biocontrol of take-all diseases of wheat (\textit{Gaëumannomyces graminis var. tritici}).

In pot culture experiments, \textit{S. vermifera}+\textit{T. harzianum} (T11) induced maximum plant height in lentil plants. The longest roots with respect of F1 isolate come respectively from plants inoculated with \textit{P. indica}+\textit{T. viride}+\textit{T. harzianum} (T15) and \textit{P. indica}+\textit{S. vermifera} (T7) within time 3, followed by soil treated with \textit{S. vermifera}+\textit{T. harzianum} (T11). Dubey et al. (2006) reported that root colonization by Ranchi isolate of \textit{T. viride} induced a maximum growth of roots and shoots in chickpea plants followed by \textit{T. harzianum}. Harman (2000) reported that strains 22 of \textit{T. harzianum} increased root development in maize and in several other
Table 3: Effect of antagonistic treatments on root dry weight and disease severity of *Lens culinaris* in pots inoculated with two isolates (F1 and F2) of *Fusarium oxysporum* f.sp. *lentis* in three times.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Mean root dry weight (g)</th>
<th>Mean disease severity</th>
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<tbody>
<tr>
<td></td>
<td>Time 1</td>
<td>Time 2</td>
</tr>
<tr>
<td>T1</td>
<td>0.25 a</td>
<td>0.25 a</td>
</tr>
<tr>
<td>T2</td>
<td>0.14 bc</td>
<td>0.10 a</td>
</tr>
<tr>
<td>T3</td>
<td>0.18 abc</td>
<td>0.16 a</td>
</tr>
<tr>
<td>T4</td>
<td>0.21 ab</td>
<td>0.27 a</td>
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<tr>
<td>T5</td>
<td>0.16 abc</td>
<td>0.18 a</td>
</tr>
<tr>
<td>T6</td>
<td>0.19 abc</td>
<td>0.18 a</td>
</tr>
<tr>
<td>T7</td>
<td>0.14 bc</td>
<td>0.16 a</td>
</tr>
<tr>
<td>T8</td>
<td>0.17 abc</td>
<td>0.18 a</td>
</tr>
<tr>
<td>T9</td>
<td>0.11 c</td>
<td>0.15 a</td>
</tr>
<tr>
<td>T10</td>
<td>0.10 c</td>
<td>0.19 a</td>
</tr>
<tr>
<td>T11</td>
<td>0.20 abc</td>
<td>0.16 a</td>
</tr>
<tr>
<td>T12</td>
<td>0.12 bc</td>
<td>0.14a</td>
</tr>
<tr>
<td>T13</td>
<td>0.12 bc</td>
<td>0.19a</td>
</tr>
<tr>
<td>T14</td>
<td>0.18abc</td>
<td>0.20a</td>
</tr>
<tr>
<td>T15</td>
<td>0.19 abc</td>
<td>0.13a</td>
</tr>
<tr>
<td>T16</td>
<td>0.13 bc</td>
<td>0.13a</td>
</tr>
<tr>
<td>T17</td>
<td>0.16 abc</td>
<td>0.18 a</td>
</tr>
</tbody>
</table>

T1= Control (non-pathogen); T2= Control (Pathogen); T3= Pathogen+*P. indica*; T4= Pathogen+*S. vermifera*; T5= Pathogen+*T. viride*; T6= Pathogen+*T. harzianum*; T7= Pathogen+*P. indica+S. vermifera*; T8= Pathogen+*P. indica+T. viride*; T9= Pathogen+*P. indica+T. harzianum*; T10= Pathogen+*S. vermifera+T. viride*; T11= Pathogen+S. vermifera+*T. harzianum*; T12= Pathogen+*T. viride+T. harzianum*; T13= Pathogen+*P. indica+S. vermifera+T. viride*; T14= Pathogen+*P. indica+T. harzianum*; T15= Pathogen+*P. indica+T. viride+T. harzianum*; T16= Pathogen+*S. vermifera+T. viride+T. harzianum*; T17= Pathogen+*P. indica+S. vermifera+T. viride+T. harzianum*

Values in the same column, followed by the same letters are not significantly different (Tukey’s test, P< 0.05).
crop plants under either greenhouse or field conditions.

The highest plant dry weight with respect to F1 isolate was observed for time 3 treatment and soil treated with S. vermifera+T. harzianum (T11). Isolate F2 treated with T. harzianum (T6) exhibited a maximum plant dry weight for time 3 treatment. The effect of 17 treatments on root dry weight revealed that the highest root dry weight with respect to F2 isolate was observed in the pots inoculated with T. harzianum (T6) and for time 3. Serfling et al. (2007) assessed the potential of P. indica to protect wheat plantlets from the root pathogen F. culmorum, the stem base pathogen Pseudocercospora herpotrichoides, and the leaf pathogen Blumeria graminis f. sp. Triticci reporting that the fresh weights of roots, shoots, and of the entire plants were significantly increased at all time points in plants grown on sand or soil as compared with the controls.

The least disease severity with respect to two isolates of FOL was observed in time 3 treatment and soil treated with S. vermifera+T. harzianum (T11). Dubey et al. (2006) reported that Trichoderma spp. significantly reduced the wilt incidence in chickpea. Poddar et al. (2004) reported that rhizosphere isolate of T. harzianum decreased wilt incidence in chickpea. El-Hassan and Gowen (2006) tested three formulations of the biocontrol agent Bacillus subtilis and found that B. subtilis significantly decreased disease severity of Fusarium oxysporum f.sp. lentis. Serfling et al. (2007) reported that Pseudocercospora herpotrichoides disease severity was significantly reduced in plants colonized by P. indica in field experiment. In the present study T. harzianum+S. vermifera treatment (T11) was found as the most effective against FOL. Fakhro et al. (2010) reported that limitation of disease severity caused by Verticillium dahliae by more than 30% was observed on tomato plants colonised by the endophyte P. indica.

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REFERENCES

419


پاتنسل بیوکنترل فارج های اندوفیتیش رشد و گونه های تریکودرما روی پسرگی
فوزاریومی عدس در شرایط درون شیشه ای و گلخانه ای

ج. کاری دولت‌آبادی، ا. محمدی گل تیه، ن. محمدی، م. ریبیعی، ن. روحانی و 1. وارما

چکیده

پسرگی عدس با عامل *Fusarium oxysporum* f.sp. *lentis* کریکه‌گی کشت عدس می‌باشد. تأثیر چهار فارج آنتاگونیست شامل *Piriformospora indica* و *Trichoderma harzianum* و *Trichoderma viride Sebacina vermifera* هنگامی از آن‌ها روی پسرگی فوزاریومی عدس مورد ارزیابی قرار گرفت. در آزمایش‌های درون شبیه

ای، تأثیر فارج های آنتاگونیست روی پیمارگ در کشت متقالی، مواد فارار و کلینیک‌سیون مورد ارزیابی

419
قرار گرفت. نتایج نشان داد که فارم های آنتاگونیست یک ناحیه بازداری خوب می توانند ایجاد کنند.
در آزمایش های گلدانی، ۲ ایزوله بیمار گیر در ۳ زمان، ۱۰ روز قبل از کاشت بذر، همزمان با کاشت بذر و ۱۰ روز بعد از کاشت بذر مایه زنی شد. ۱۷ تیمار که شامل ۲ شاهد و ۱۵ ترکیب مختلف فارم های آنتاگونیست بود، همزمان با کاشت بذر به خاک گلدان ها مایه زنی گردیدند. ارتفاع گیاه و وزن خشک هوا به طول رشد، وزن خشک ریشه و شدت بیماری در مرحله گلدهی مورد ارزیابی قرار گرفت.
بهترین ارتفاع و کمترین شدت بیماری با در نظر گرفتن ۲ ایزوله بیمار گیر در گلدان های تیمار شده با S. vermifera + T. harzianum مشاهده شد.